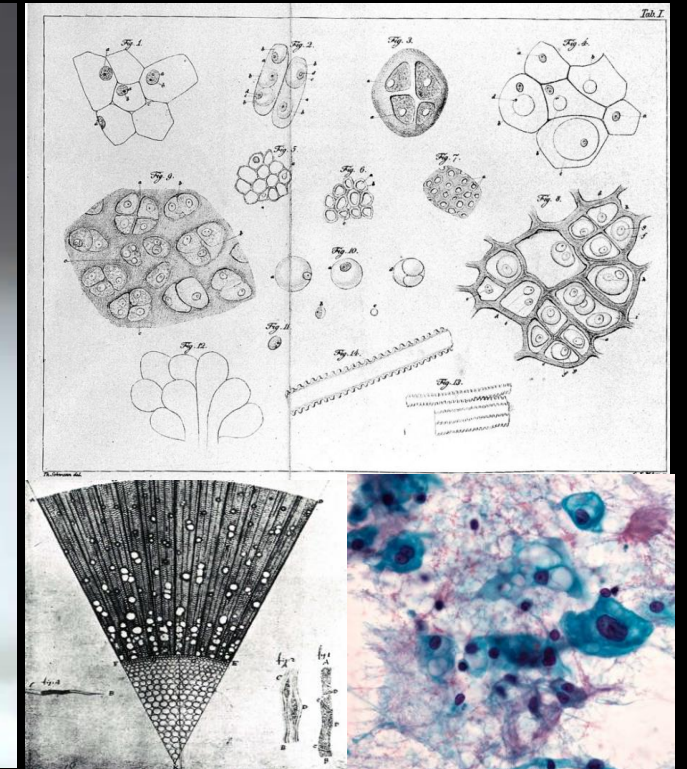


Go with the Flow: Clinical Insights Through Flow Cytometry

Alexis S. Dadelahi PhD

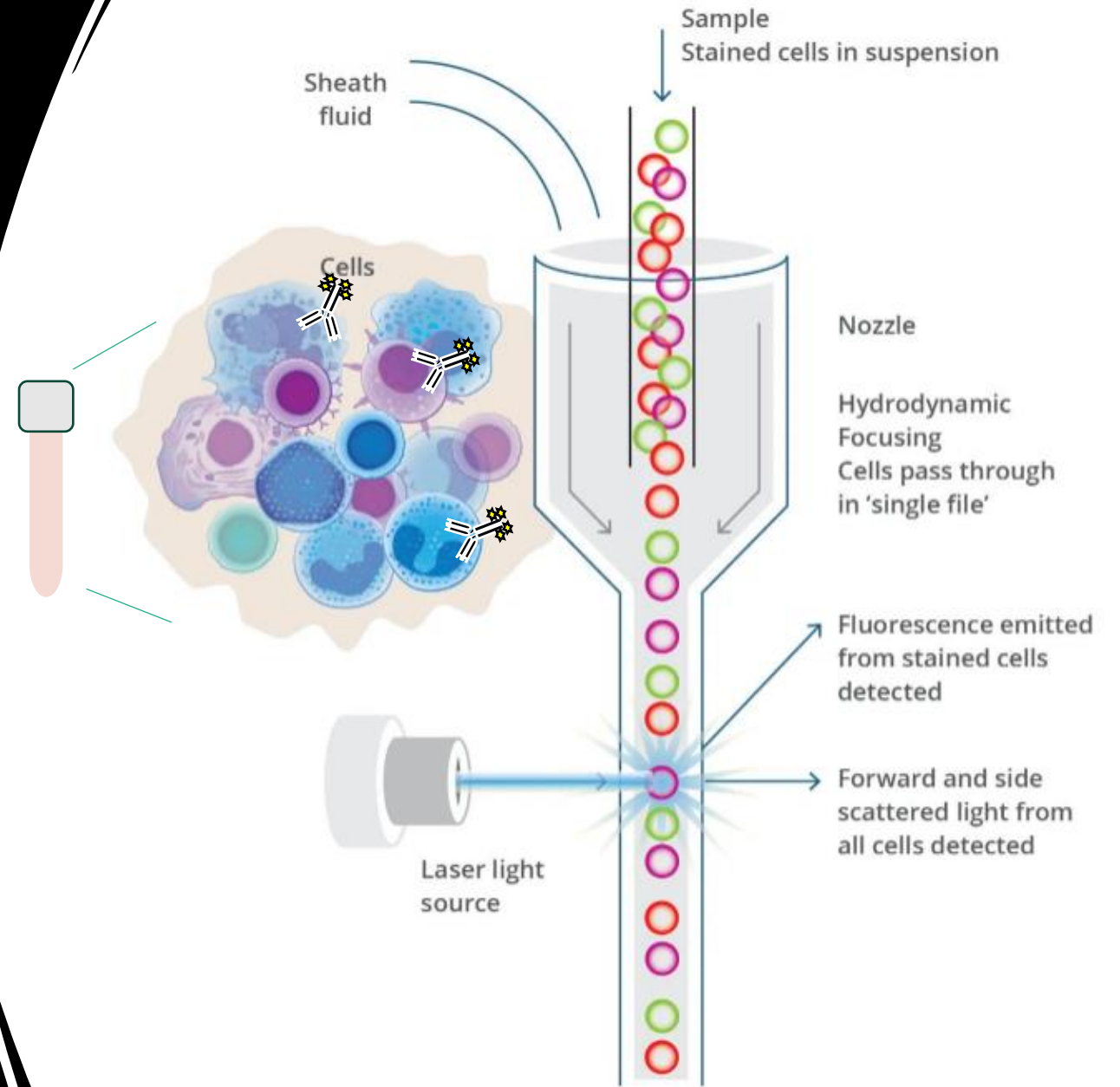


Seeing is Believing

Cytometry- “cell measurement”

What is Flow Cytometry?

- Technology that analyzes the physical and chemical characteristics of particles in **fluid** as it passes through at least one laser.
- Single cell (and beyond) interrogation
- HIGH Throughput (populations)
- Utilizes both light scatter and fluorescence

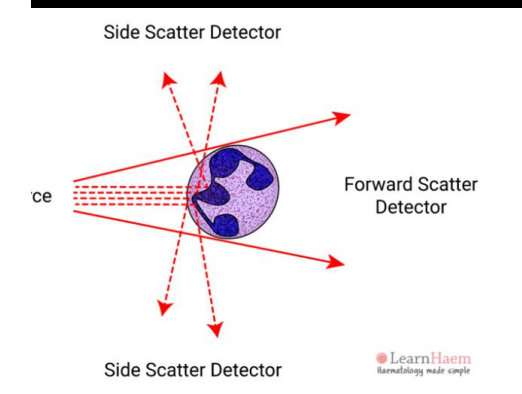
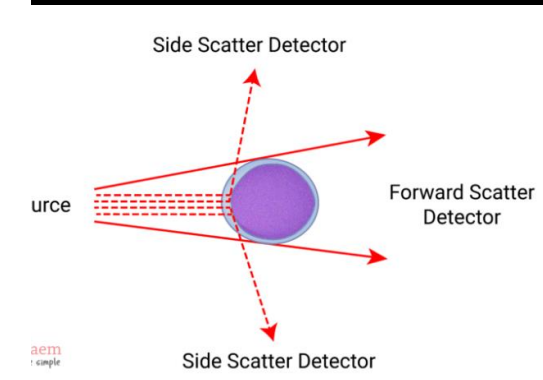
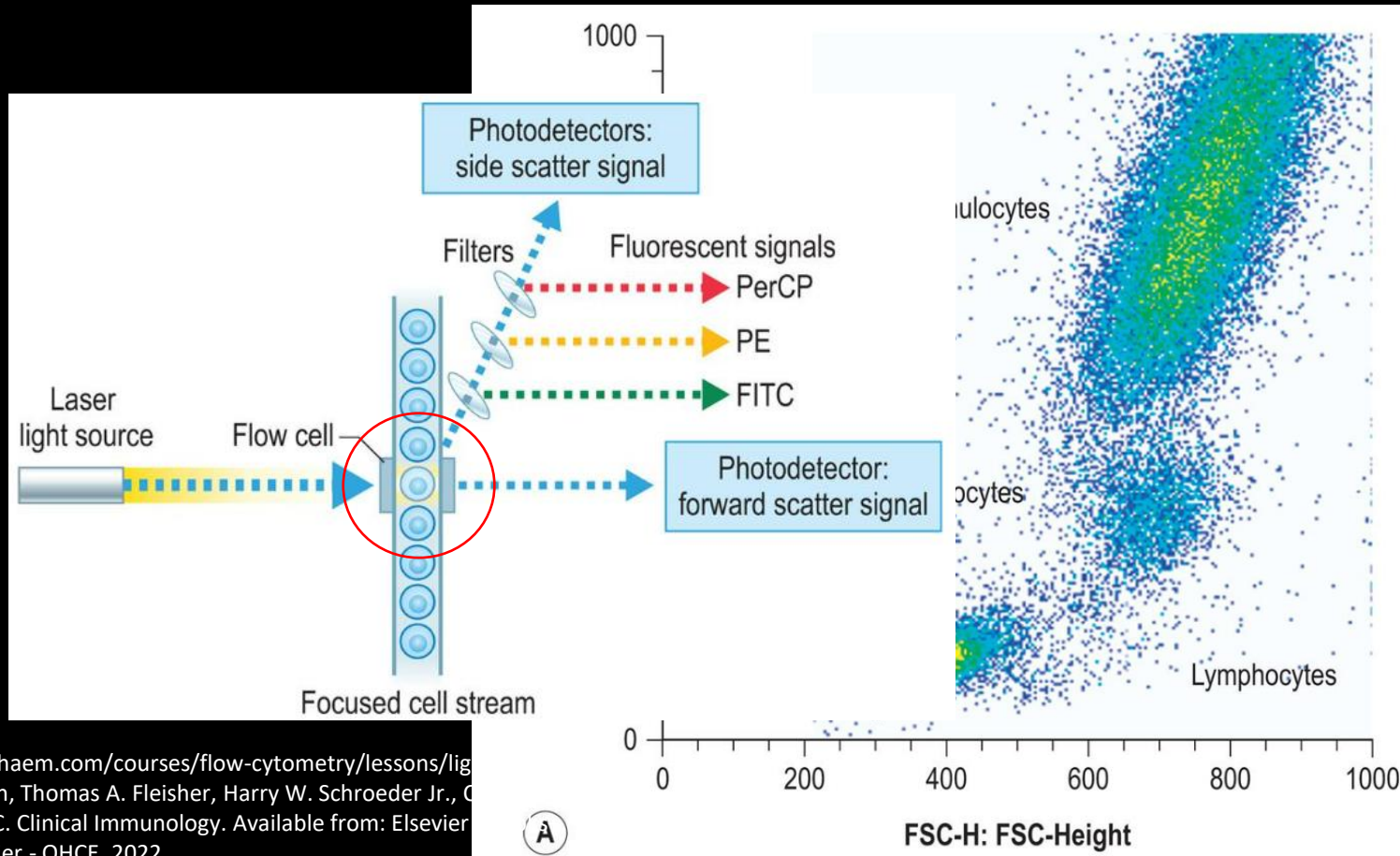


Learning Objectives:

- Review the principles of flow cytometry and examine approaches for successful assay design.
- Discuss various applications of flow cytometry for clinical use.
- Evaluate and apply clinical cytometry techniques to address clinical questions.

