

Colorectal Cancer Molecular Diagnostics

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Disclosures

Dr. Bronner has no disclosures pertaining
To this presentation.

Molecular CRC Testing

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- MSI, MMR IHC, Sporadic, Lynch
- KRAS
- BRAF
- Other genes and mutations in clinical trials

MSI Testing

- MSI-H Sporadic: 15% CRC
- MSI-H Lynch: 2-3% CRC
- MSS: 92% CRC

Lynch Syndrome Cancers

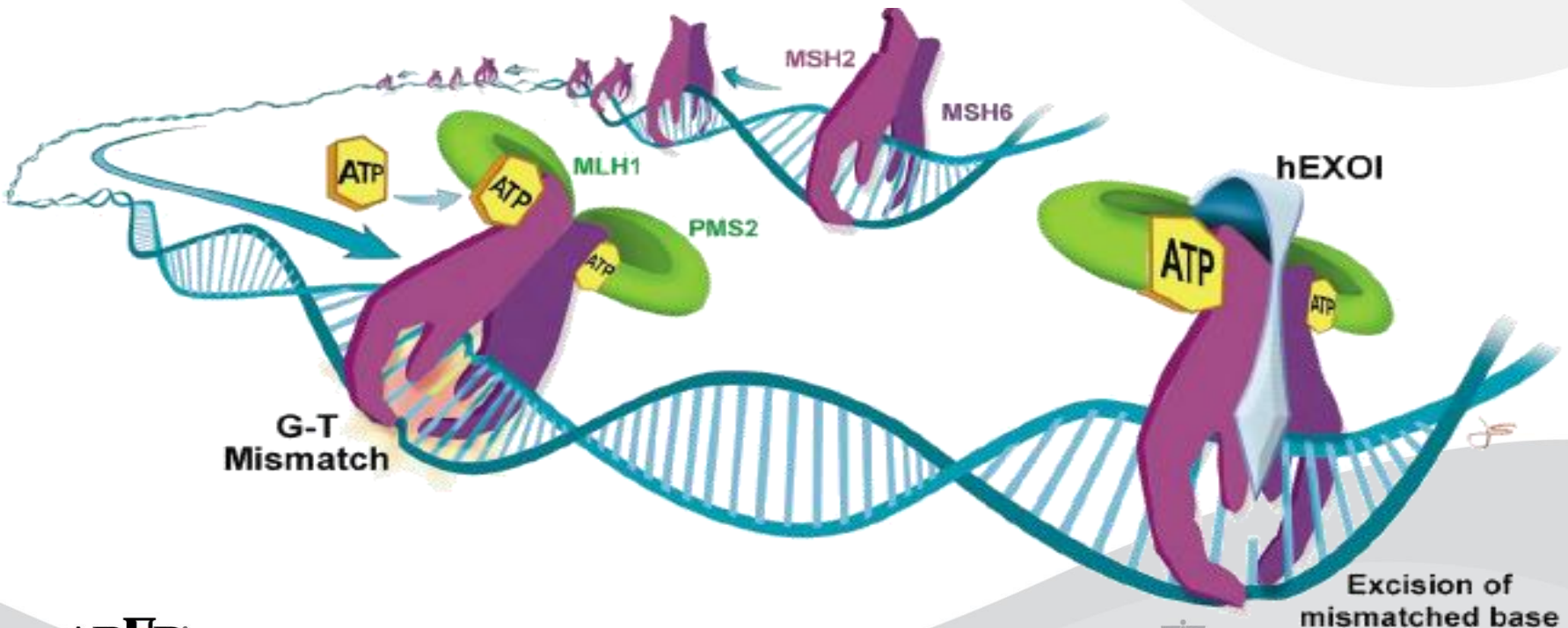
- Colorectal CA 80% & Endometrial CA 50%
- Other CA's: panc/bil, gastric, small bowel, sebaceous skin (Muir Torre), ovarian, GU, GBM (Turcot's)
- Screening: Age 25 or 10 yrs < youngest in family
Annual colonoscopy & endometrial bx, periodic EGD, EUS of pancreas, pelvic exam, brain scans, urine cytology

**HUGE & LIFELONG IMPACT ON
LYNCH PATIENTS: DX IS CRITICAL**

Microsatellite instability (MSI)

Microsatellites: Short, stably inherited repetitive DNA sequences prone to error during replication

Normally repaired by MMR proteins: proofreading complex for the DNA polymerase



Microsatellite instability (MSI)

- Lynch Syndrome: GERMLINE autosomal dominant mutation in a MMR gene
- MMR genes: **MLH-1, MSH-2, MSH-6, PMS-2**
- MSI: microsatellites of altered lengths accumulate throughout the genome due MMR deficiency
- Lynch phenotype not so obvious (nonpolyposis)
- Family history not always obvious or available
- MMR deficiency permits molecular diagnosis

MSI-High Colon Cancer

- **Sporadic:** Tumor limited, nearly all MLH1 methylation
 - Rare somatic MMR mutations described
- **Lynch:** Germline mutations
 - MLH1 60%, MSH2 35%, PMS2, MSH6, EPCAM, POL 5%
- Lynch & Sporadic Pathology: identical

Reasons to Diagnose MSI-H CRC

- Hereditary and syndromic components of Lynch
- Improved survival for sporadic CRC in randomized/stage-matched trials
- Chemo- and immuno-therapy selection
 - 5FU out
 - Immunotherapy in

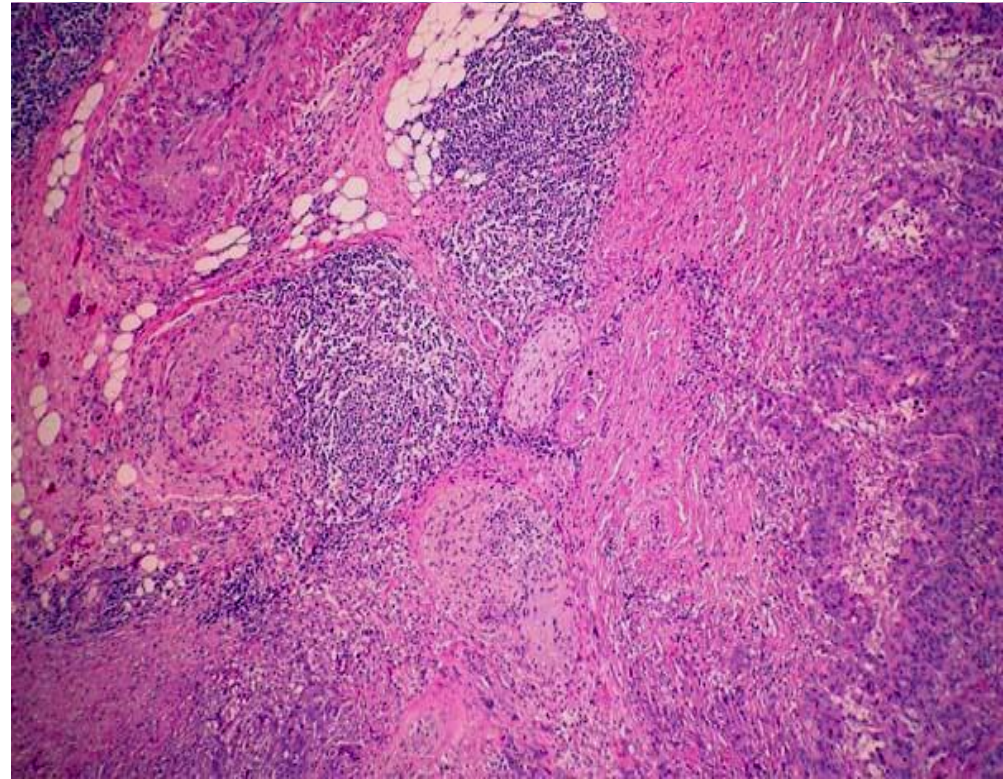
Andre T, et al. N Engl J Med 383:2207;2020

Zaanan A, et al. Clin Cancer Res 17:7470;2011

Ribic CM, et al. NEJM 349:247;2003

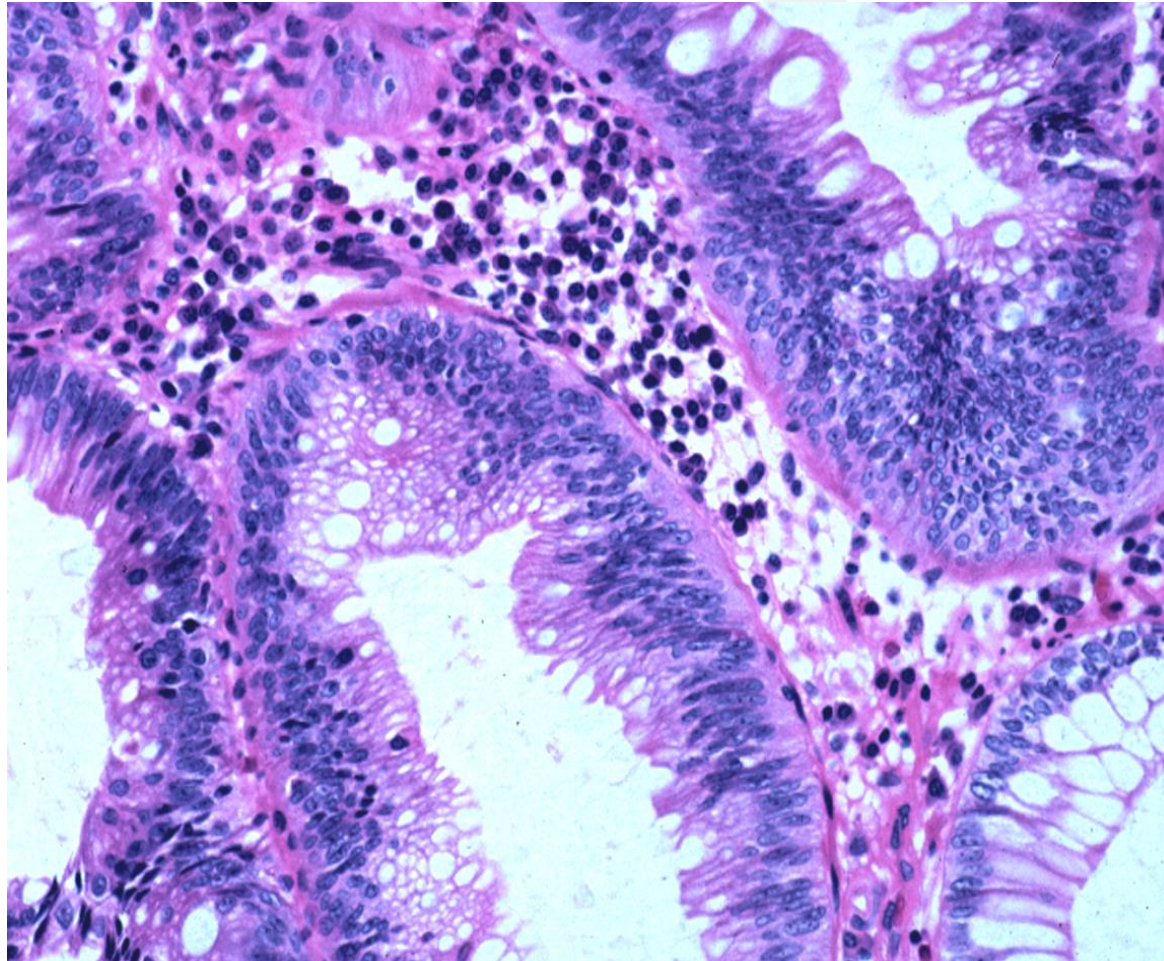
MSI-H CRC: Clinicopathologic Features

- Right-sided location
- Age < 50 years (Lynch)
- Poor differentiation
- Absence of dirty necrosis
- >2 tumor infiltrating lymphs/hpf
- Mucinous change
- Crohn's-like lymphoid reaction



Greenson JK, et al. *Am J Surg Pathol* 27:563-570, 2003.

Duodenal or Gastric Adenoma



Consider FAP and Lynch Syndrome

How do we work up Lynch syndrome?

- Determine mismatch repair deficiency
 - PCR for microsatellite instability
 - IHC for mismatch repair proteins
- Determine mismatch repair deficient tumor type
 - Sporadic: no germline testing
 - Possibly inherited: germline testing needed

Lynch Testing

Tumor screening assays (90% sens)

Detect affected **patients with tumor**

– MSI by PCR: paraffin works well

– MMR Immunohistochemistry:

- MLH-1, MSH-2, MSH-6, PMS2 (work well with correct positive controls)