Flow Cytometry: The Interrogation Point

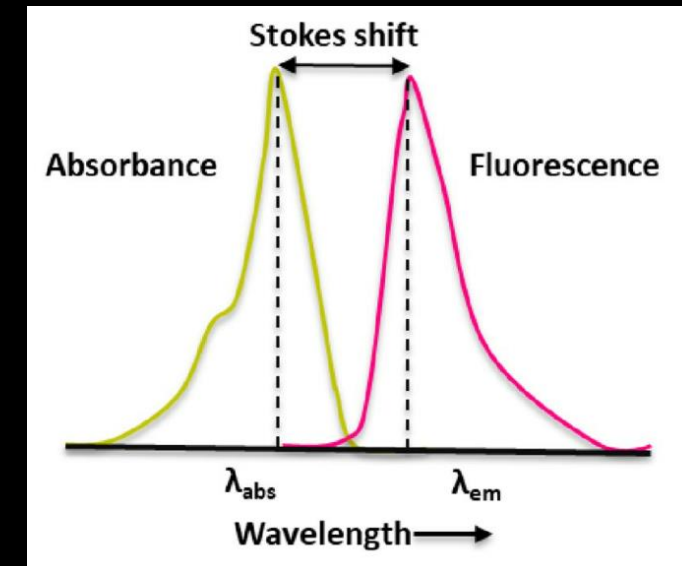
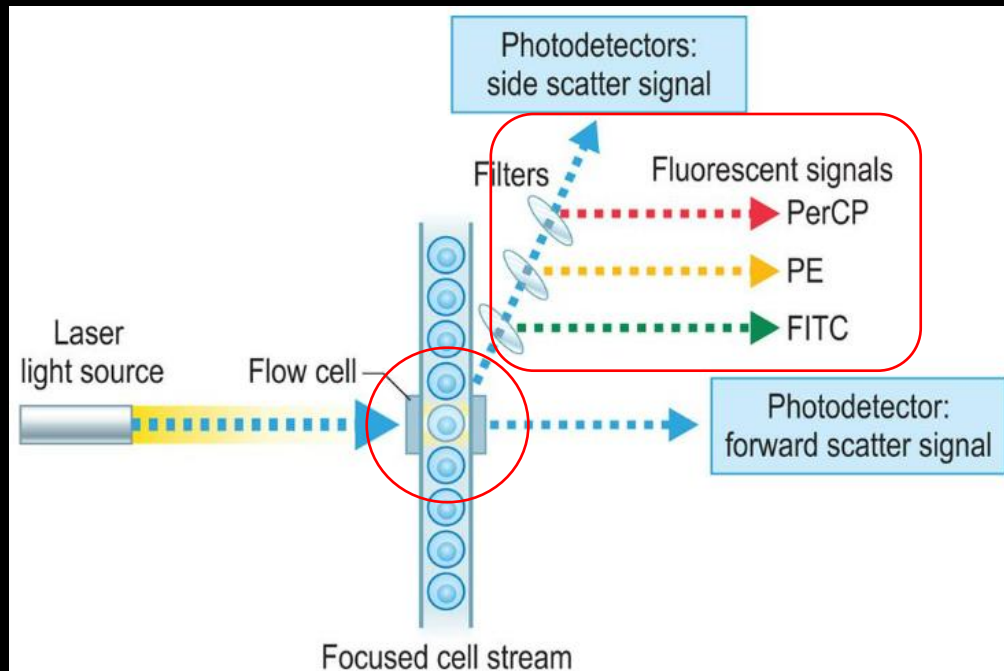
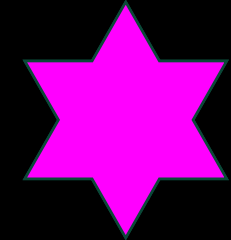
- Light scatter
 - Forward scatter → Rough estimate of cells size
 - Side scatter → Rough estimate of cell “complexity” (granules, organelles, density)
- Together, provide information about cells in our fluid suspension
 - Scatter characteristics differ by population



Flow Cytometry: The Interrogation Point

- Fluorescence
 - Predictable emission wavelength following laser excitation

Fluorophore excited at 633nm



Fluorescence in Flow Cytometry...Its to *Dye* For

- Types of fluorescent reagents
 - “dyes” or fluorescent chemicals → viability, proliferation, DNA content, function etc.
 - Fluorescent proteins → green fluorescent protein, red fluorescent protein etc.
 - Antibodies chemically bound (conjugated) to various fluorophores



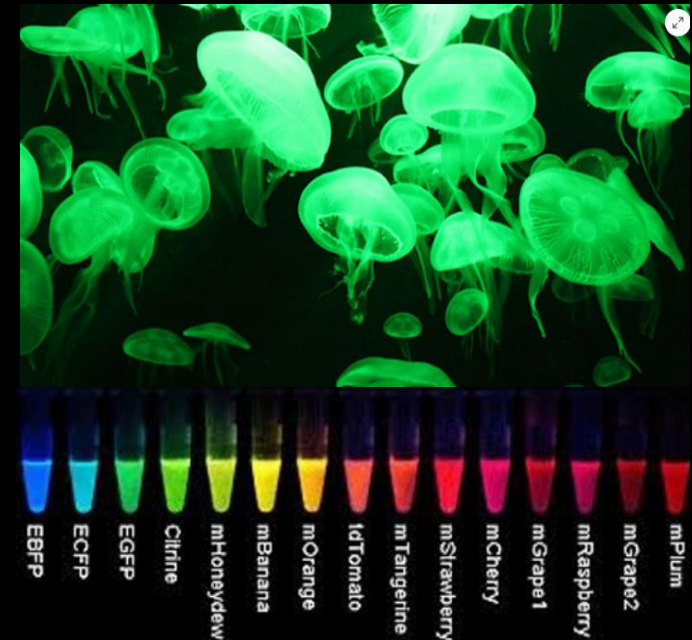
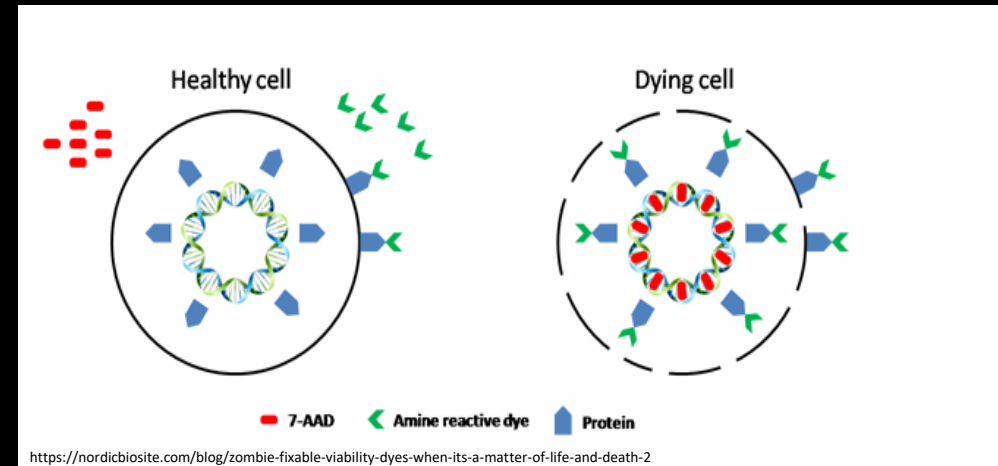
Anti-CD4
FITC



Anti-CD3
APC

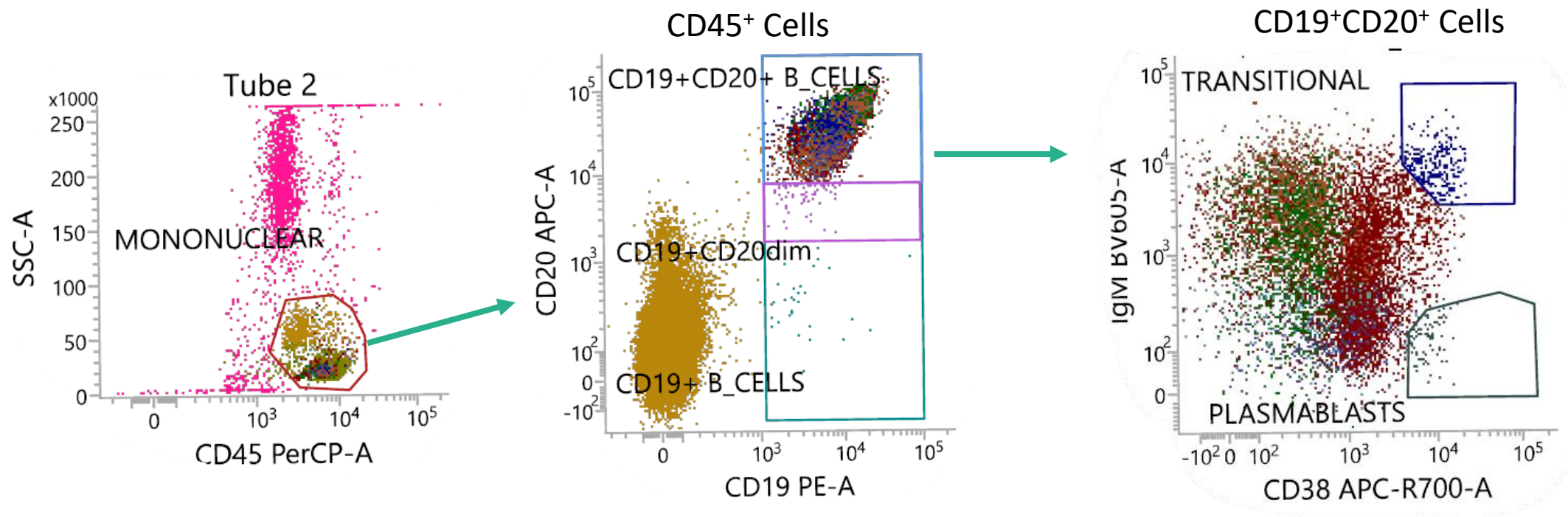


Anti-CD45 PE-
Cy7



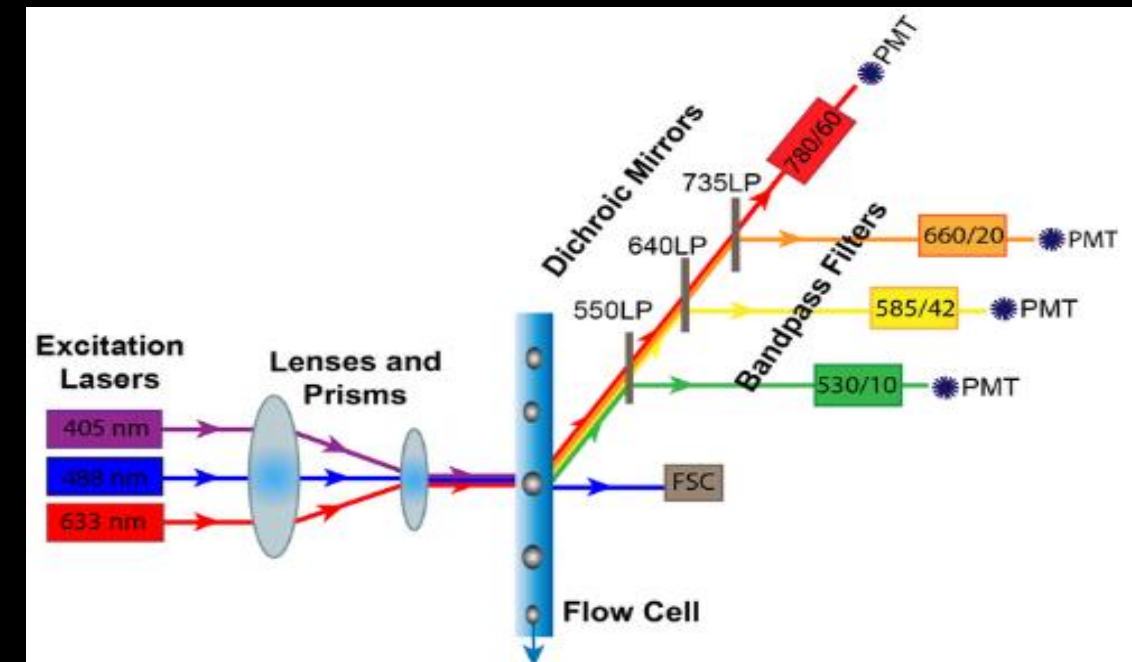
Gating

- Defining Populations of interest using what we know about light scatter and cellular characteristics



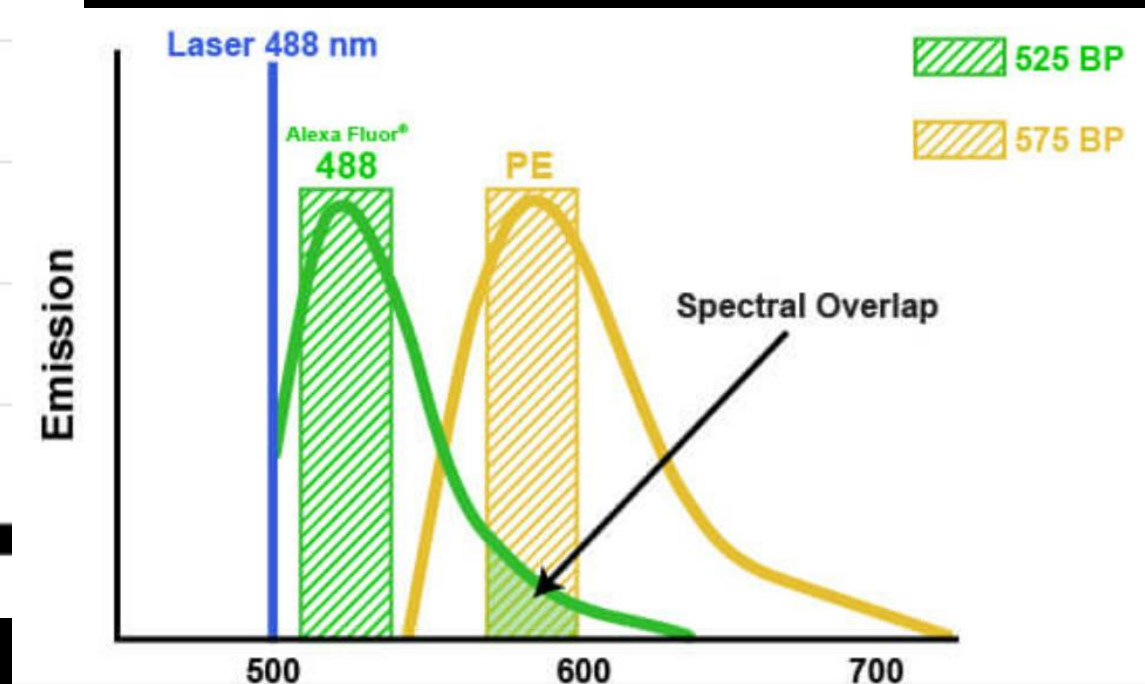
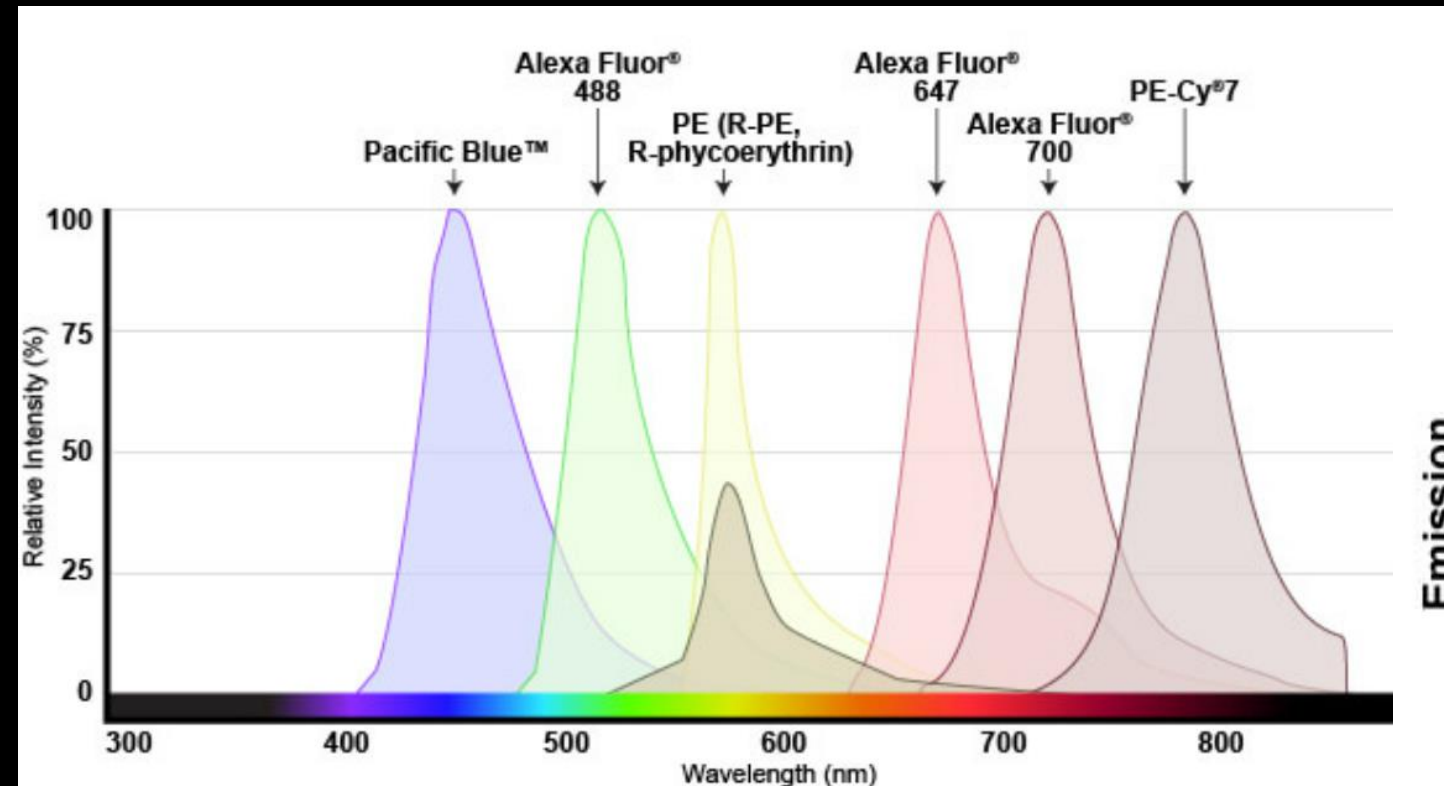
More Lasers = More Colors

- Increasing number of lasers increases the number of fluorophores available for use
 - UV (350nm)
 - Violet (405nm)
 - Blue (488)
 - Green (532nm)
 - Yellow (560nm)
 - Orange (610nm)
 - Red (633nm)
- More colors improve our ability for multiparameter analysis (many things at once)



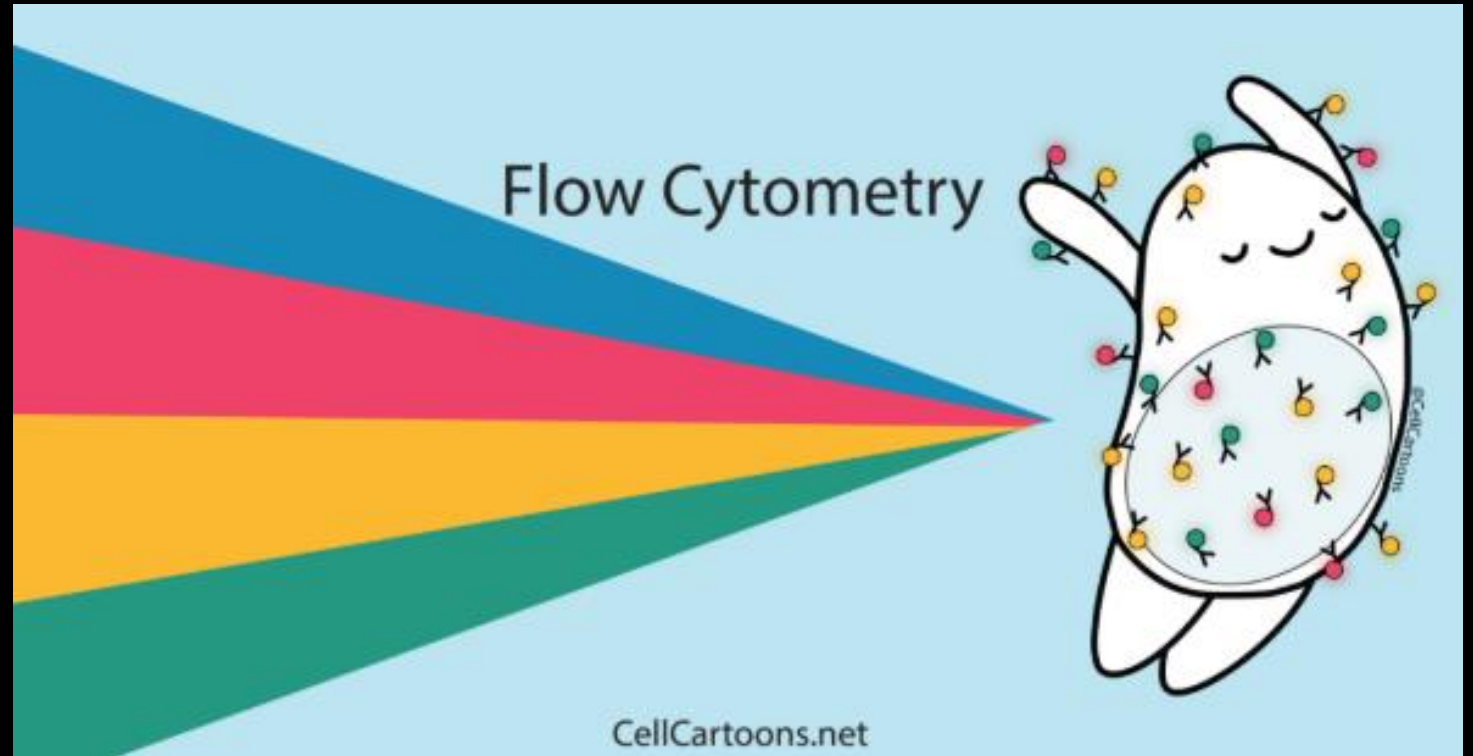
More Colors, More Problems

- Increasing the number of fluorophores, increases panel complexity
 - Unique emission spectra
 - Overlap (spillover) increases as number of fluorophores increases



Advantages of Flow Cytometric Analysis

- Multiparameter analysis → observe many things at once
 - Populations, subpopulations and single cells
 - Plethora of staining reagents available
 - Extracellular and intracellular investigation possible
- Fixed or live cell analysis
- High throughput platform
- Highly sensitive and specific



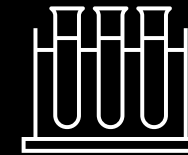
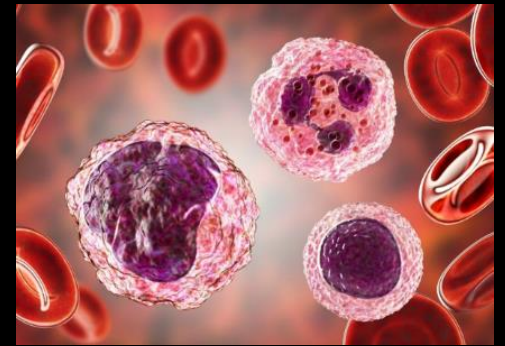
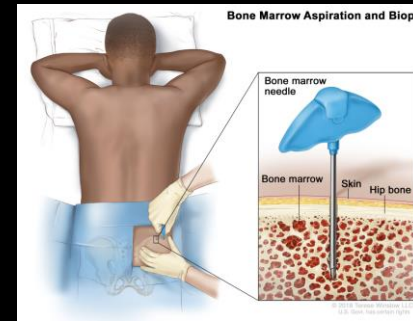
Factors Affecting Panel Design