Blood germline mutation analysis

Detects **family members without tumor**

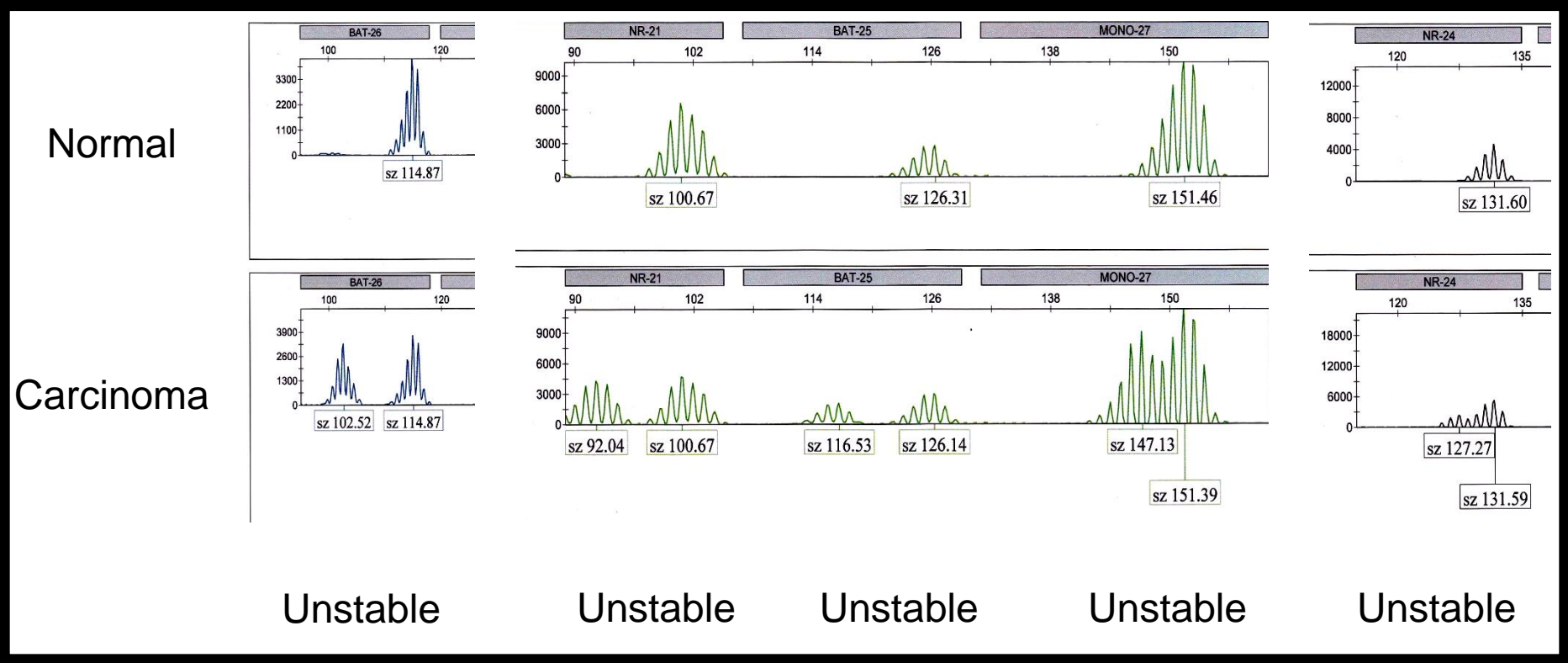
MSI Requirements

- *Tumor*
- *Normal DNA*
 - Non-tumor paraffin tissue
 - Blood
 - Buccal swab

Mononucleotide repeat panel

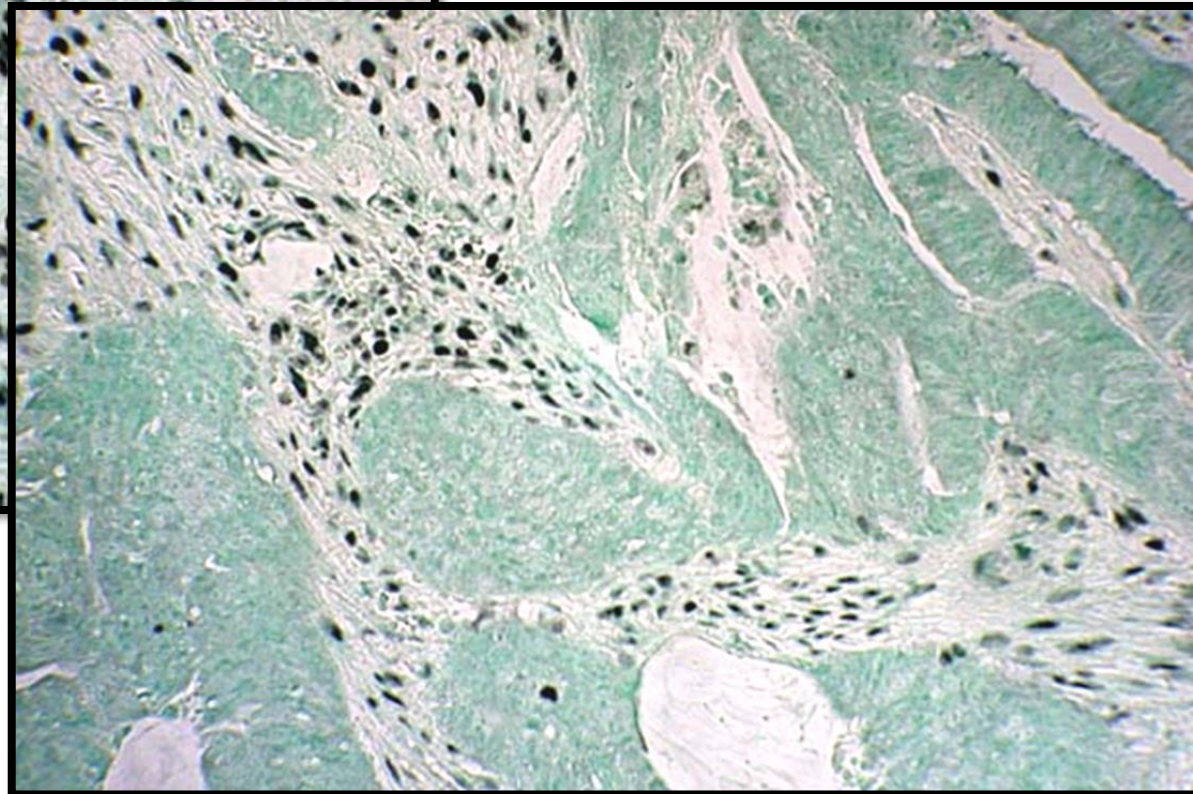
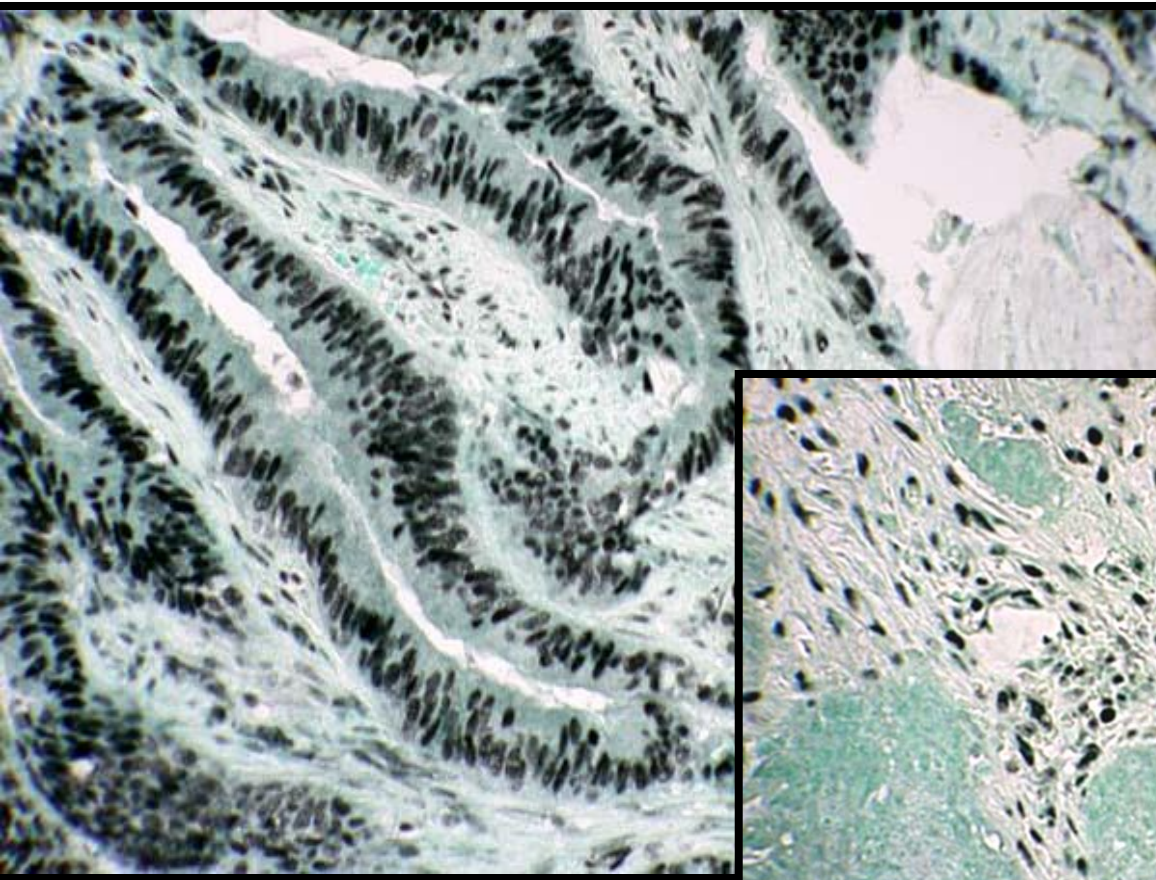
- Mononucleotide repeats are probably more sensitive and specific for MMR deficiency
- 5 mononucleotide repeat panel
 - MSI high: 2 or more unstable, although typically all (or almost all) repeats are unstable
 - Since instability in even one mononucleotide repeat may indicate MMR deficiency, instability in one repeat is termed “indeterminate” rather than MSI low

MSI Electropherogram Results



Microsatellite-High (MSI-H)	Instability at ≥ 2 loci (usually all 5)
Microsatellite-Low (Indeterminate)	Instability at 1 locus
Microsatellite stable (MSS)	None of 5 loci unstable

IHC in Lynch Syndrome

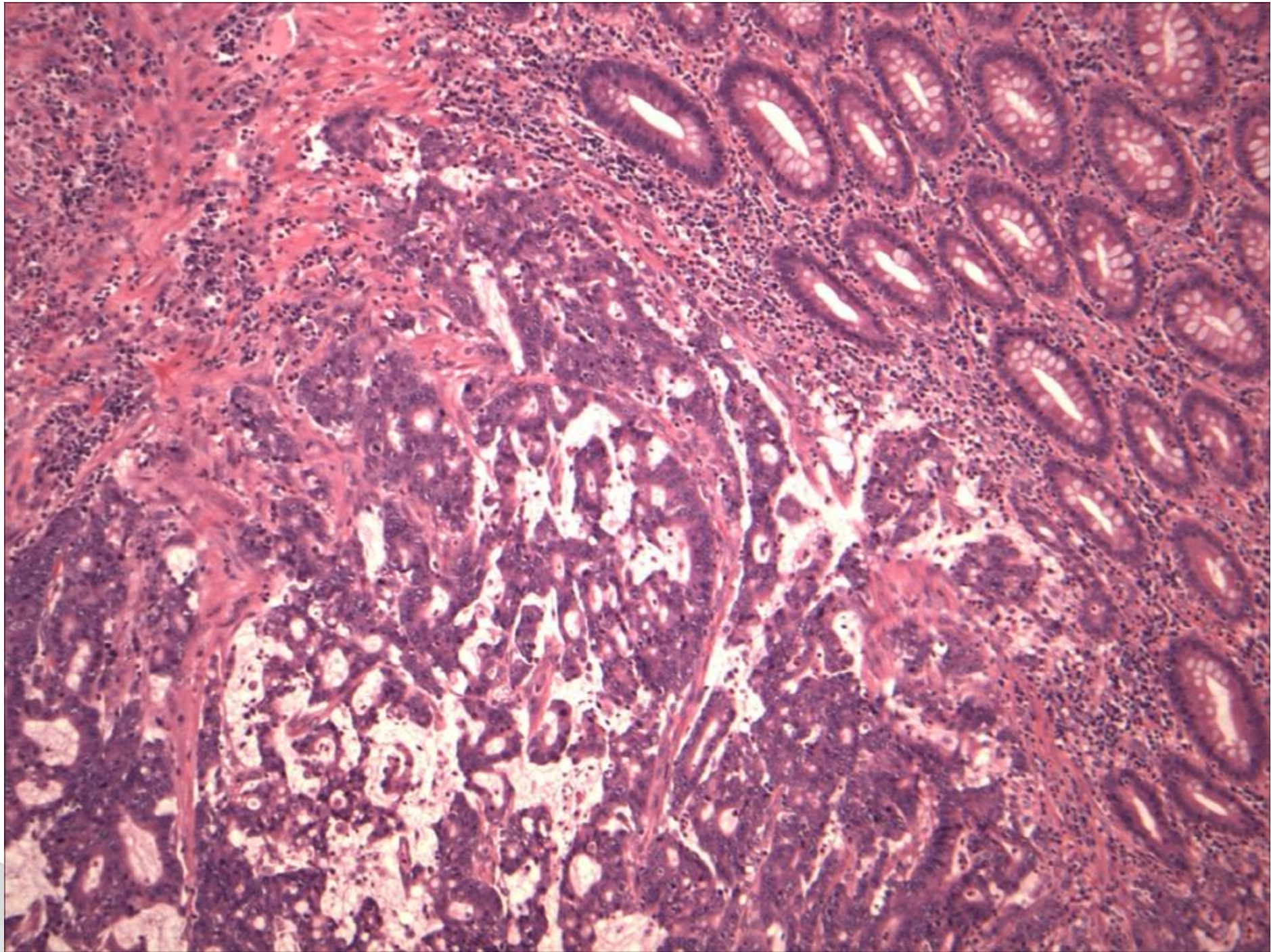


How do we interpret MMR IHC stains?

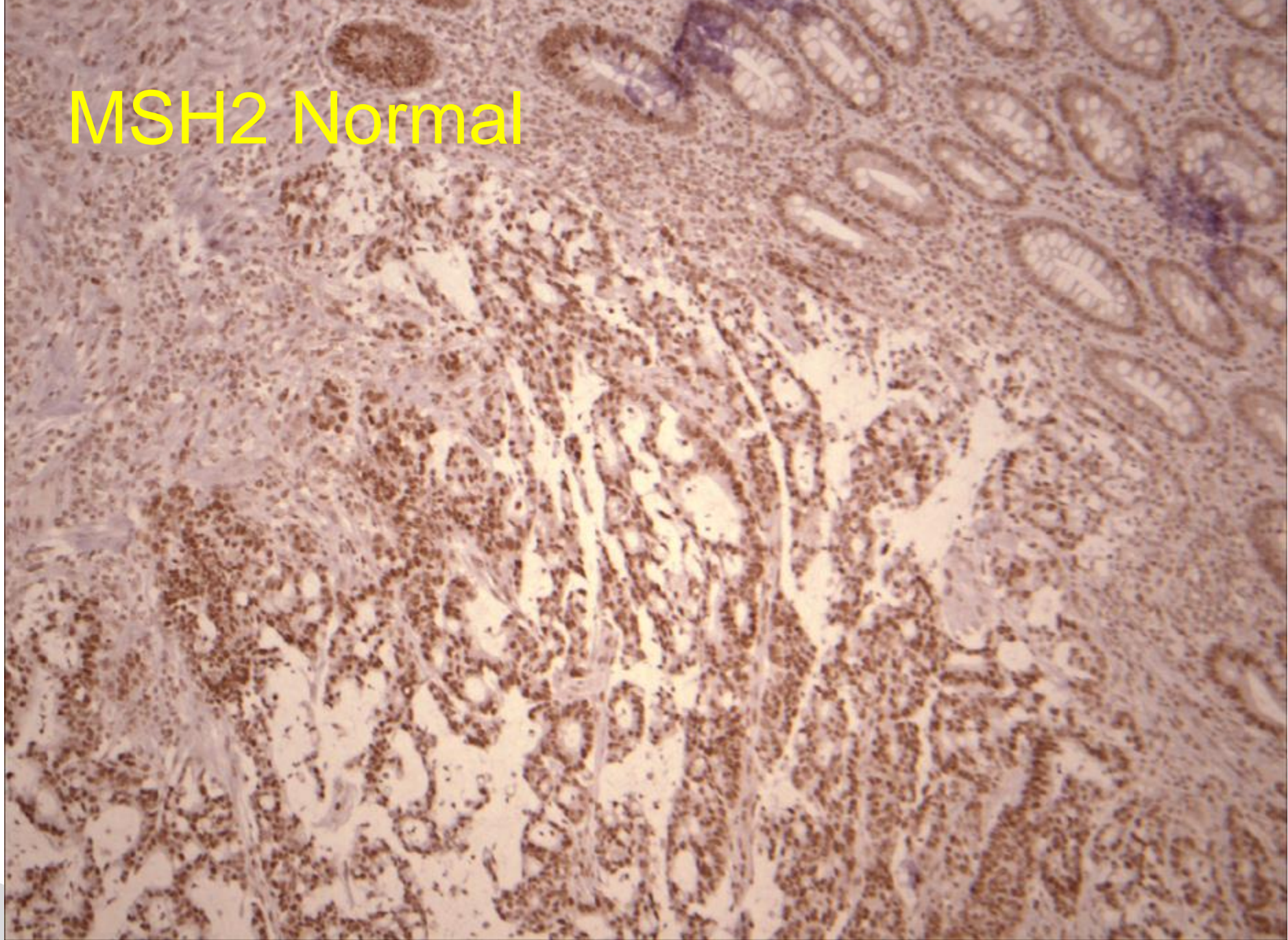
- Two MMR protein complexes:
 - MLH1/PMS2
 - MSH2/MSH6
- Stability of PMS2 and MSH6 depends upon these complexes
- Therefore, loss of staining of MLH1 leads to PMS2 loss
- Similarly, loss of staining of MSH2 leads to MSH6 loss
- MLH1 and MSH2 are stable without the complex; therefore, MSH6 or PMS2 mutations result in *solitary* IHC losses of either protein