WHAT: What exactly is the goal → informs what to target

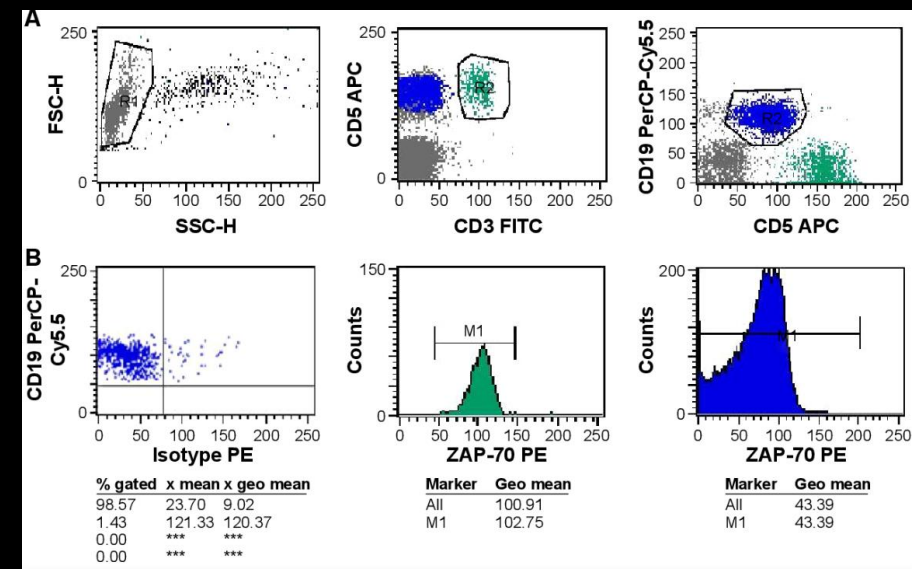
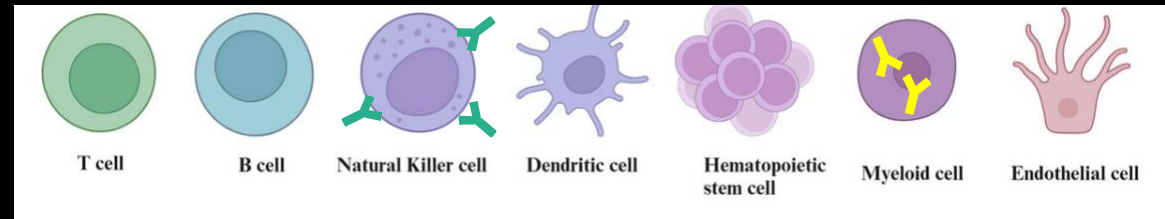
WHERE: Intracellular, extracellular

HOW: Fixation vs live? Permeabilization? Quantitative vs qualitative?

READ OUT: How will data be analyzed to interpret results? Quantitative vs Qualitative assays?



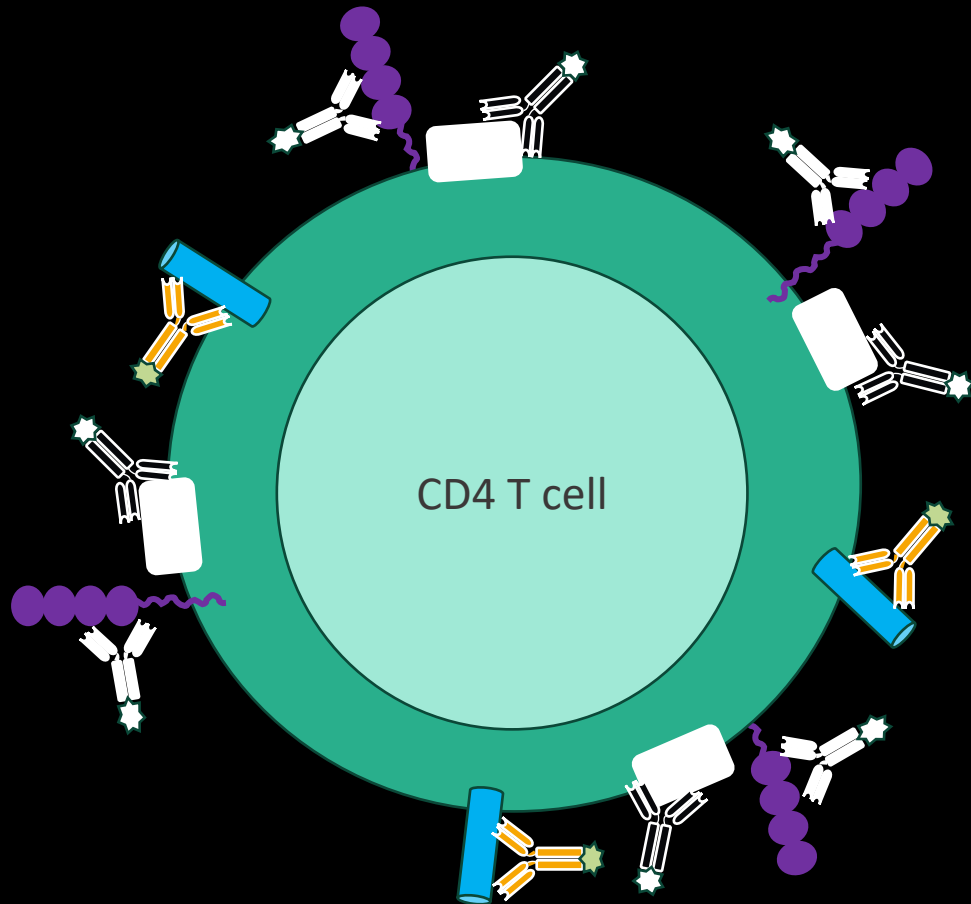
Sample Type



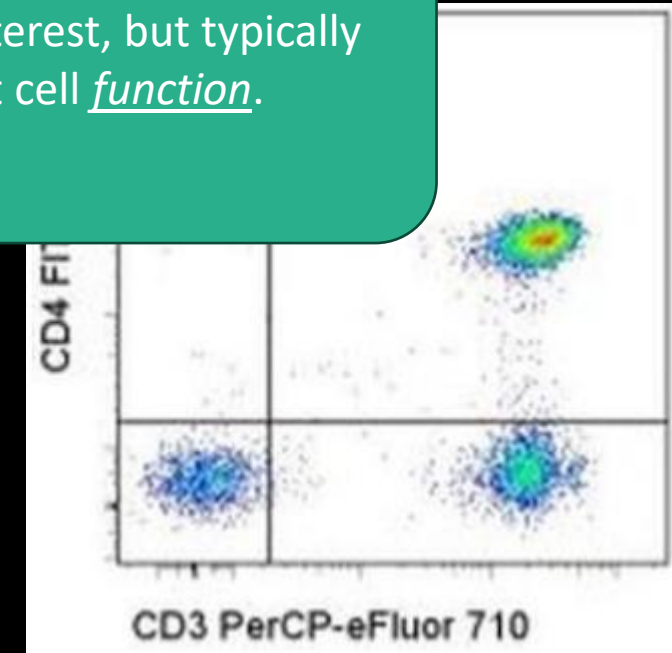
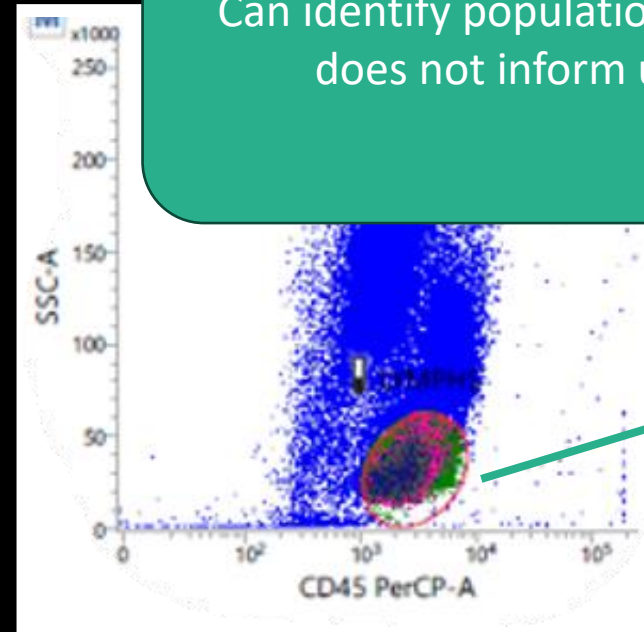
<https://nci-media.cancer.gov/pdg/media/images/554337.jpg>; <https://www.the-scientist.com/accelerating-immune-research-with-cryopreserved-peripheral-blood-mononuclear-cells-70242>; <https://www.assaygenie.com/blog/immunophenotyping-by-flow-cytometry/>;

Wu, Yu-Jie et al. Using the geometric mean fluorescence intensity index method to measure ZAP-70 expression in patients with chronic lymphocytic leukemia. *OncoTargets and therapy* 9 (2016): 797 - 805.

Extracellular Panel Approaches



Can identify populations of interest, but typically does not inform us about cell function.



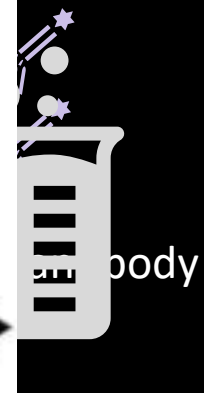
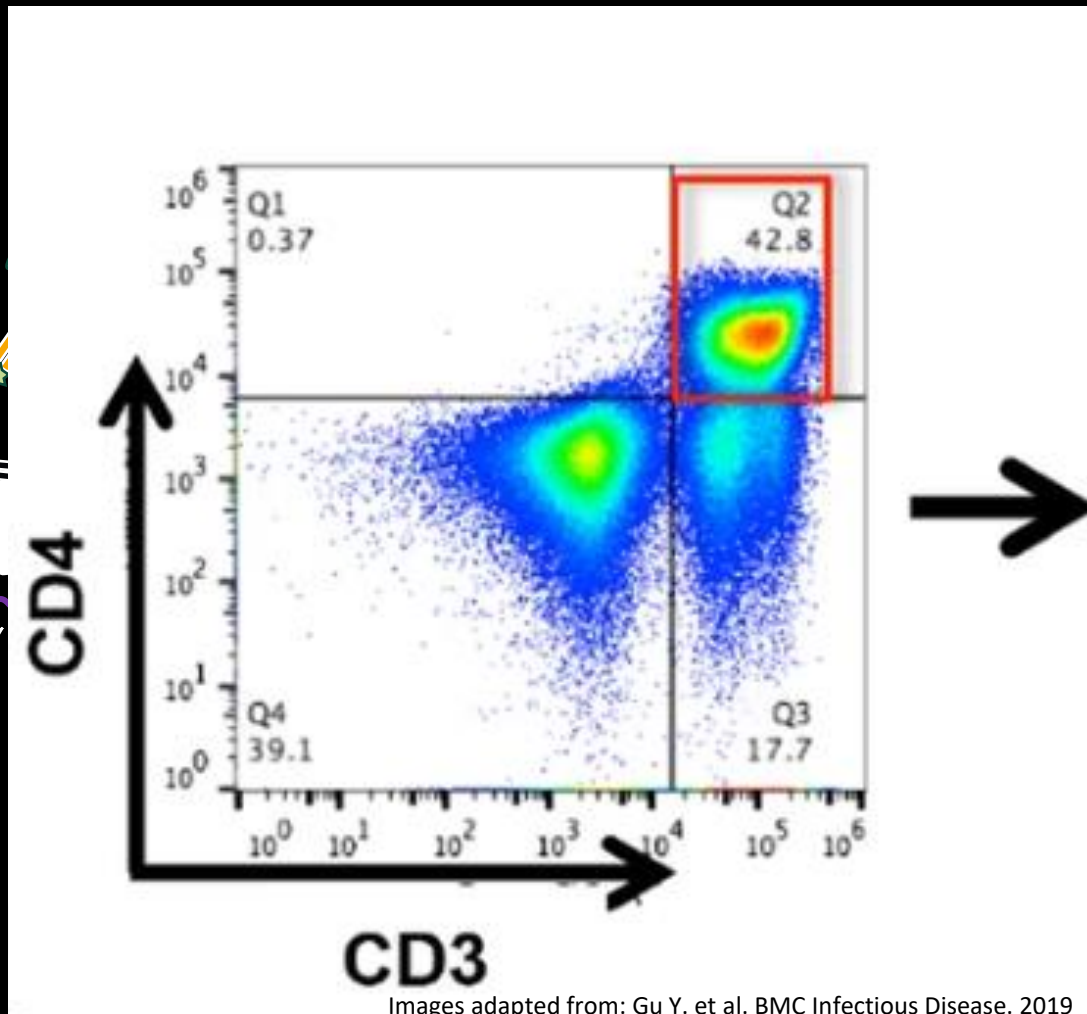
<https://www.thermofisher.com/antibody/product/CD3-CD4-CD8a-Antibody-clone-SK3-SK7-SK1-Cocktail/22-0306-71>