IHC interpretation

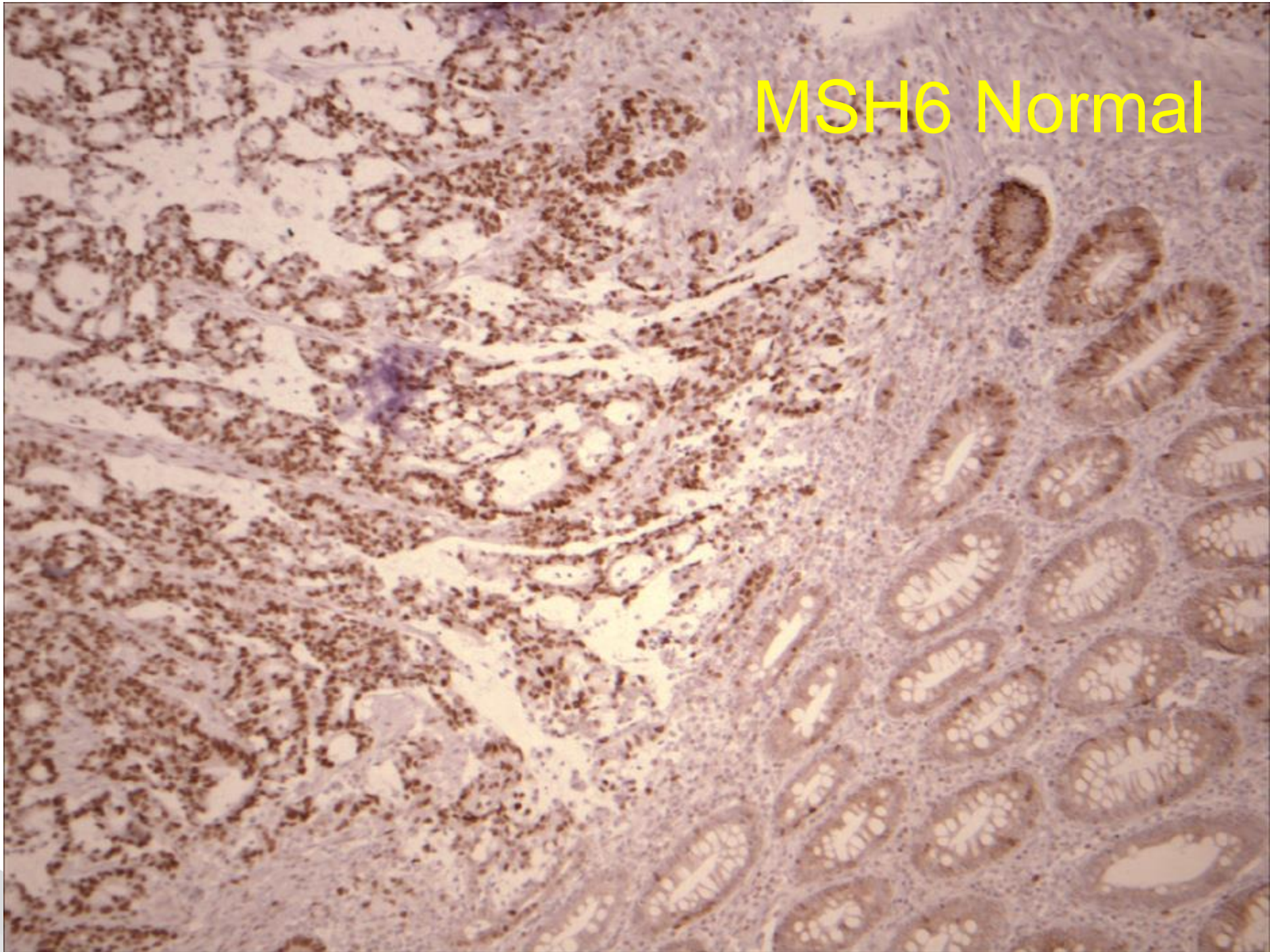
- Defect in MLH1: loss of MLH1/PMS2
- Defect in MSH2: loss of MSH2/MSH6
- Defect in MSH6: *isolated* loss of MSH6
- Defect in PMS2: *isolated* loss of PMS2
- There are exceptions
 - Isolated loss of PMS2 has been associated with MLH1 mutations
- Panel testing makes this less important



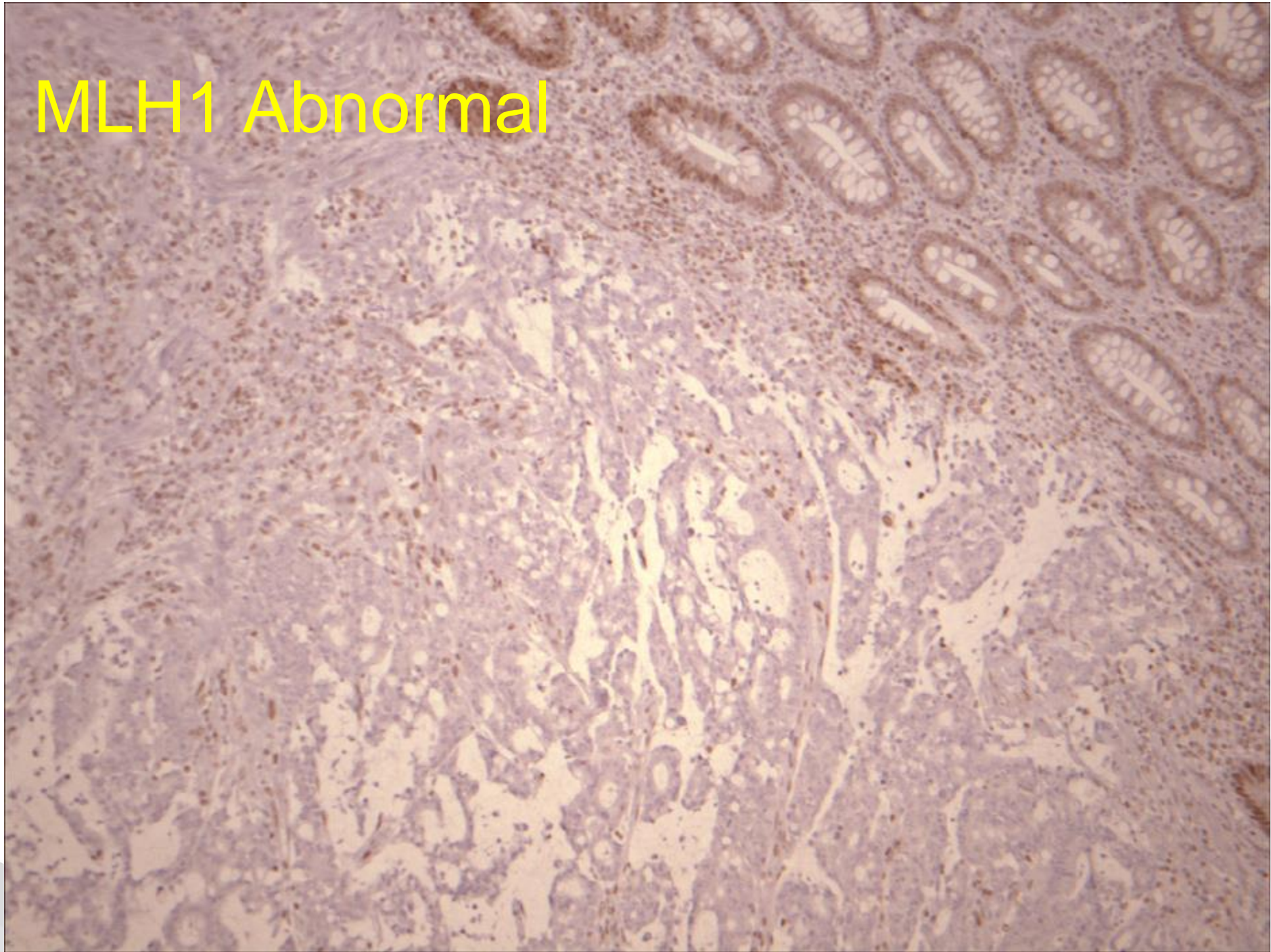
MSH2 Normal



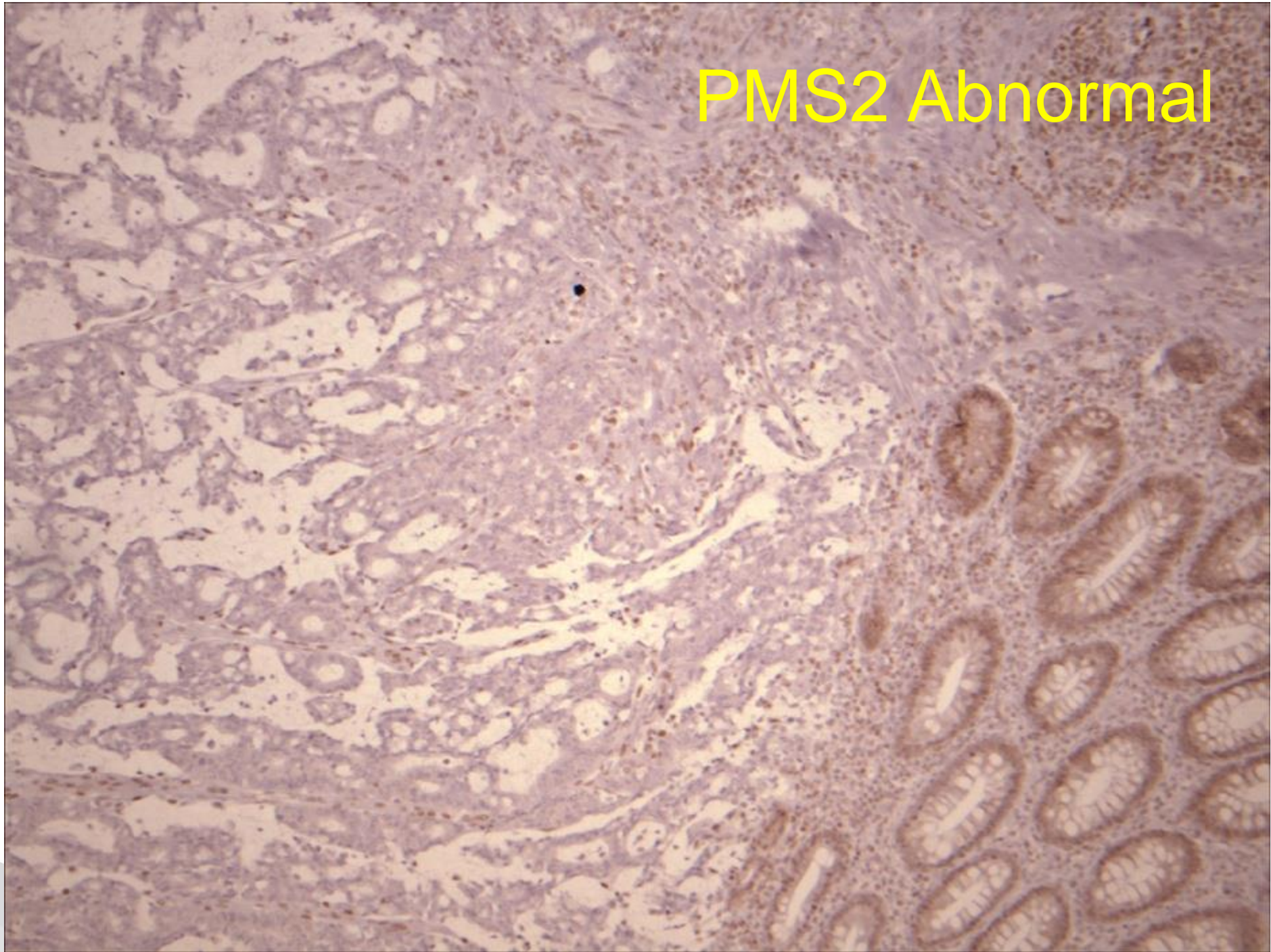
MSH6 Normal



MLH1 Abnormal



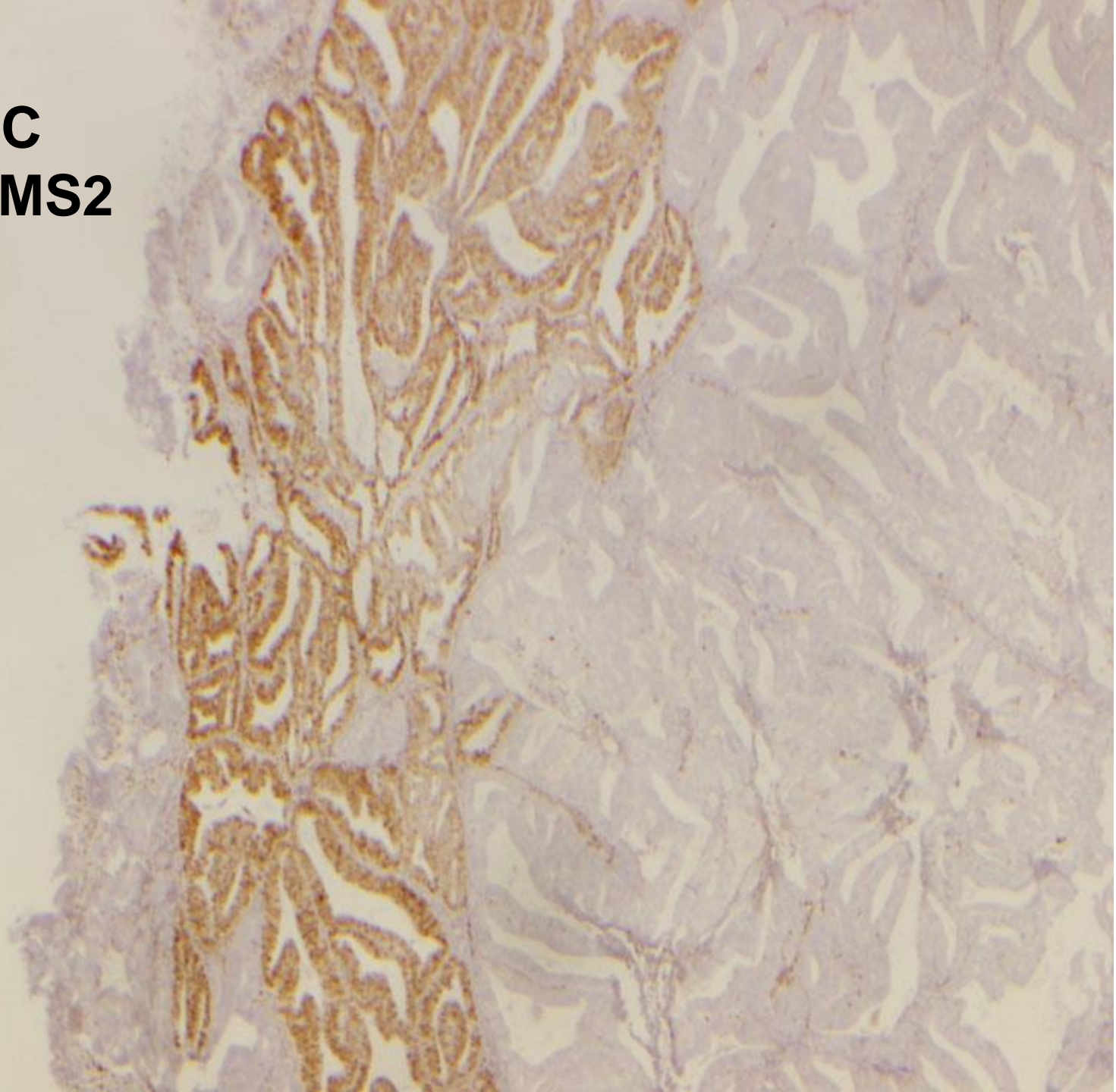
PMS2 Abnormal



“Clonal” MSH6 loss

- Due to instability in a coding mononucleotide repeat in MSH6 (Shia, Modern Path 2013)
- Leads to focal (sometimes nearly complete/complete) MSH6 loss
- Primary cause of instability usually something else
 - MLH1 defect, either acquired methylation or germline
 - PMS2 defect

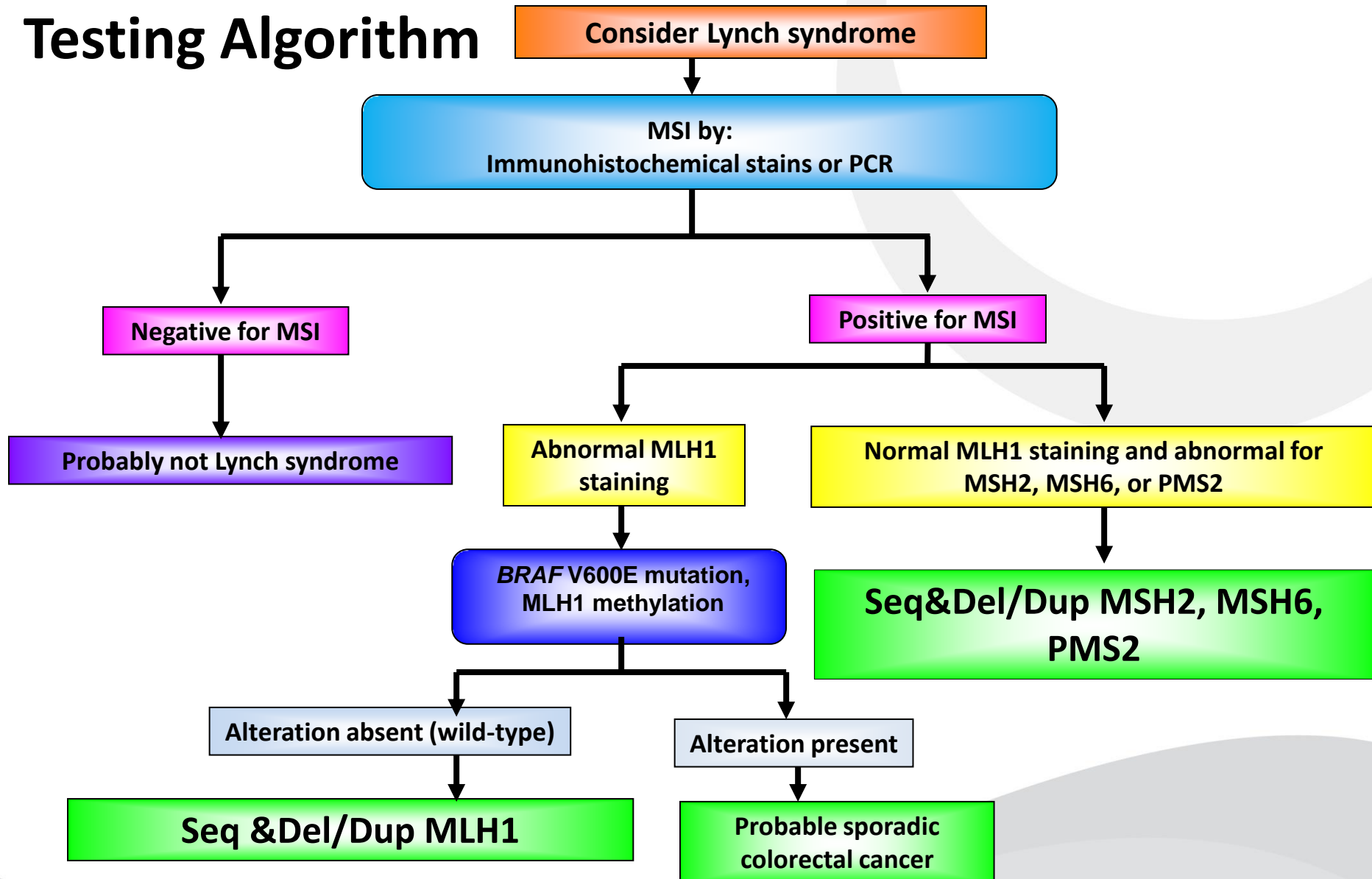
**MSH6 IHC
(MLH1/PMS2
loss)**



Sporadic vs Lynch CRC

- Family history
- MSH2, MSH6, PMS2 IHC loss
- MLH1 promoter methylation (sporadic)
- BRAF point mutation V600E (sporadic)
 - NOTE: Not applicable to non-CRC tumors:
endometrial
- *Germline* MMR gene mutation

Lynch Syndrome Testing Algorithm



Mistake #1: IHC controls

- MMR IHC requires use of known positive and negative controls
 - Need 2 known control tumors: one with MSH2 loss, the other with MLH1 loss
 - Need two MMR stains: PMS2 & MSH6
 - 2 control slides per run with punches of both control tumors stained by PMS2 and MSH6
- Run these controls with *every* MMR IHC run
 - Need to see that antibodies stain tumors they should stain, and don't stain tumors they shouldn't
 - A tonsil doesn't show you this
 - Normal internal control cells surrounding a CRC (ex: lymphocytes) don't show you this

U149 PMS2 OV



CONTROL
ARUP

12/28/14

18

U176 MSH6 OV



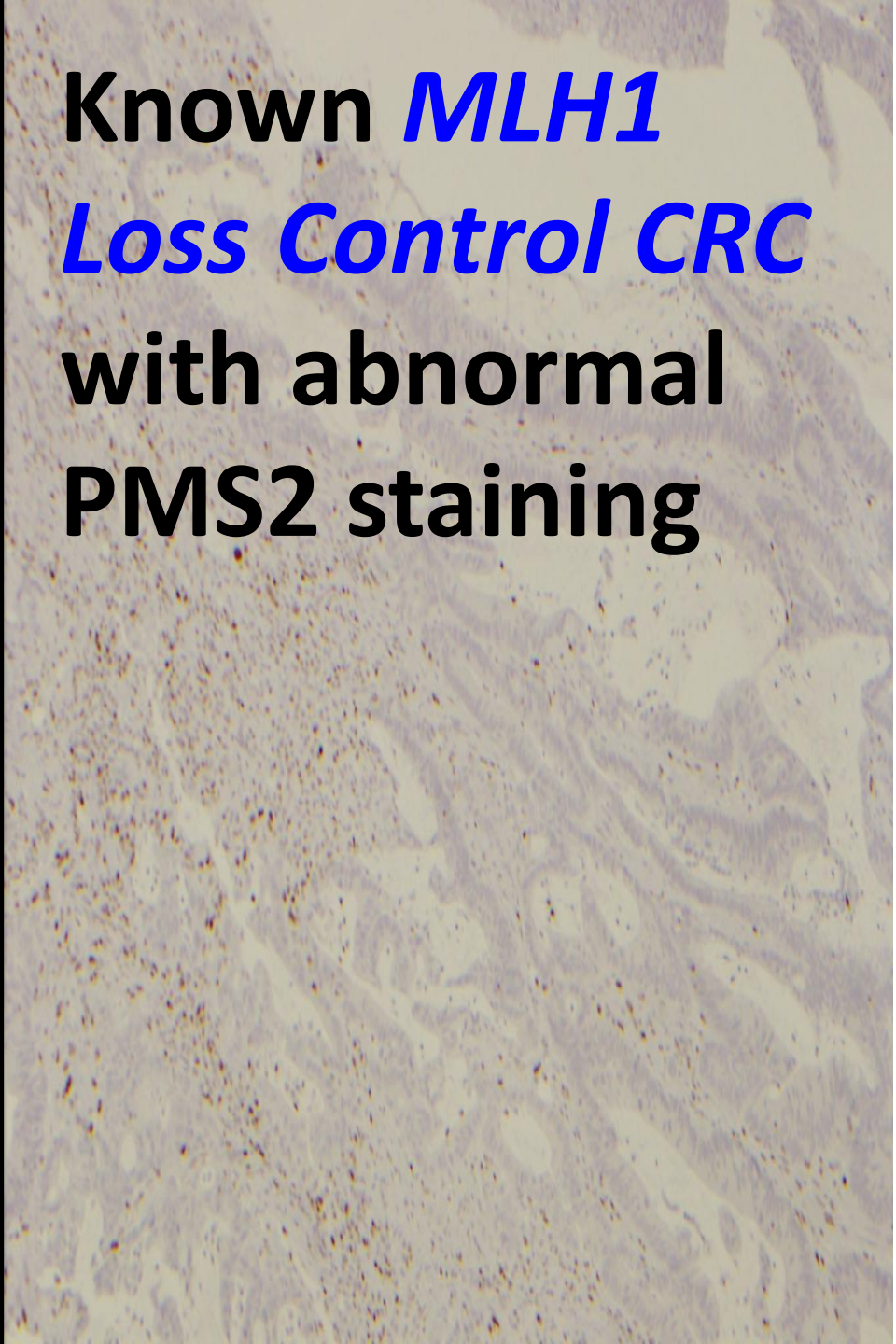
CONTROL
ARUP

12/28/14

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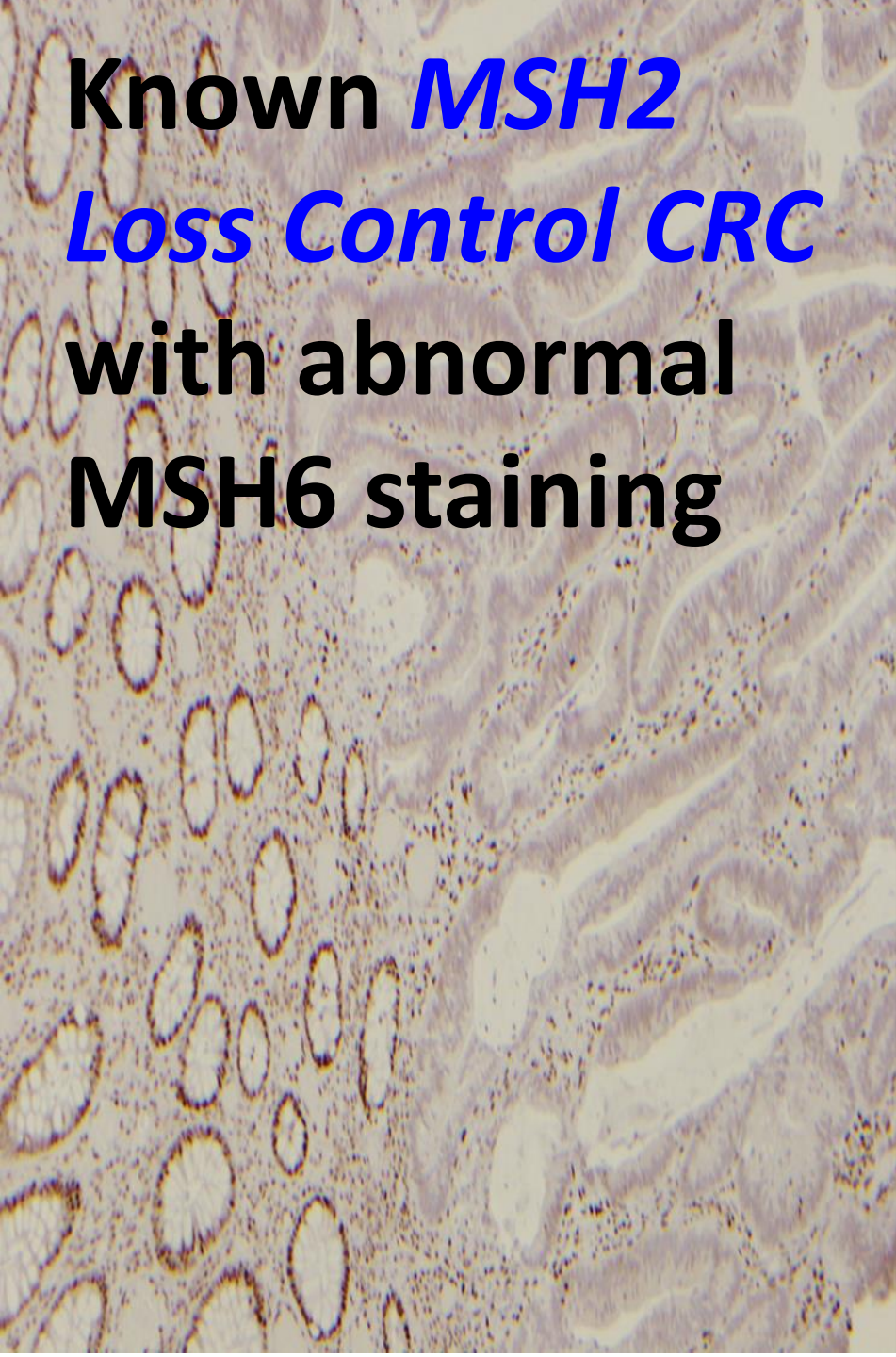
This image shows a histological section of colorectal tissue. The glandular structures are well-organized and lined by a single layer of columnar epithelial cells. The nuclei are stained dark brown, indicating normal PMS2 expression. The overall architecture is typical of healthy colonic mucosa.

Known *MSH2*
Loss Control CRC
with normal
PMS2 staining

This image shows a histological section of colorectal tissue. The glandular structures are present but the staining is significantly reduced or absent, particularly in the nuclei of the epithelial cells, indicating a loss of PMS2 protein. This is characteristic of a microsatellite instability (MSI) phenotype.

Known *MLH1*
Loss Control CRC
with abnormal
PMS2 staining

Known *MSH2*
Loss Control CRC
with abnormal
MSH6 staining

This image shows a histological section of colorectal cancer tissue. The glandular structures are visible, but the brown staining for MSH6 is significantly reduced and patchy, indicating a loss of normal MSH6 expression. This is characteristic of a microsatellite instability-high (MSI-H) or loss of control of cell cycle (LCC) CRC.

Known *MLH1*
Loss Control CRC
with normal
MSH6 staining

This image shows a histological section of colorectal cancer tissue. The glandular structures are visible, and the brown staining for MSH6 is strong and uniform across the tumor cells, indicating that MSH6 expression is maintained. This is characteristic of a microsatellite stable (MSS) or loss of control of cell cycle (LCC) CRC.

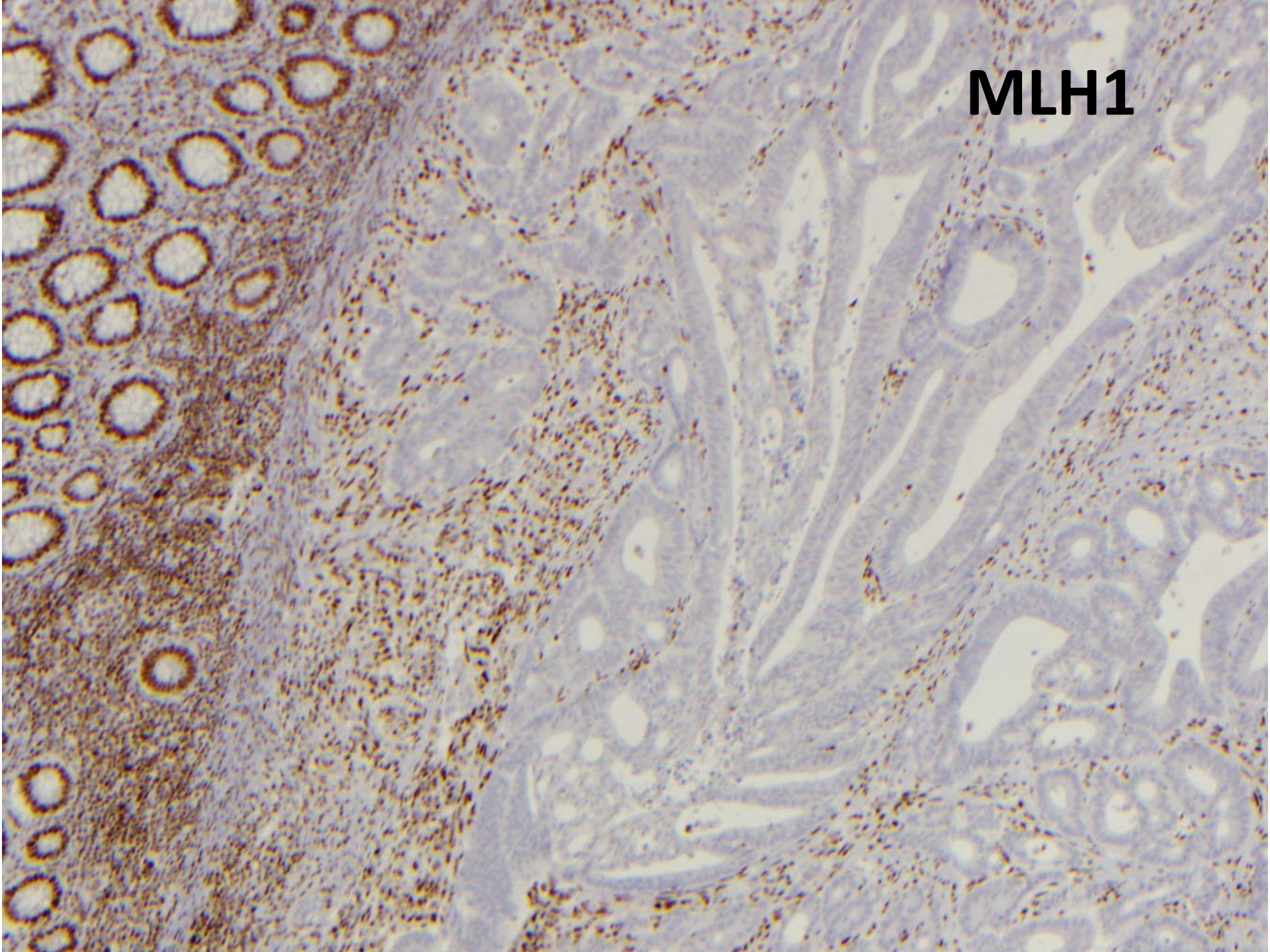
Mistake #2: Reporting IHC results

- Don't describe IHC staining as “positive” or “negative”—confusing
- Report clearly; get feedback from clinicians (we say “normal” and “abnormal”)
- Don't report results that no one sees or acts upon
 - Interact with colleagues who deal with results
 - Make sure your reports are comprehensible and clinicians are reacting appropriately (genetic counselors probably best)