CD45-pan leukocyte marker

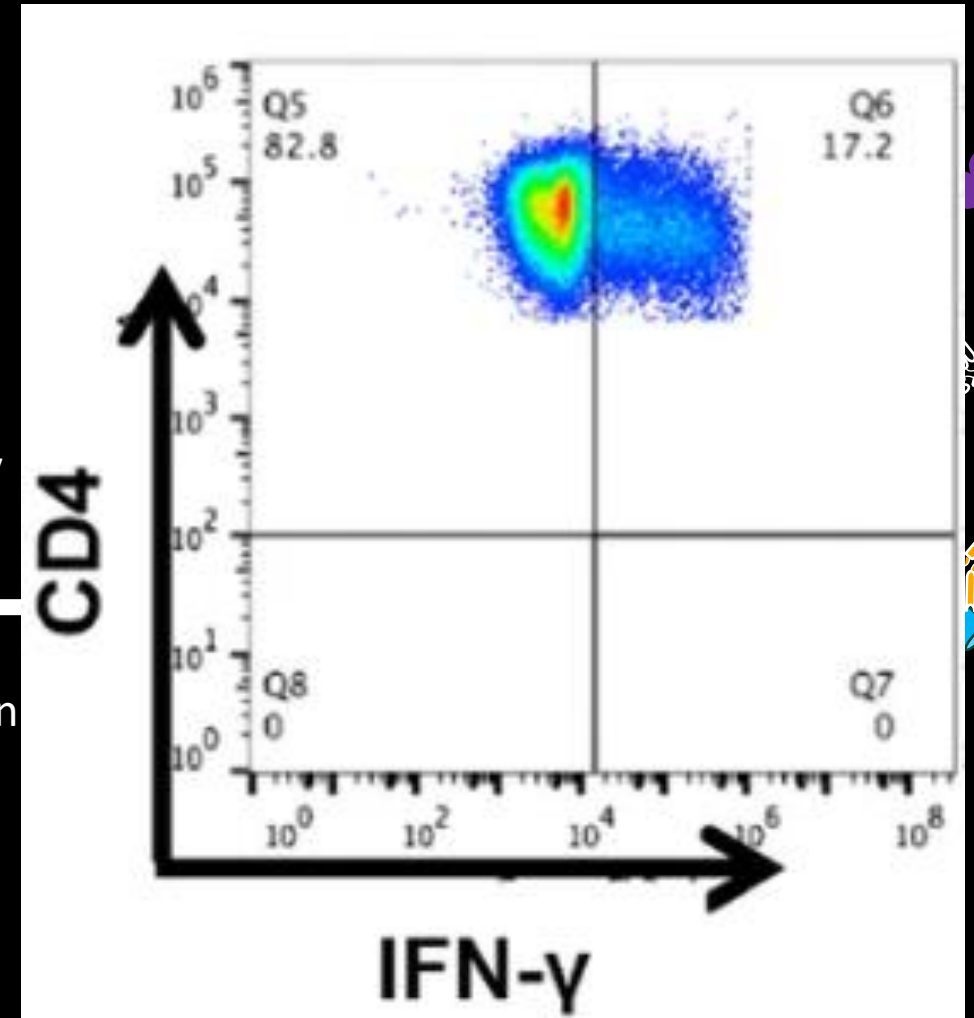
CD4-T subset-specific co-receptor

CD3-T cell signaling molecule (pan T cell marker)

Panel Design: Intracellular Targets



tion and
stabilization
(ethanol or
propen)



CD45-pan leukocyte marker

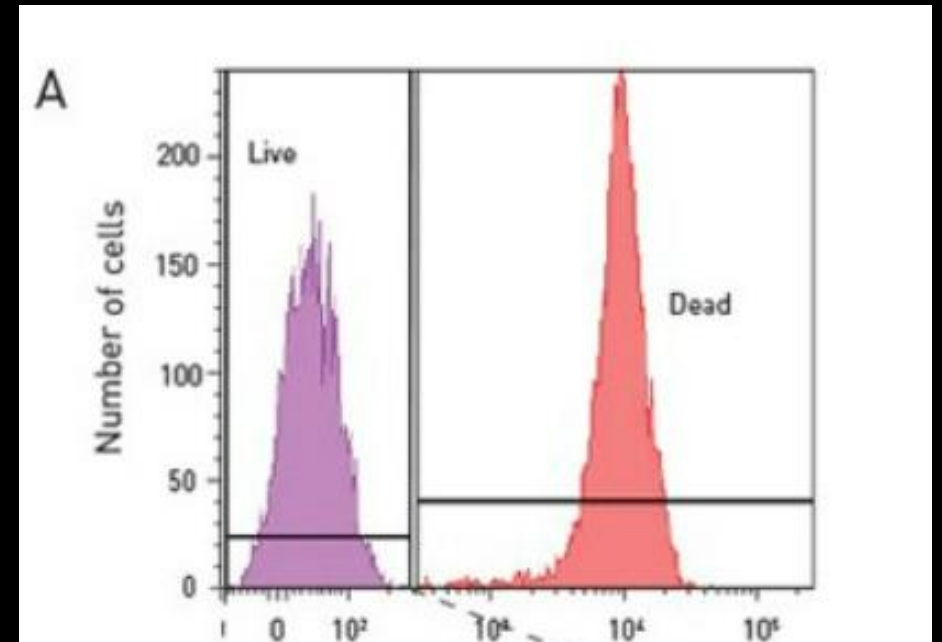
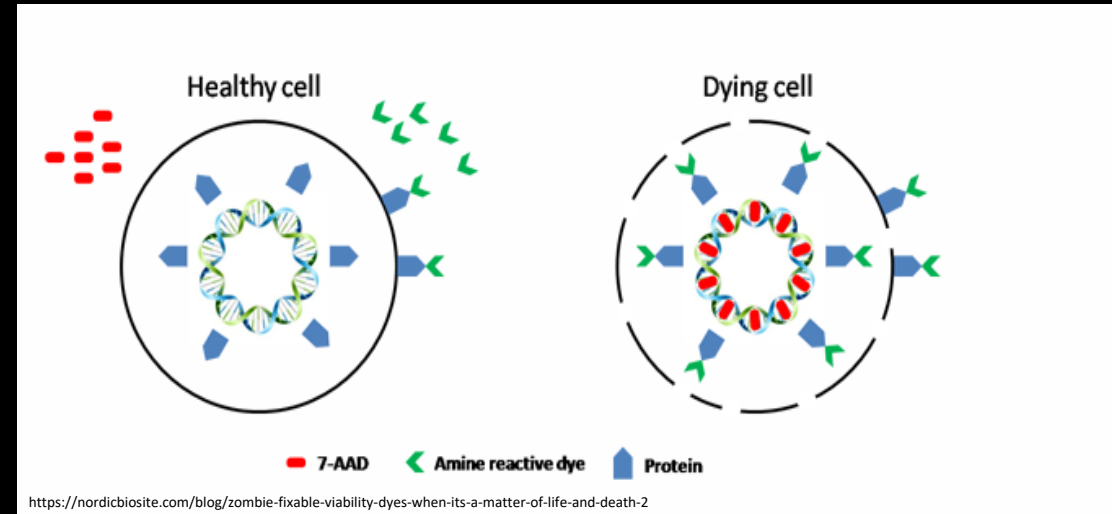
CD4-T subset-specific co-
receptor

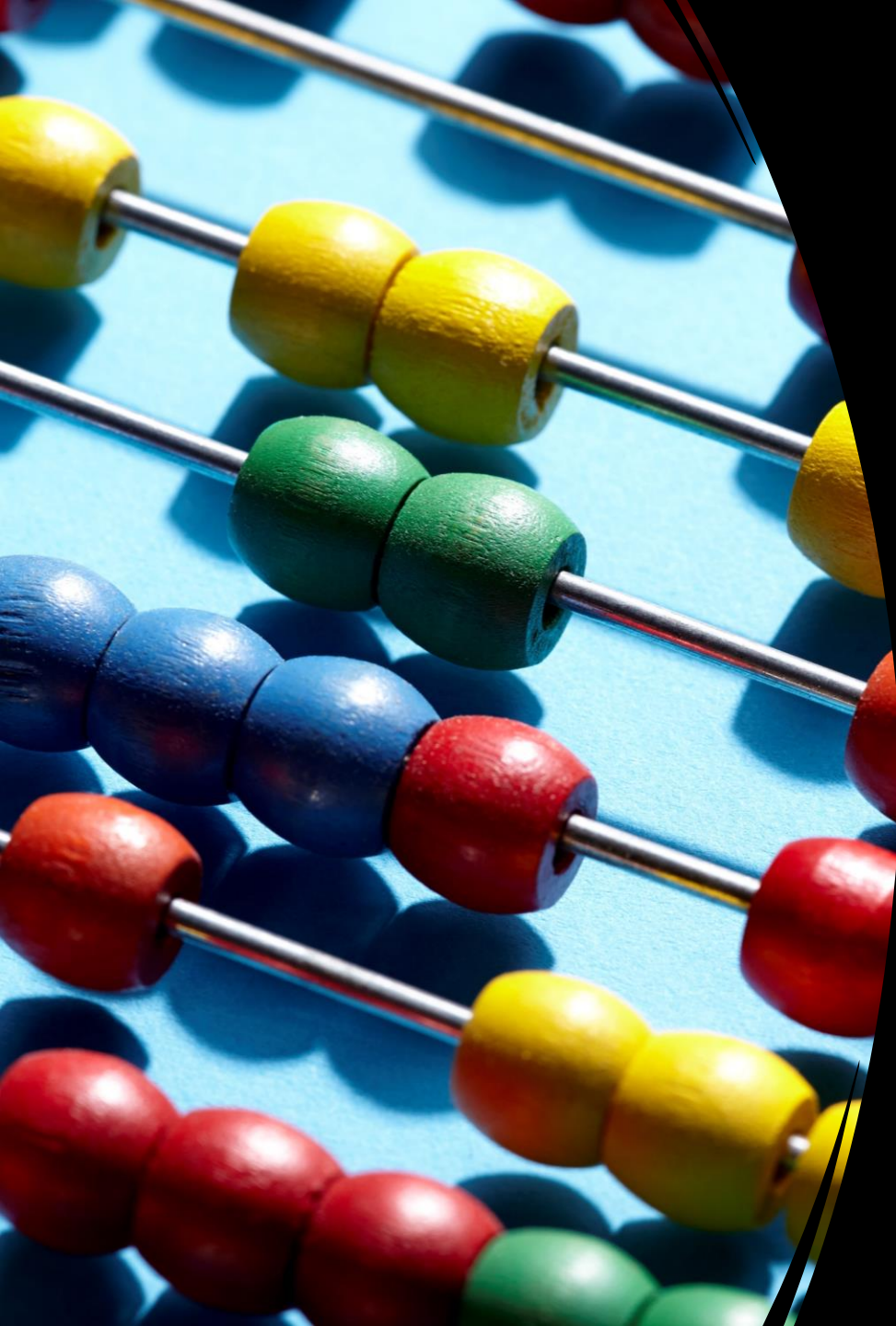
CD3-T cell signaling molecule
(pan T cell marker)