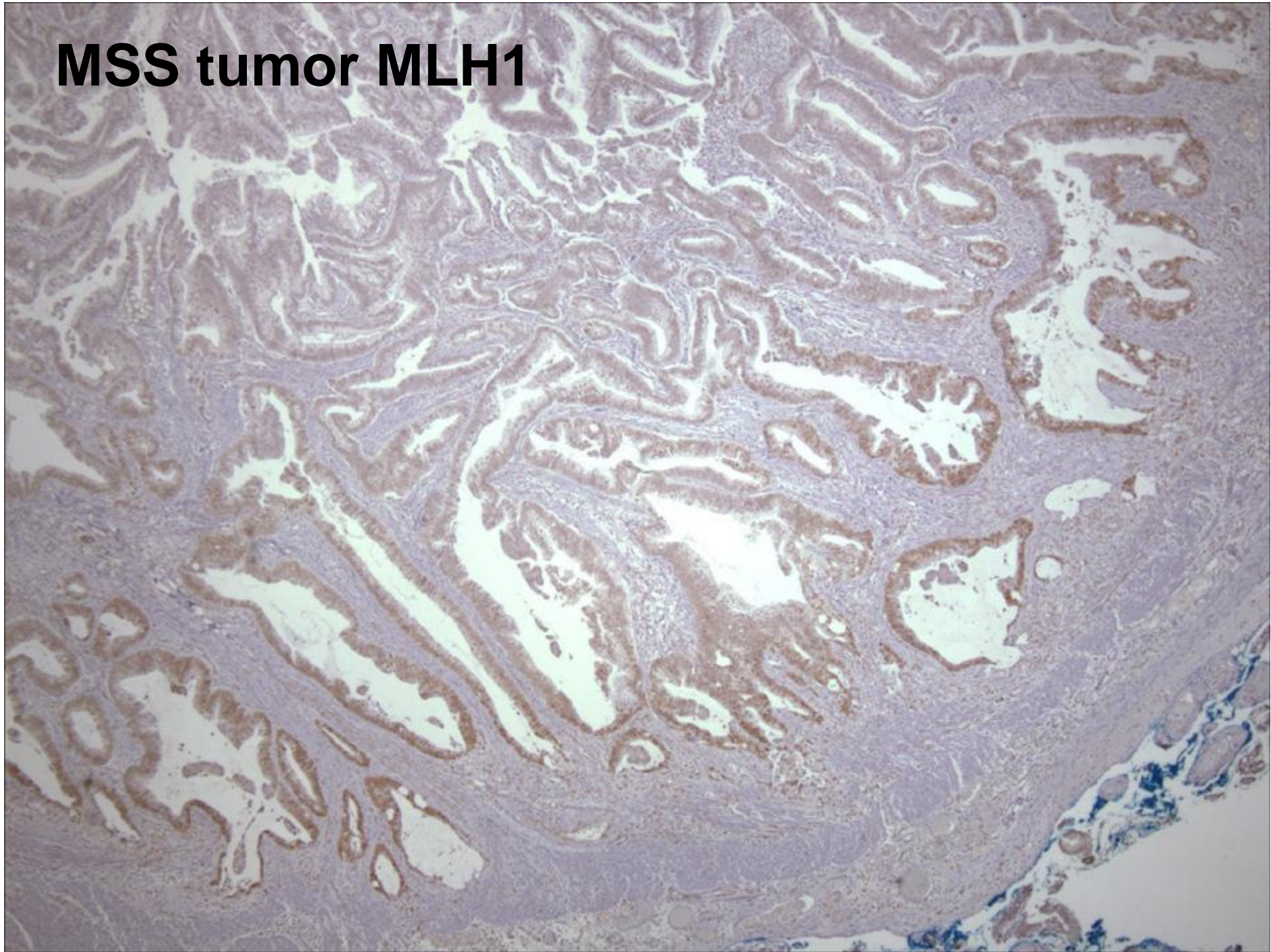
Mistake #3: IHC interpretation

- Loss of tumor staining without contiguous *internal control* staining is uninterpretable: don't call this abnormal
- Decreased staining intensity, unless quite marked, probably doesn't mean anything: this is a qualitative test
 - If marked, suggest confirmatory MSI PCR testing

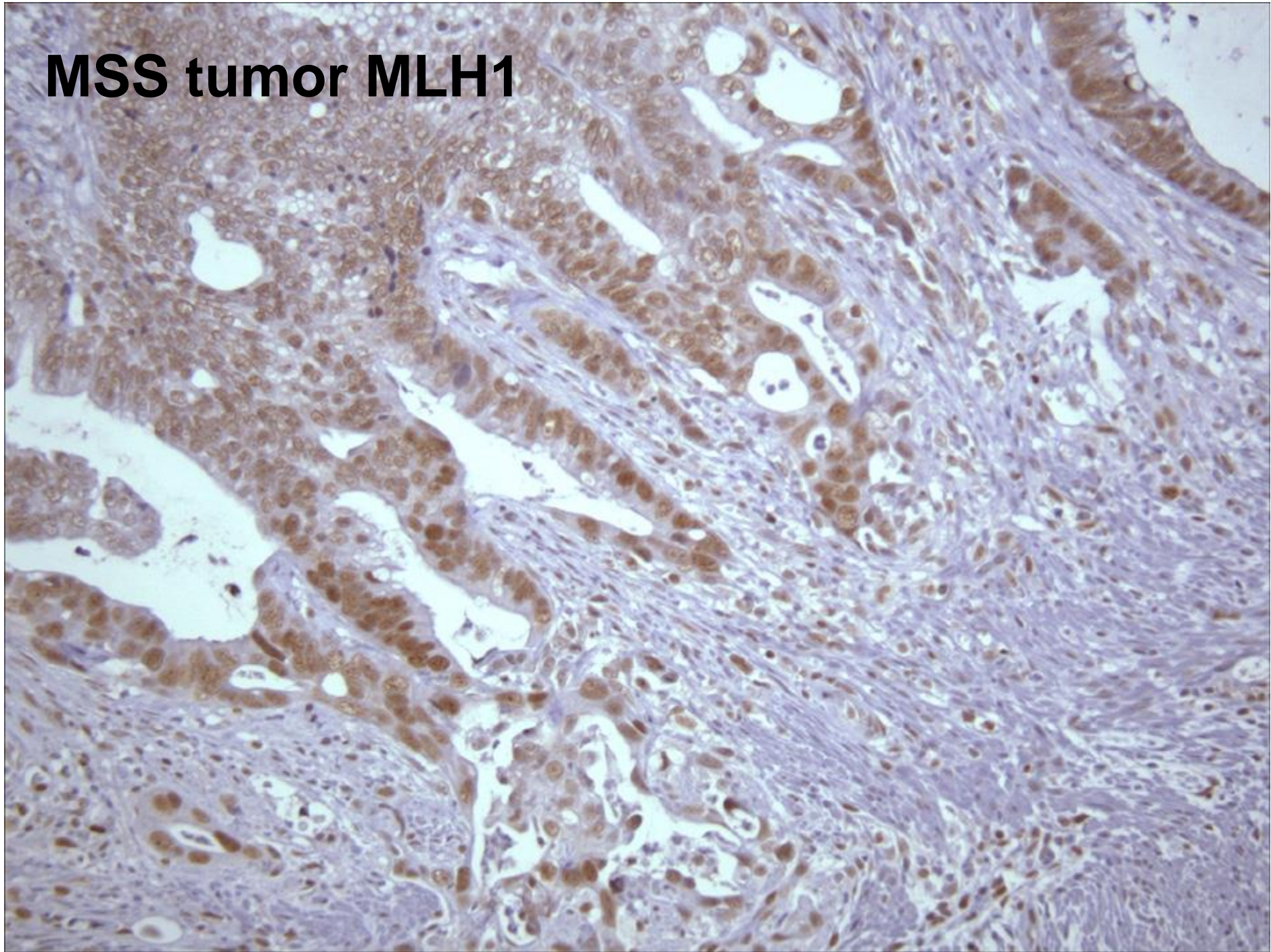
MLH1



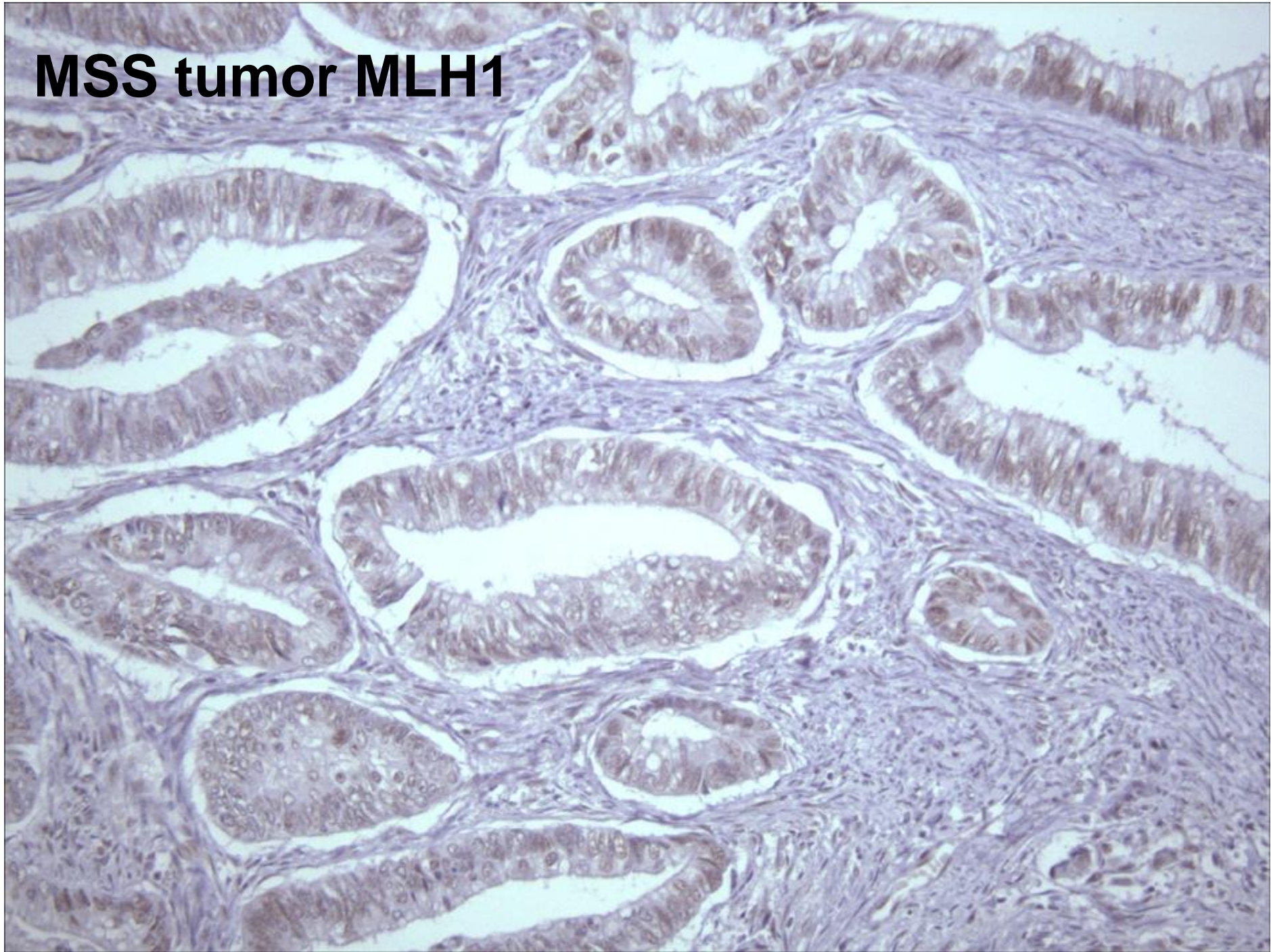
MSS tumor MLH1



MSS tumor MLH1



MSS tumor MLH1



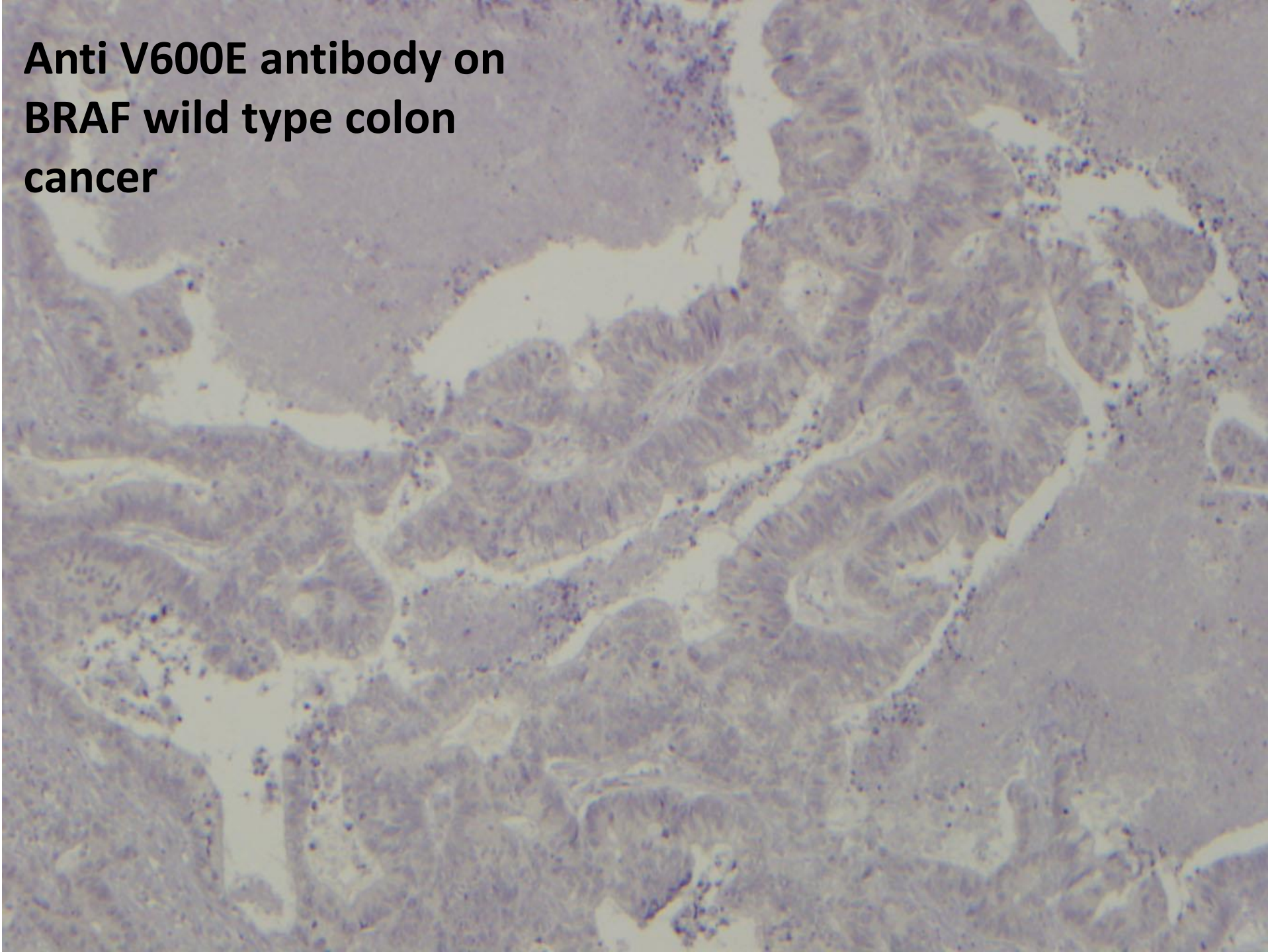
Mistake #4: Inappropriate BRAF testing

- Non-colorectal (e.g., **endometrial**) cancers rarely mutate BRAF, DON'T ORDER TEST
- Need to test MLH1 methylation for non-colorectal cancers
- Need to test MLH1 methylation for potentially sporadic colorectal cancers without BRAF mutations

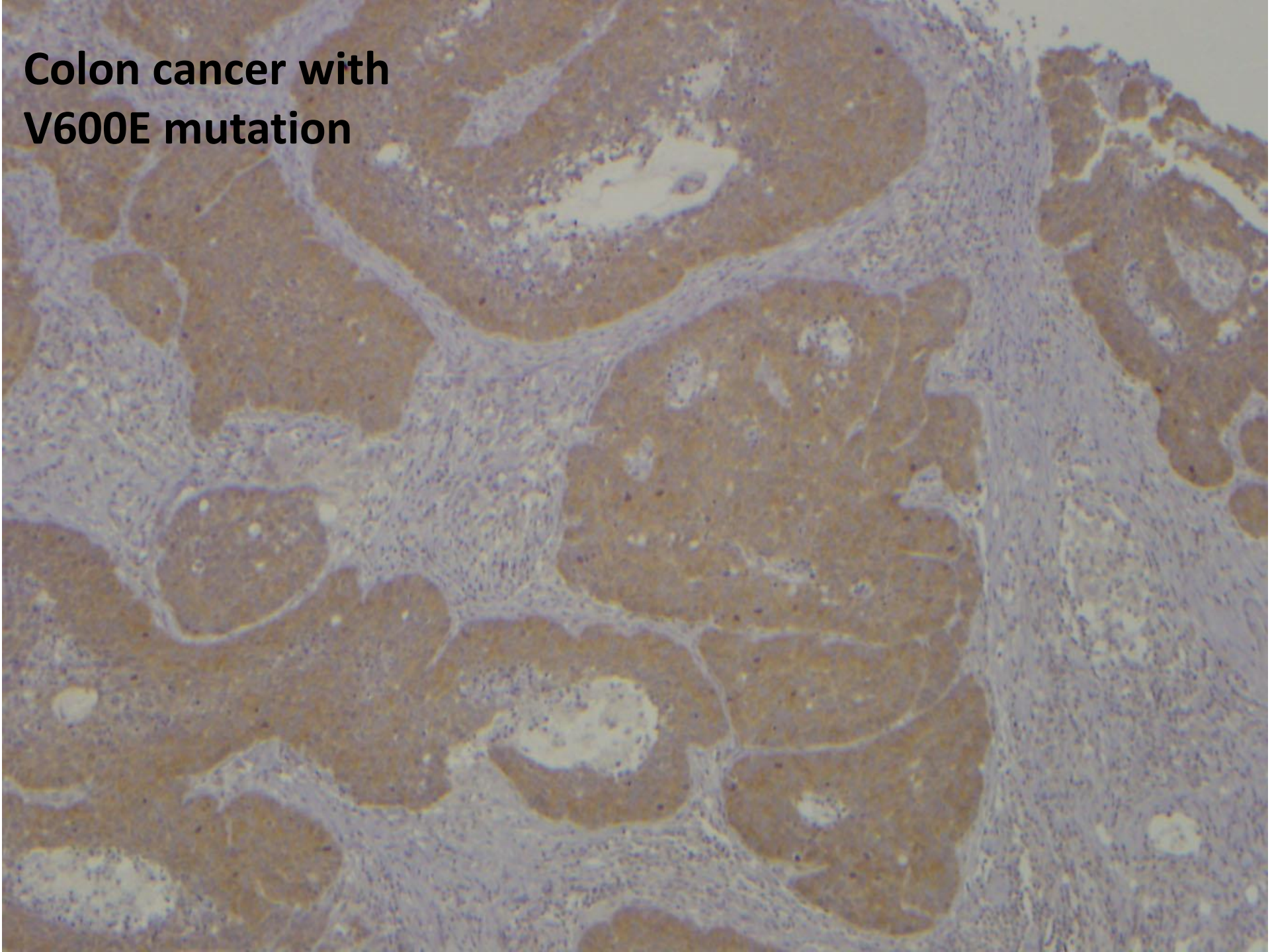
Mistake #5: All IHC Lynch work-up

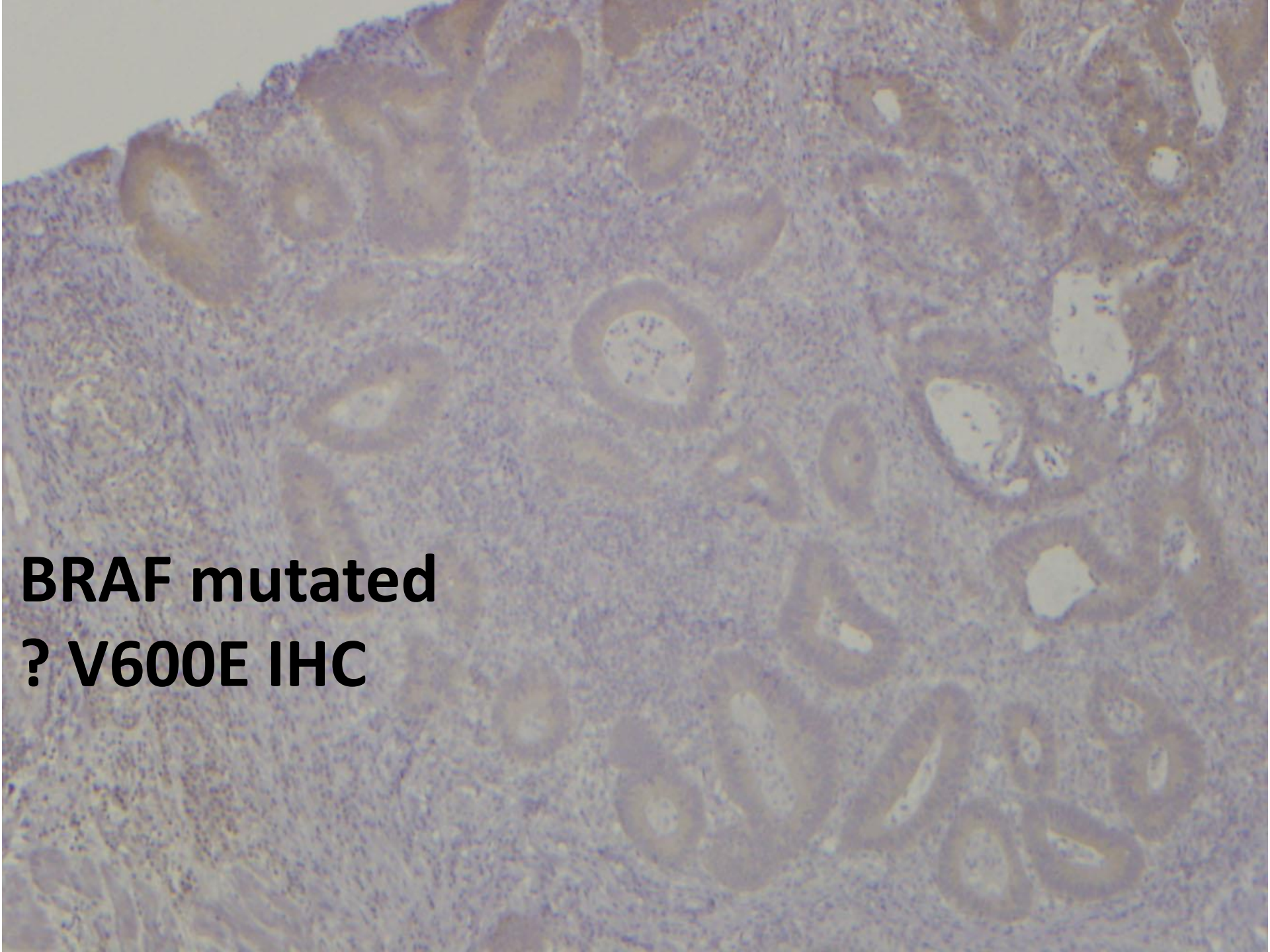
- BRAF antibody: detects BRAF V600E mutation (Affolter, Samowitz, Bronner; GCC 2013)
- Has same problems as all other IHC tests, including staining variability and difficult interpretation
- No internal controls for antibody staining
- Research vs. clinical test
 - Clinical test needs to be robust, easily interpretable

**Anti V600E antibody on
BRAF wild type colon
cancer**



**Colon cancer with
V600E mutation**





BRAF mutated
? V600E IHC

Mistake #6: Testing of serrated lesions

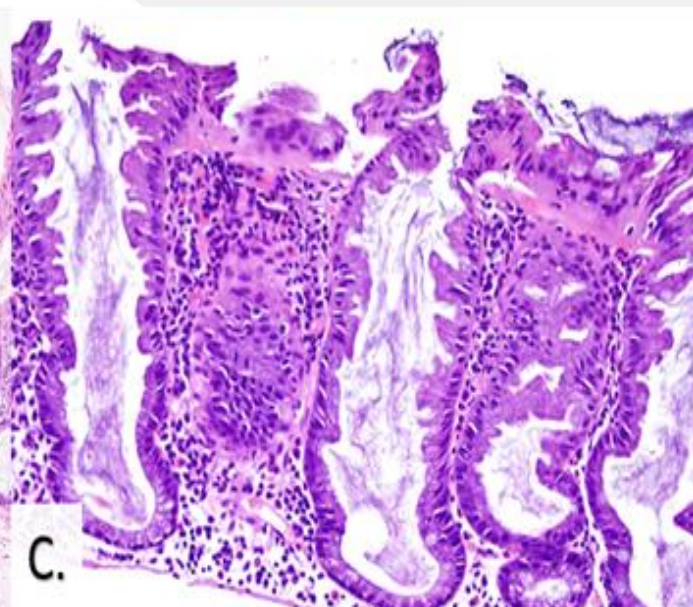
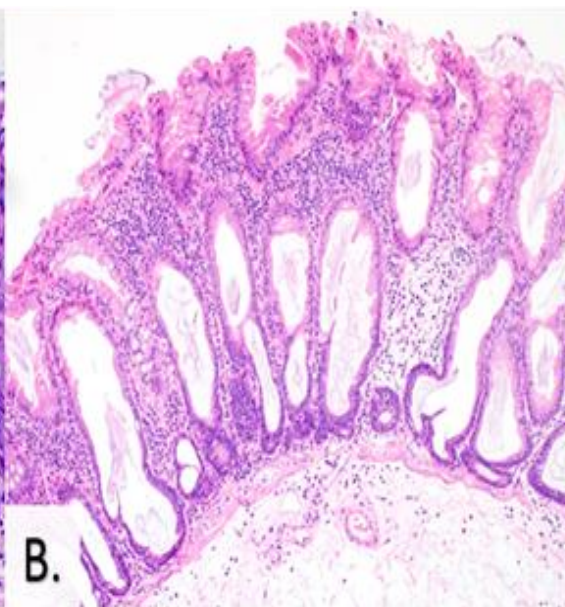
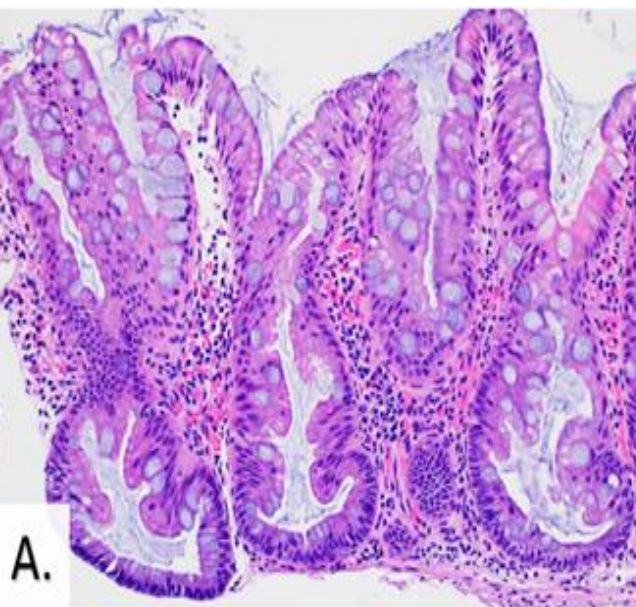
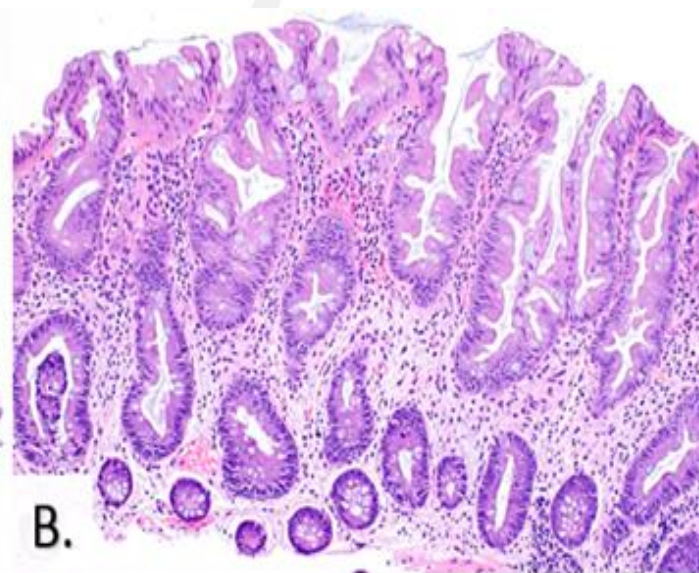
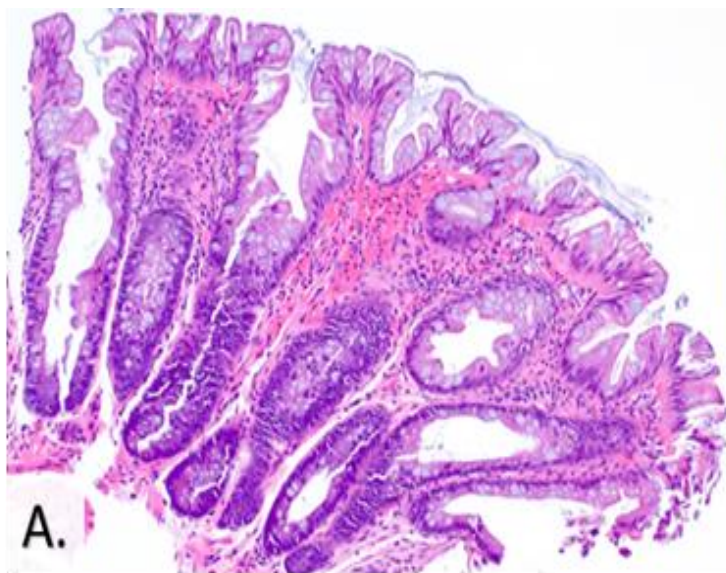
- Evaluating serrated lesions for *MMR deficiency: USELESS*
 - Based on incorrect notion that MSI will separate SSP's from HP's
- Evaluating serrated lesions for *BRAF mutations: USELESS*
 - Both SSP's and HP's commonly have BRAF mutations