IFN- γ (cytokine)

Panel Design: Intracellular Targets

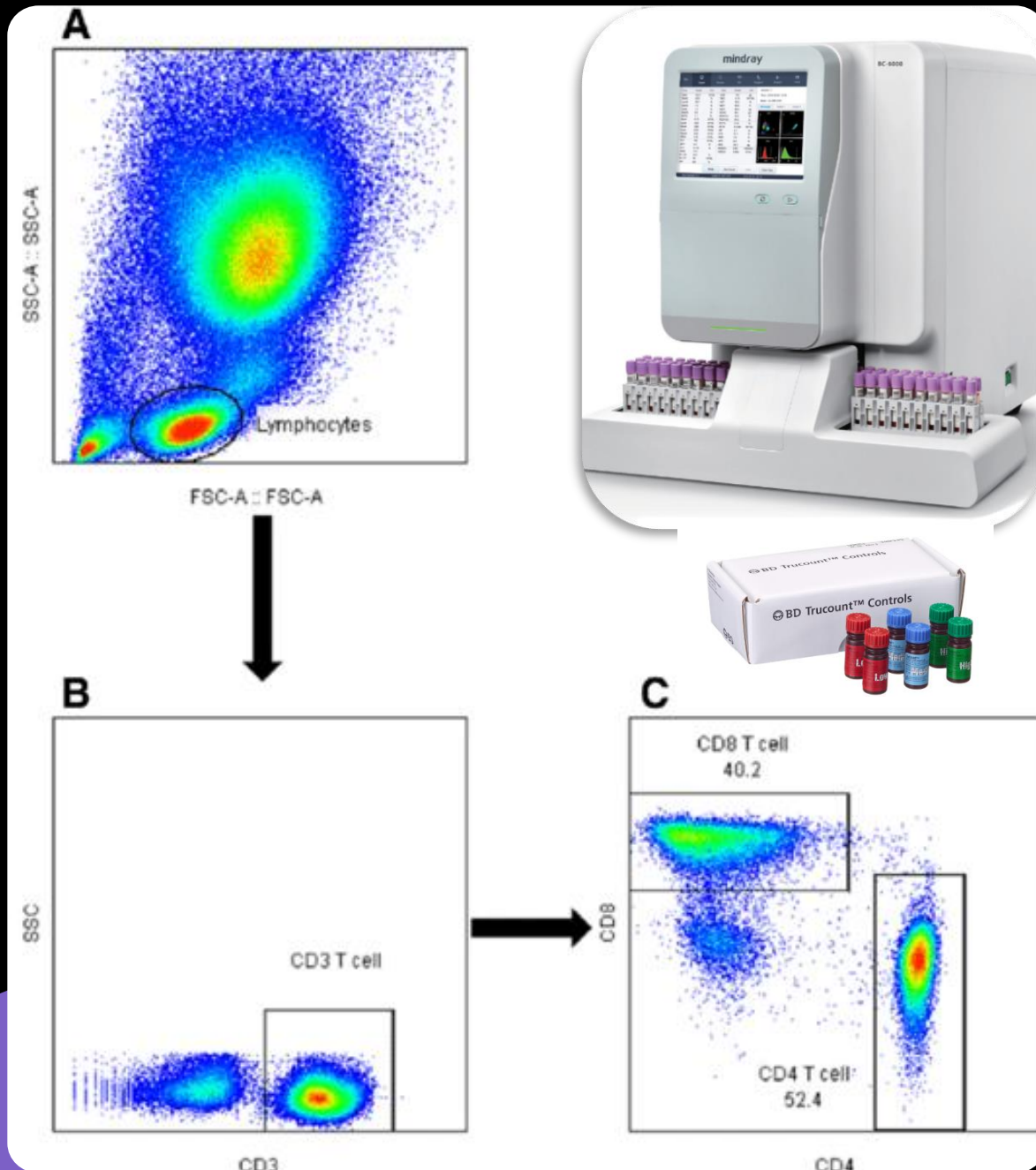
- Viability-DNA binding or amine reactive dyes
- Proliferation-DNA binding, pyrimidine analogs,
- Other-transcription factors, phosphorylated proteins, , chemical changes etc.





Who's There and How Much? Quantitative and Qualitative Flow Cytometry

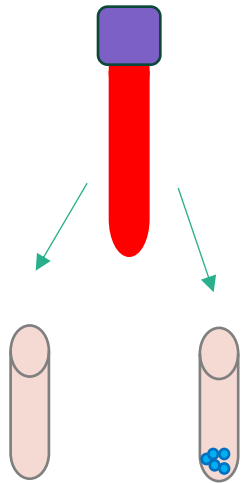
- How is the assay reported?
 - Quantitative flow assays: reports out a quantitative result value (i.e. cells/ μ L)
 - Qualitative flow assays: detect the presence or absence of cell population where reported value is based on a relative frequency (i.e. % of CD45⁺ lymphocytes)



Quantitative Flow Cytometry

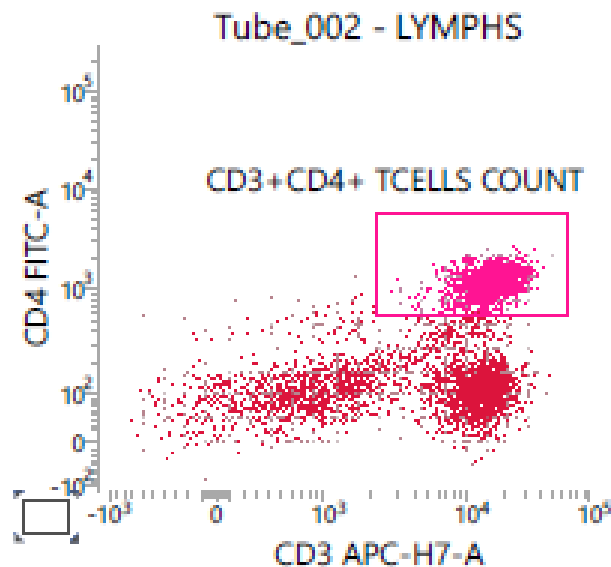
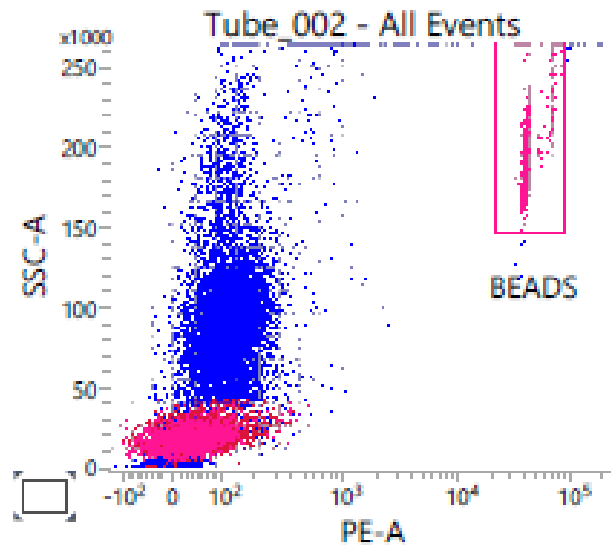
- Flow cytometric analysis can measure the frequency of cell subsets within the sample.
- Using this relative percentage, absolute cell counts for subpopulations can be back calculated from absolute cell counts or reference material (known quantity of fluorescent beads)

Patient sample



Tube 1: aliquot of patient sample stained with all markers of interest

Tube 2: aliquot of patient sample stained with lineage markers, and combined with known quantity of fluorescent beads

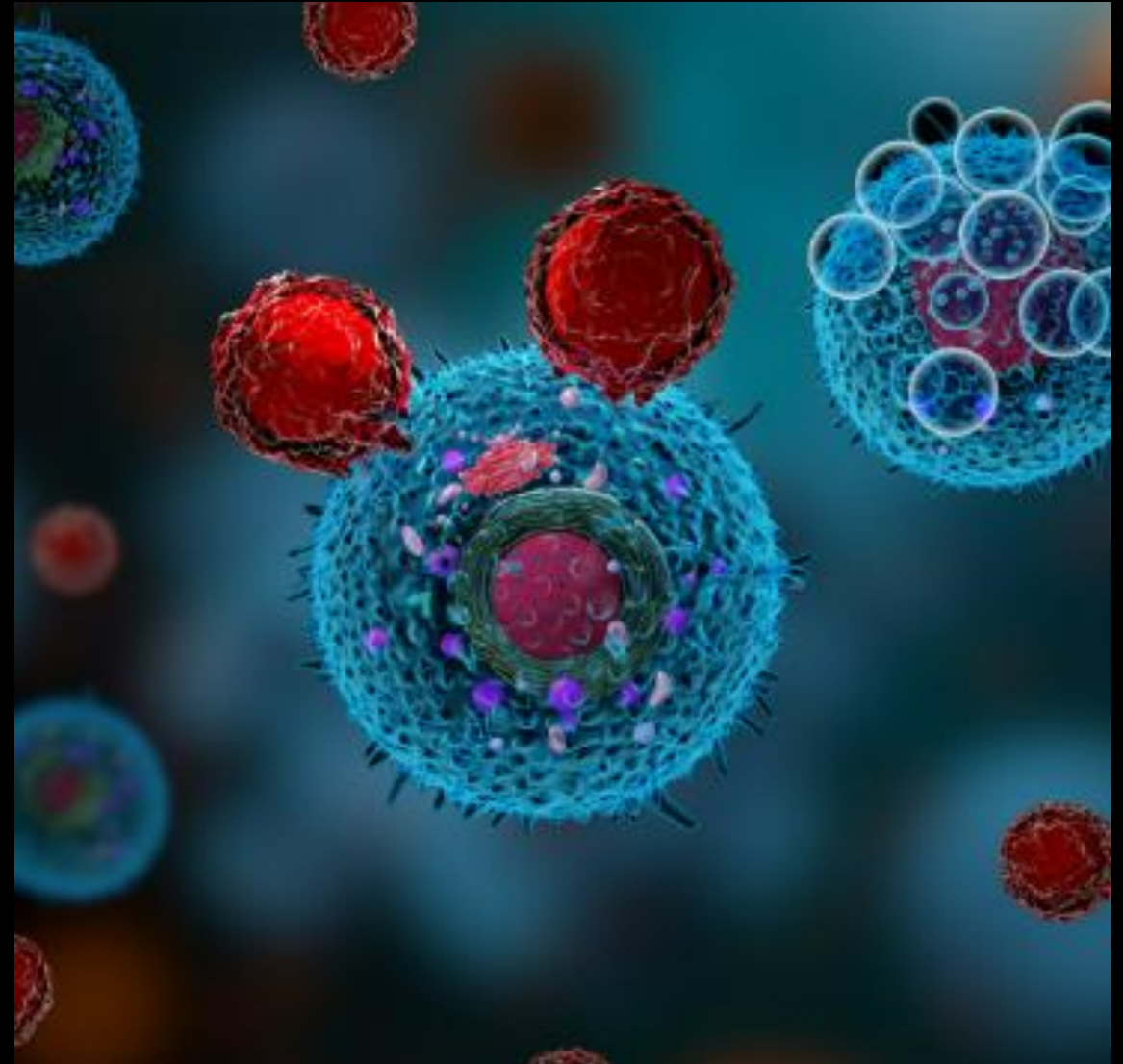


Quantitative Flow Cytometry-HIV Monitoring

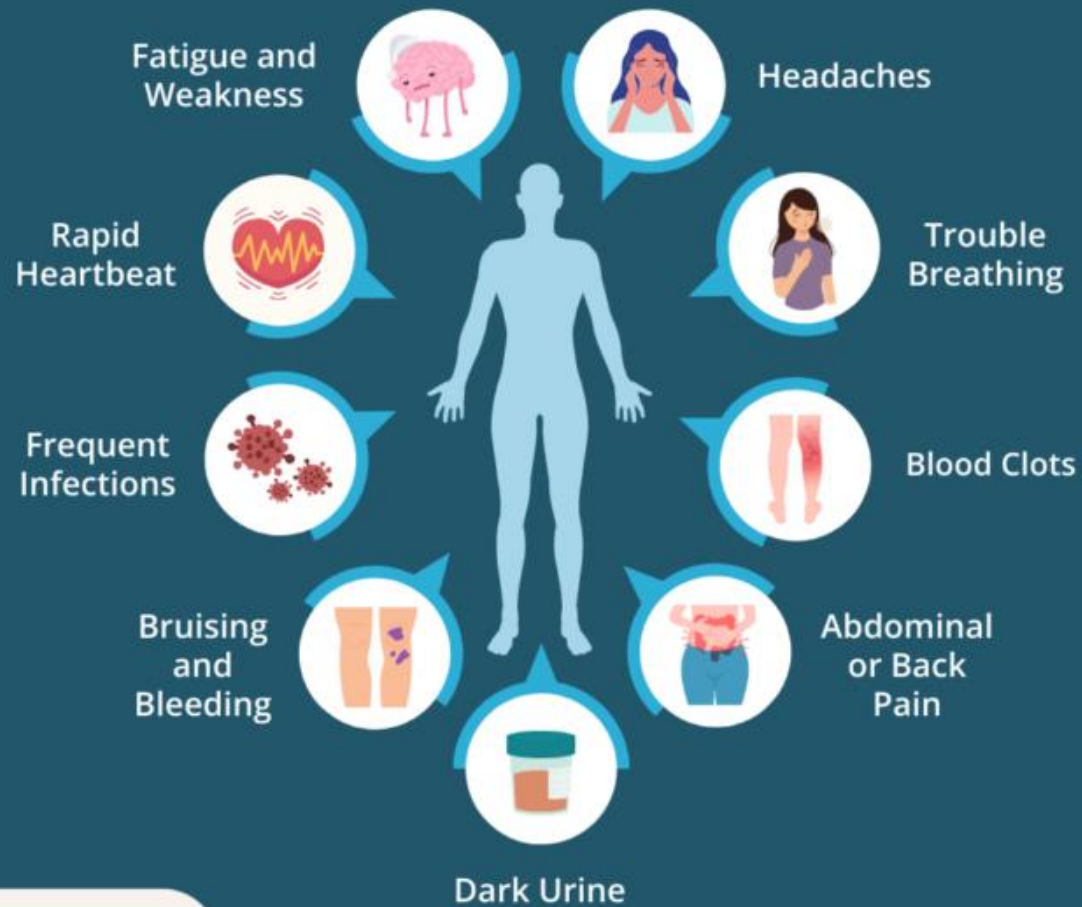
- Percentage and total number of lymphocytes can often aid in informing next diagnostic steps
- Enumeration of CD4 T cells and HIV
 - CD4 T cell counts ≤ 200 cells/ μ L is AIDS-defining event
 - Useful for monitoring disease progression, and prevention of opportunistic infection

Qualitative Clinical Flow Cytometry

- Assessing the presence or *absence* of...
 - Cell population(s)
 - Target(s) of interest on a cell population
 - Expression level of targets of interest (how much?)
- Often useful for diagnosis/support of diagnosis



How PNH Affects the Body



EVERYDAY HEALTH

Paroxysmal Nocturnal Hemoglobinuria (PNH)

- Rare blood disorder characterized by a *reduction or absence* of glycosyl phosphatidylinositol (GPI)-anchored membrane proteins
 - Lack of GPI expression on red blood cells (RBCs) and white blood cells (WBCs) leads to increased susceptibility to complement activity → increased cell lysis

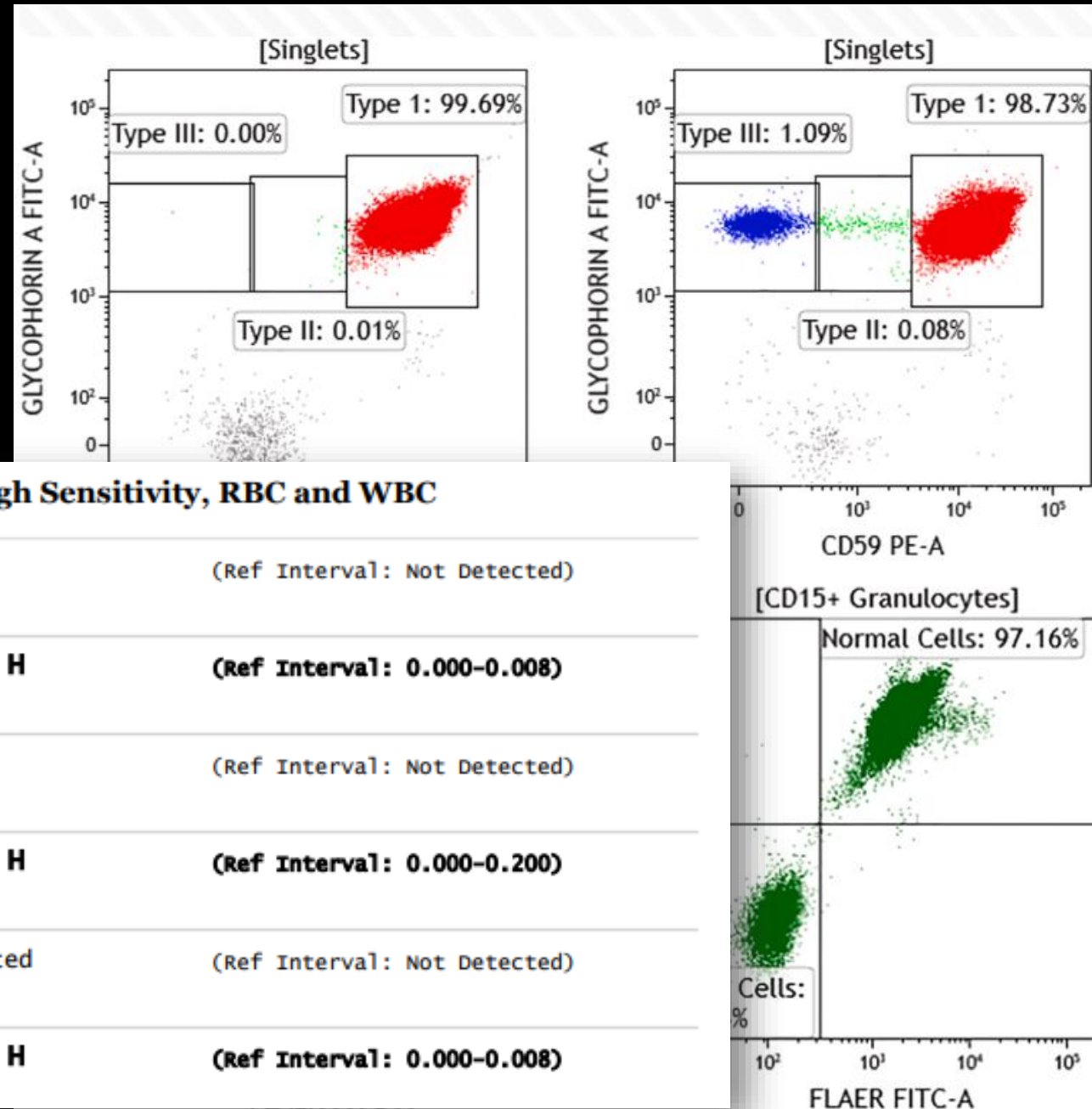


PNH: Detecting Deficiency

- Flow cytometric analysis of GPI-anchored proteins on RBCs and WBCs is the gold standard for diagnosis and monitoring of disease
- Purpose of the assay demands special considerations
 - Tasked with measuring what *isn't* there
 - Limited number of events detected within a given population → Rare event analysis
 - Minor PNH clones <1%-4% of total cells
 - PNH clones are sensitive to destruction

Detecting PNH Clones with Flow Cytometry

- Sufficient sensitivity → Increase events (≥100,000)
- Assess both RBC and WBC for presence of GPI-anchored proteins

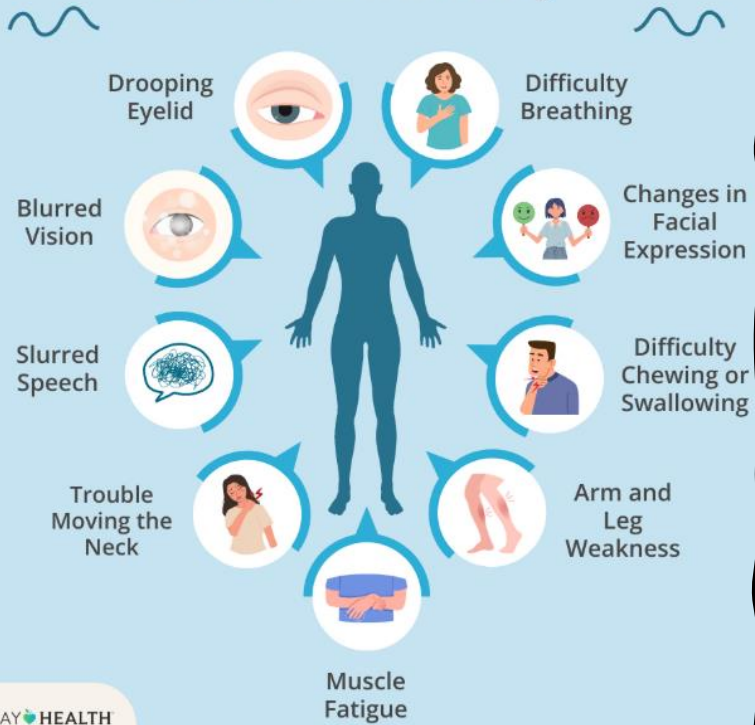




Form \neq Function: Investigating Cell Function Using Flow Cytometry

- Presence of a particular marker can indicate cell phenotype, lineage, and suggest function, but the presence of a marker alone does not necessarily indicate *functionality*
- Flow cytometric techniques can still be leveraged to investigate cell function, with clever assay design
 - Read out for function: phosphorylation, proliferation, cytokine production, specific protein regulation etc.
- Powerful strategy for investigating functional immune responses