SSP vs. HP

- No molecular test reliably separates these lesions
- Use polyp site (R vs L), size (>2 biopsies), and histology to distinguish these lesions

Criteria to Distinguish HP from SSL

Criteria	ROC AUC
Morphology	69.3%
Size (cutoff bx number 2 or more)	87.3%
Endoscopic size	55.2%
Location	82.3%
Morphology, Size and Location	93.7%





KRAS Testing

The Metastasis Problem

- 50-60% CRC patients present with or develop metastases
- 5-yr survival

Stage I + II (N0) → 91%

Stage III (N1,2) → 70%

Stage IV (M1) → 11%

Search for Alternative Rx's

5 FU/Leucovorin mainstay for decades

After 2000 → New Therapies

Oxaliplatin (FOLFOX)

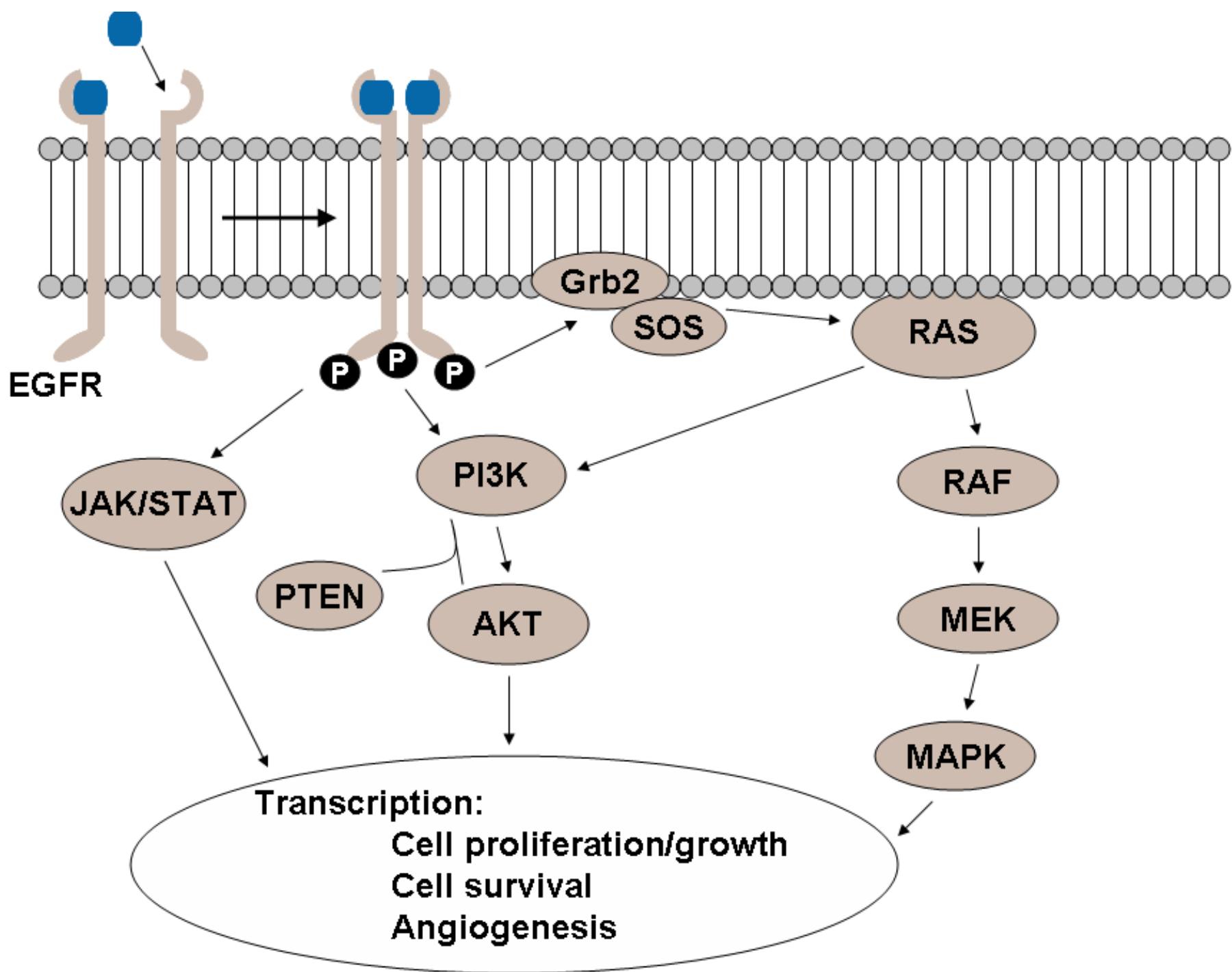
Irinotecan (FOLFIRI)

Anti-VEGF (bevacizumab)

Anti-EGFR (cetuximab, panitumumab)

EGFR inhibitor therapy for CRC

- EGFR pathway is activated (but EGFR is not mutated) in colorectal cancer
- Cetuximab is an antibody that binds to EGFR and turns off EGFR pathway
- Mutations downstream of EGFR (KRAS, BRAF, PIC3, PTEN) activate the pathway and make EGFR block irrelevant
- Bad to give a toxic and expensive drug if it won't work



KRAS mutation

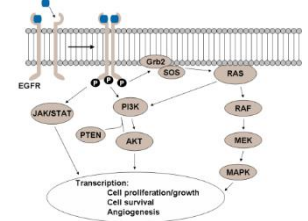
- <1% response rate to anti-EGFR Rx with codon 12 or 13 or 61 mutations (~40% of CRC)
- ~40% response rate with KRAS WT (~60% of CRC)
- But.... ~ 60% KRAS WT do not respond to EGFR inhibition
- Other markers play a role and in clinical trials

KRAS Testing: Cost Savings

- ~30,000 new *metastatic* CRC annually
- KRAS testing = \$13 million (\$452/pt)
- Cetuximab Rx= \$2.1 *billion* (\$71,120/pt)
- Mutated KRAS (~40%) excluded from cetuximab
- Cost savings: ~\$750 million annually
- High toxicity; ~2 month added survival

No need for normal tissue to
test for KRAS (unlike MSI)

Future



- Impact of *specific* KRAS 12/13 mutations?
 - Sotorasib targeted to **KRAS G12C** (~3-7% met CRC) in phase 2 trial did not meet primary endpoint (*Lancet Oncol* 2022;23:115), **unlike in NSCLC**
 - Combination trials of KRAS G12C with EGFR inhibitors or immunotherapy underway
- Other predictors of anti-EGFR response?
 - Other KRAS mutations: codon 61, others?
 - BRAF
 - EGFR copy no. (FISH,CISH,PCR,NGS), specific mutations
 - PTEN, PIK3CA mutations

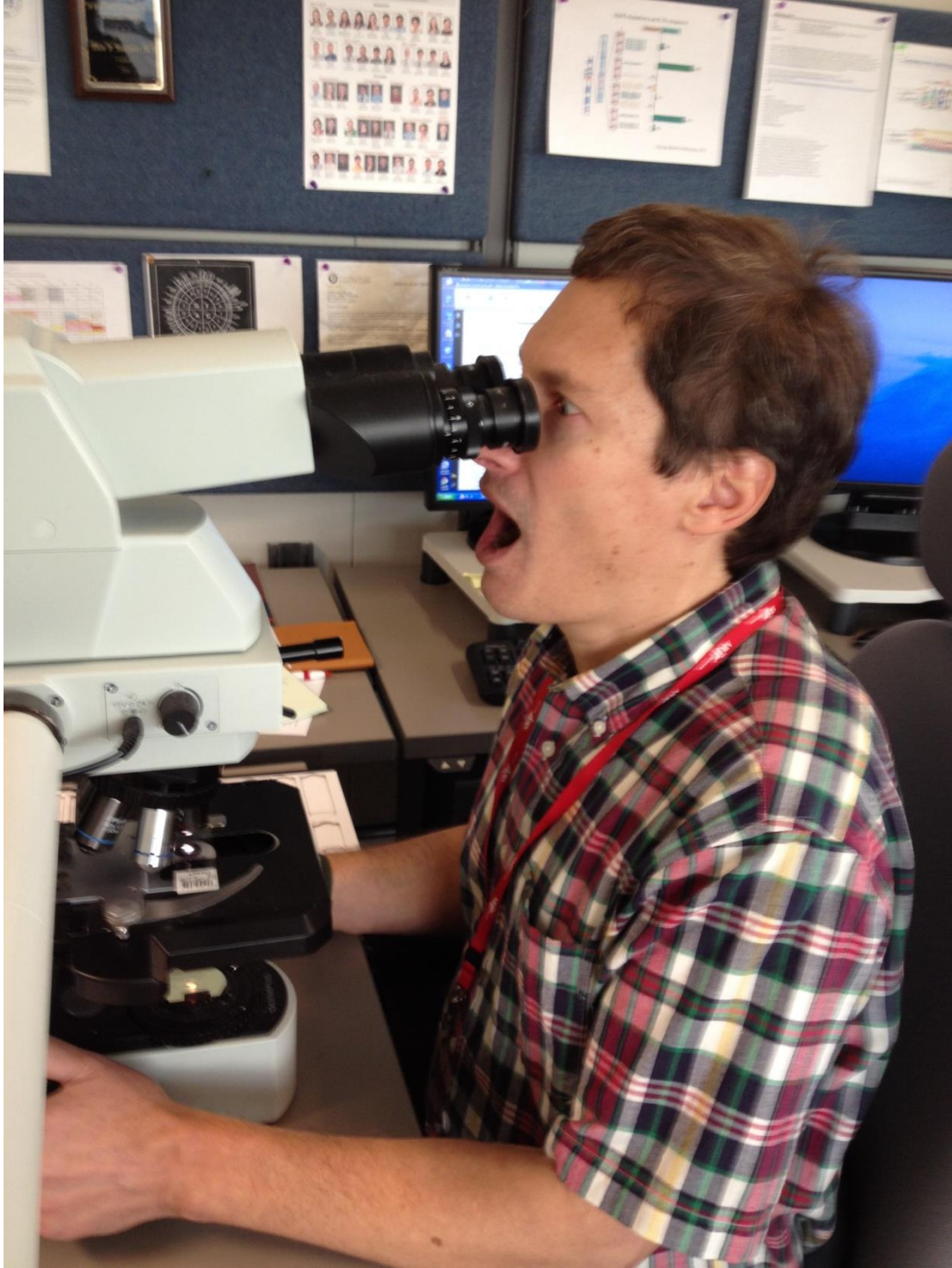
KRAS Summary

- KRAS mutations occur in 30-40% CRC's
- Highly predictive of lack of response to anti-EGFR Rx (such as cetuximab)
- Pathologists play a key role in determining best Rx for stage III-IV CRC
- BRAF, PIK3CA, PTEN downstream markers may be useful in KRAS wild type tumors
- Additional biomarkers expected and many clinical trials underway



What is our role in this?

- Selecting block to test
- Circling tumor
- Maybe performing the test, interpreting results



Mistake #7: Choosing a bad block

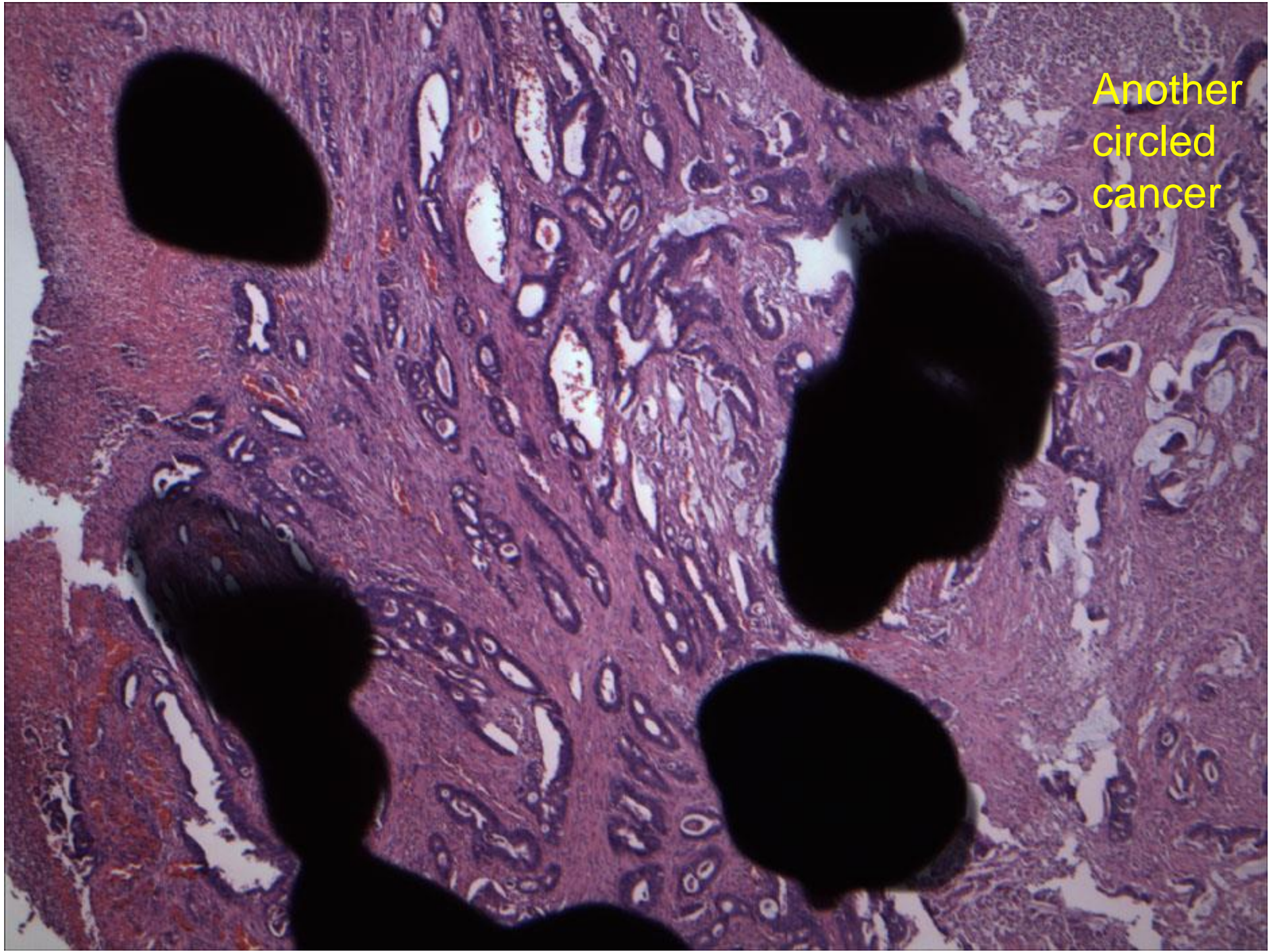
- PCR isn't magic; garbage in, garbage out still applies
- With colon cancer, finding a block with sufficient tumor usually isn't a problem
- Rectal cancers resected after chemoradiation may be hypocellular; often better to choose pre-treatment biopsy
- Don't use decalcified specimens, specimens fixed in unusual fixatives

Mistake #8: Poor circling of tumor

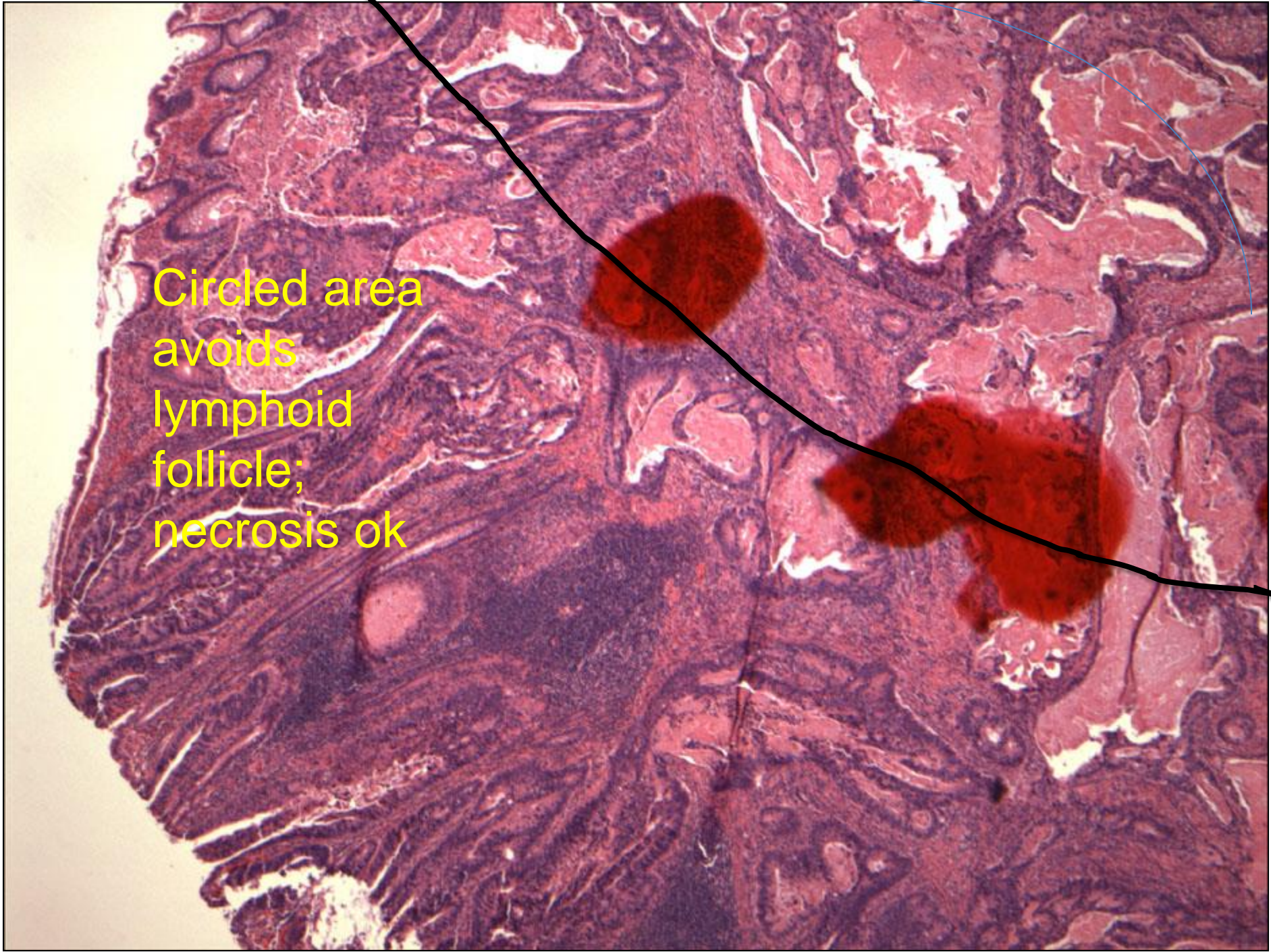
- Avoid (as much as possible) contaminating *normal cells* (lymphoid follicles, abscesses, muscle)
 - Don't be ridiculous about this, most tests will work with about 20% tumor, usually easily achievable with colon cancer
- Don't need all of the tumor
 - No need to “gerrymander” the circled area
 - Difficult to dissect, wastes everyone's time



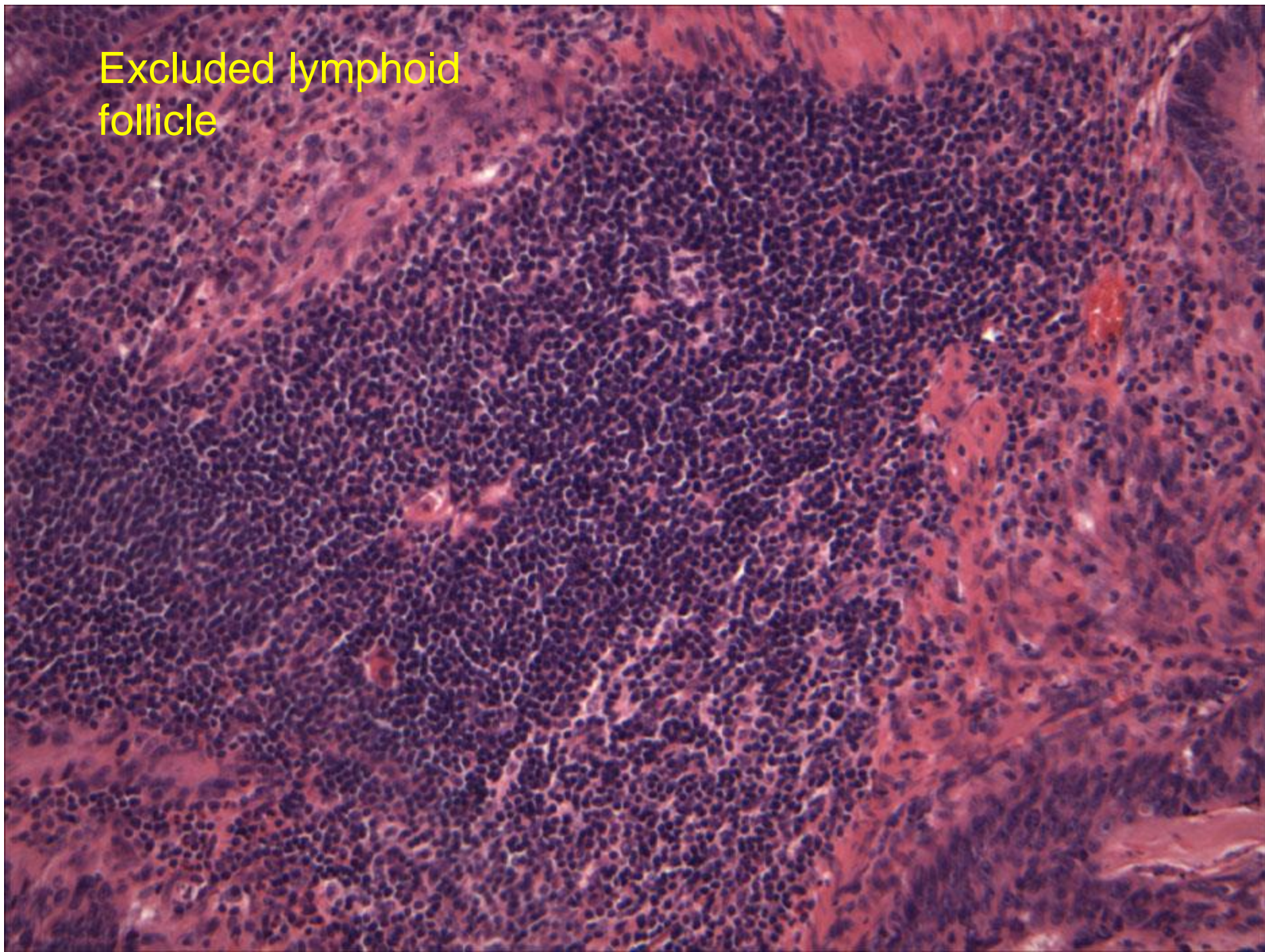
Another circled cancer

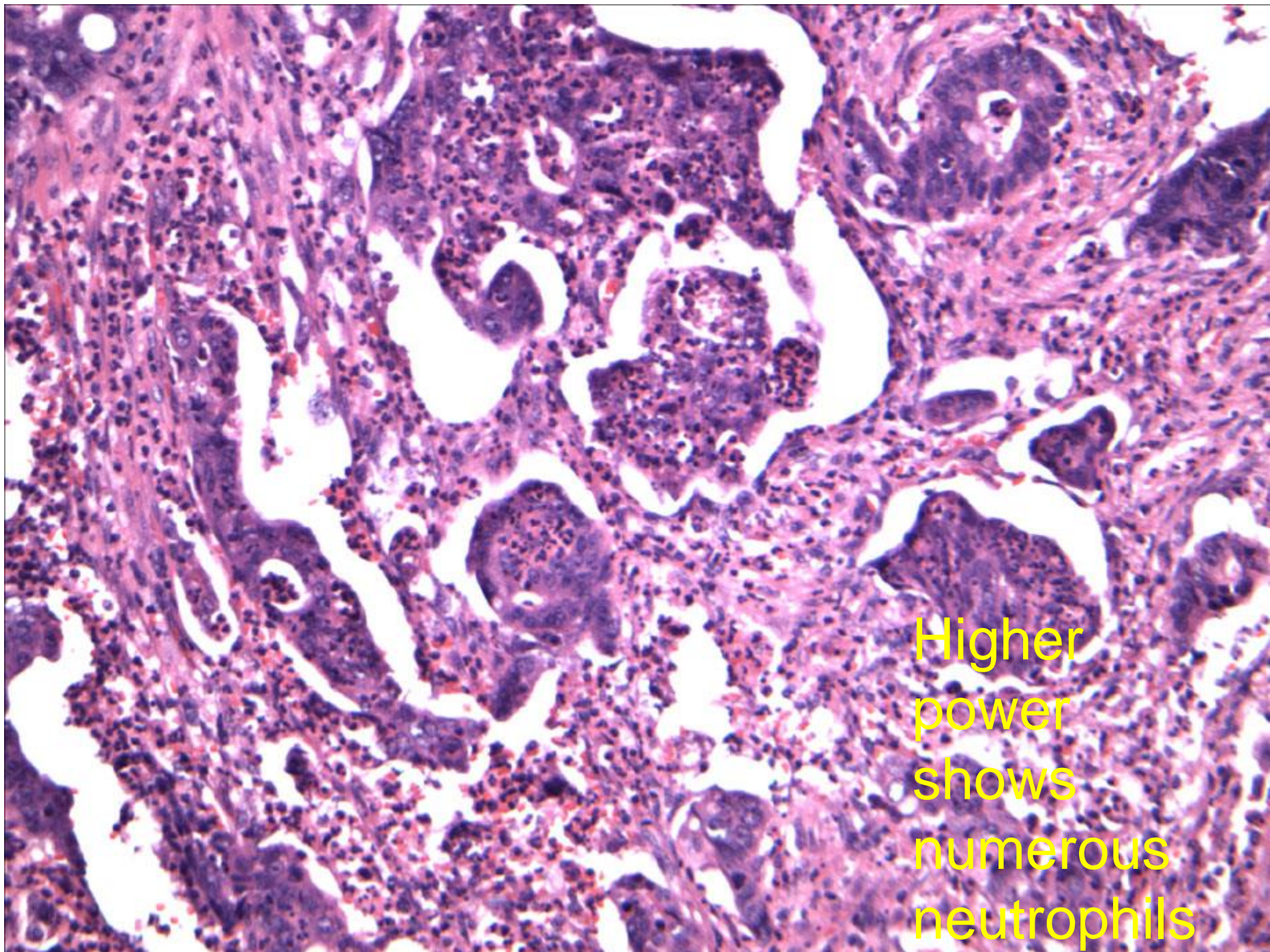


Circled area
avoids
lymphoid
follicle;
necrosis ok



Excluded lymphoid
follicle





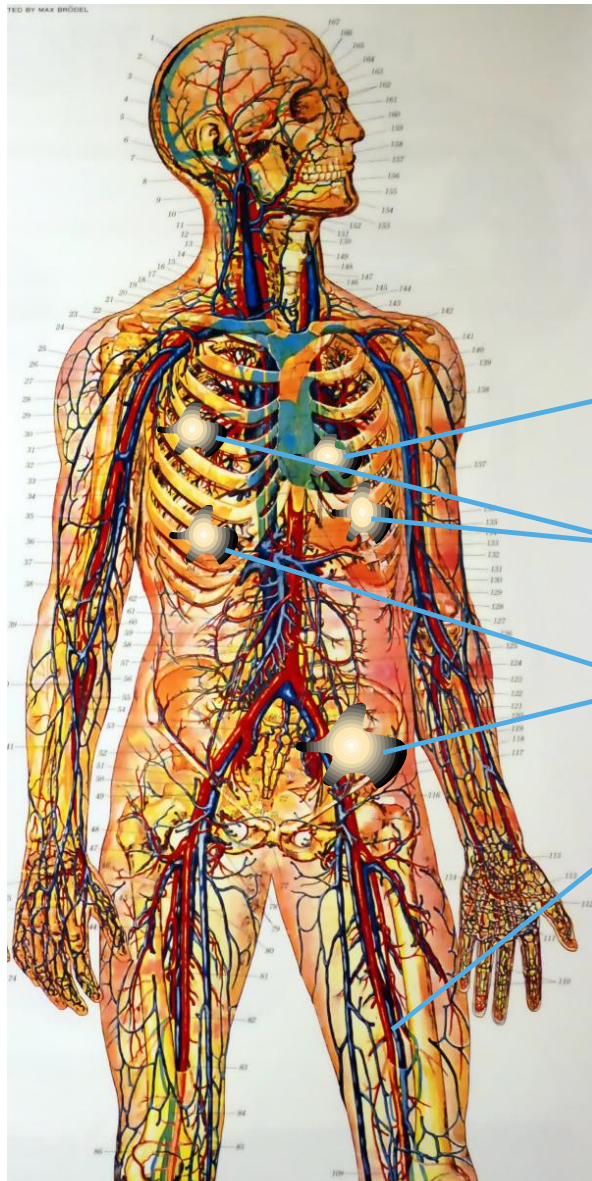
Higher
power
shows
numerous
neutrophils

Mistake #9: Assuming tumor homogeneity

- Different areas of a tumor, different metastases may have different mutations
- We unfortunately ignore this by evaluating one part of a primary, or one of many metastases
- Evaluation of circulating tumor DNA may be a way to get a mutational evaluation of the entire tumor burden (for review see Heitzer, Clinical Chemistry, 2015)

ctDNA reduces sampling error and allows analysis of entire tumor burden: primary/mets/heterogeneous clones

Blood is the “window to the body”

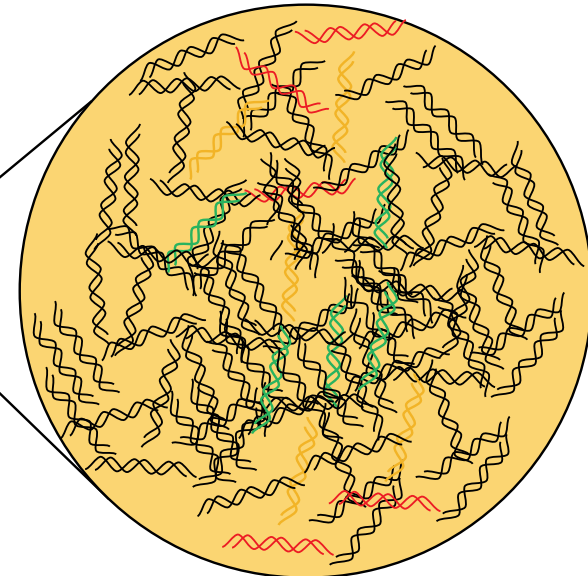


“Tumor5”


“Tumor3+4”


“Tumor1+2”


“Normal”

Summary: CRC Molecular Dx

- Sporadic MSI-H CRC (15%): MMR IHC, MSI, MLH1 Methylation, BRAF
- Lynch MSI-H CRC (2-3%): MMR IHC, MSI, MLH1 Methylation, BRAF
- Metastatic CRC KRAS WT (50-60%) selects for Anti-EGFR therapy
- KRAS G12C specific inhibitor therapy
- KRAS WT Non-responders to Anti-EGFR (60%): work continues



What about EPCAM?

- EPCAM is just five prime of MSH2
- Three prime EPCAM deletions lead to transcriptional read through, MSH2 methylation and Lynch syndrome
- EPCAM deletions associated with similar colon cancer risk as MSH2 mutations, but less of an endometrial cancer risk

Does EPCAM IHC help in Lynch work-up?

- Standard MMR IHC won't miss Lynch due to EPCAM deletions
 - IHC profile will be MSH2/MSH6 loss
- Standard germline genetic analysis for MSH2 will detect EPCAM deletions
 - Already includes probes for EPCAM deletions