How Myasthenia Gravis Affects the Body



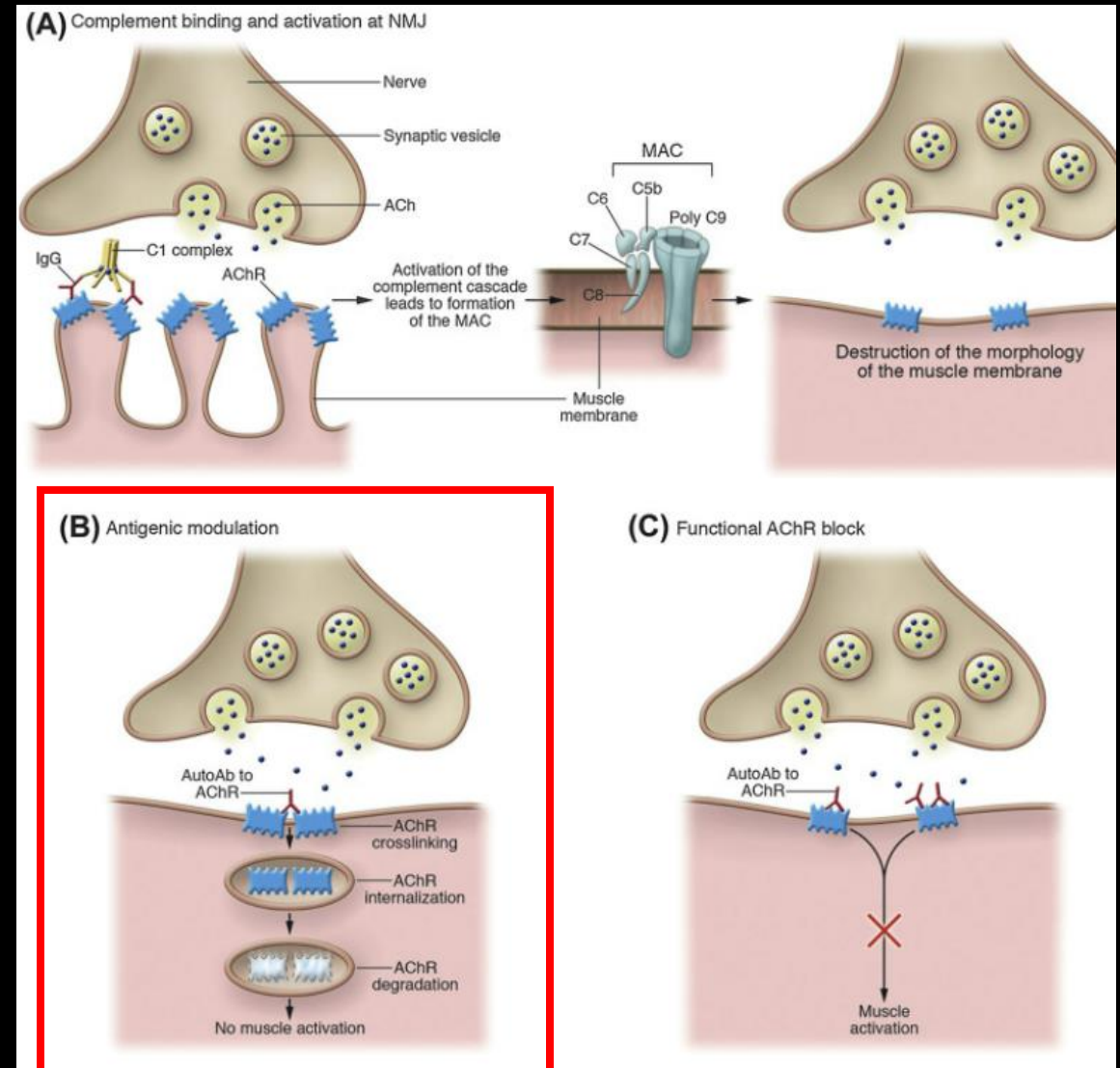
Functional Assays: Myasthenia Gravis Modulating Autoantibody Detection

- Myasthenia Gravis- autoimmune disorder in which autoantibodies are generated against the neuromuscular acetylcholine receptors (AChR)
 - Disruption of neuromuscular transmissions → voluntary muscle weakness and fatigue
 - Fluctuating weakness that worsens with activity, improves with rest

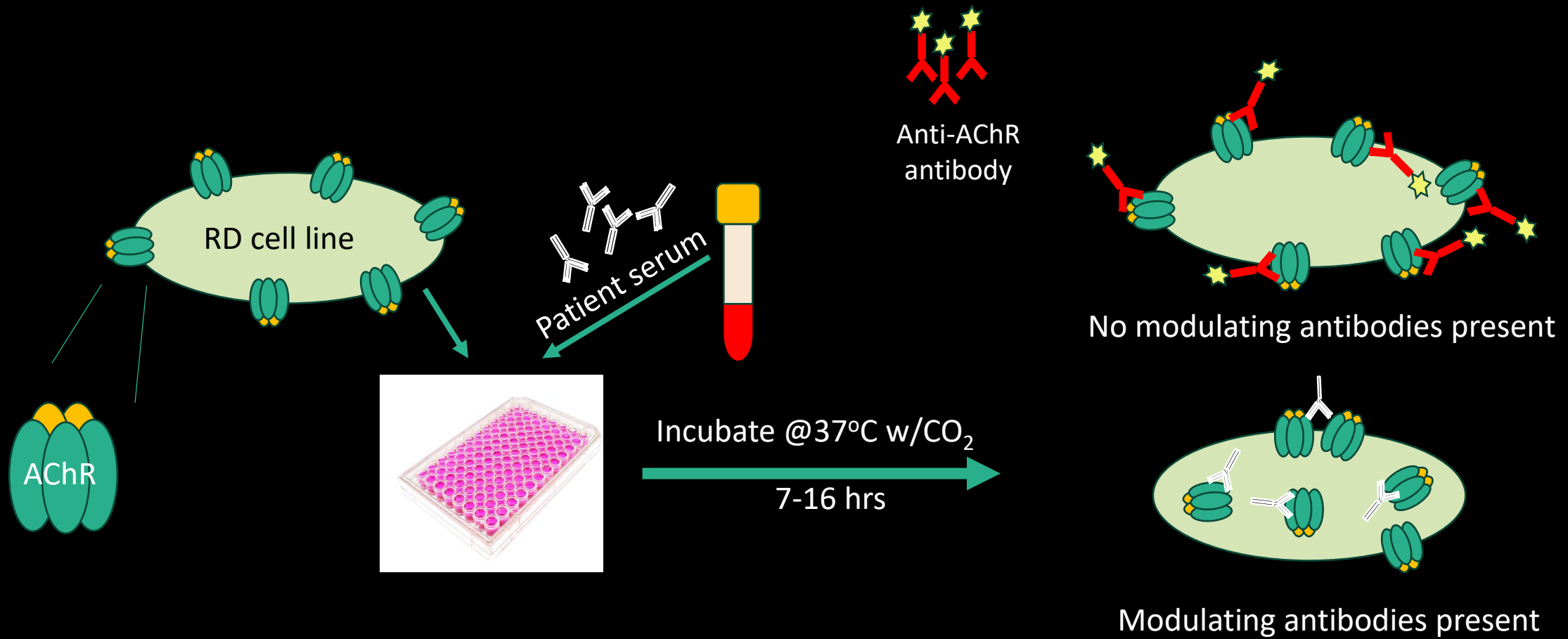


Functional Assays: Myasthenia Gravis Modulating Autoantibody Detection

- Myasthenia gravis diagnosis is often based on correlating clinical presentation with laboratory confirmation of autoantibody against AChR
- Additional characterization of autoantibody may aid in characterization and prognosis of disease
 - Modulating antibodies are correlated with enhanced disease severity

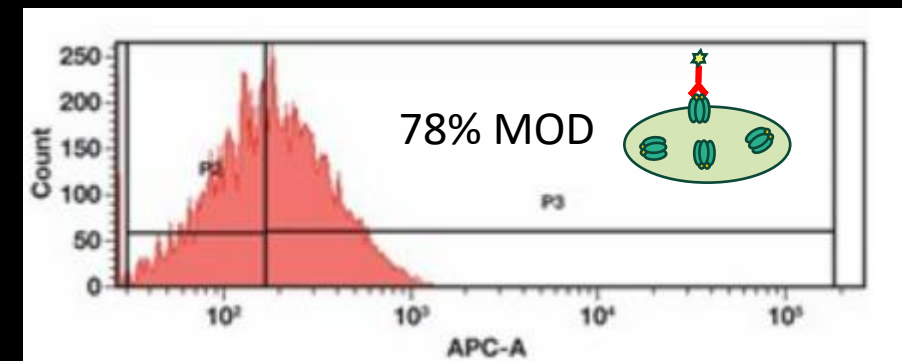
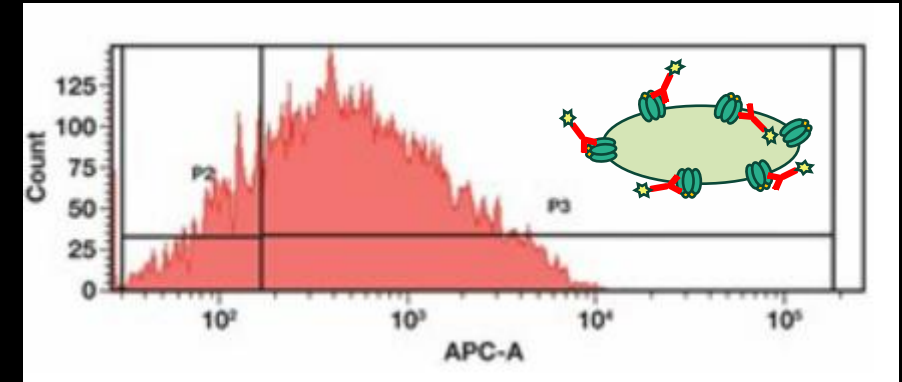
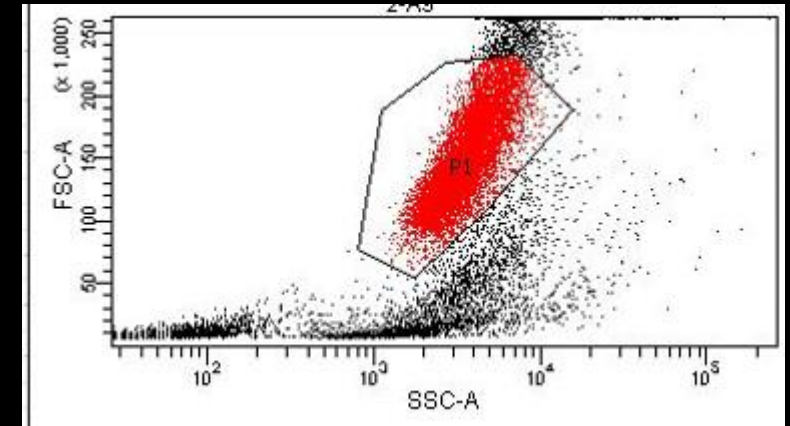


Linda L. Kusner, Henry J. Kaminski, in *Neurobiology of Brain Disorders (Second Edition)*, 2023



Functional Assays: Myasthenia Gravis Modulating Autoantibody Detection

- Fluorescent signal intensity is used to calculate %Modulation
- Compare intensity of signal to maximum calibrator (no autoantibody) and negative controls



Reference Interval		
0-45% modulation	=	Negative
≥46% modulation	=	Positive

Clinical Applications of Flow Cytometry

Functional

- Lymphocyte proliferation
- Neutrophil Oxidative Burst
- Leukocyte Adhesion Deficiency
- BTK Phosphorylation

Quantitative

- CD34⁺ Hematopoietic Stem Cell (sorting) Enumeration
- CD4⁺ T cell Enumeration
- Immunophenotyping
- Cell Cycle Analysis

Presence/Absence

- Primary Immunodeficiency
- Malignancy
- Hematologic Disorder

Rare Event Analysis

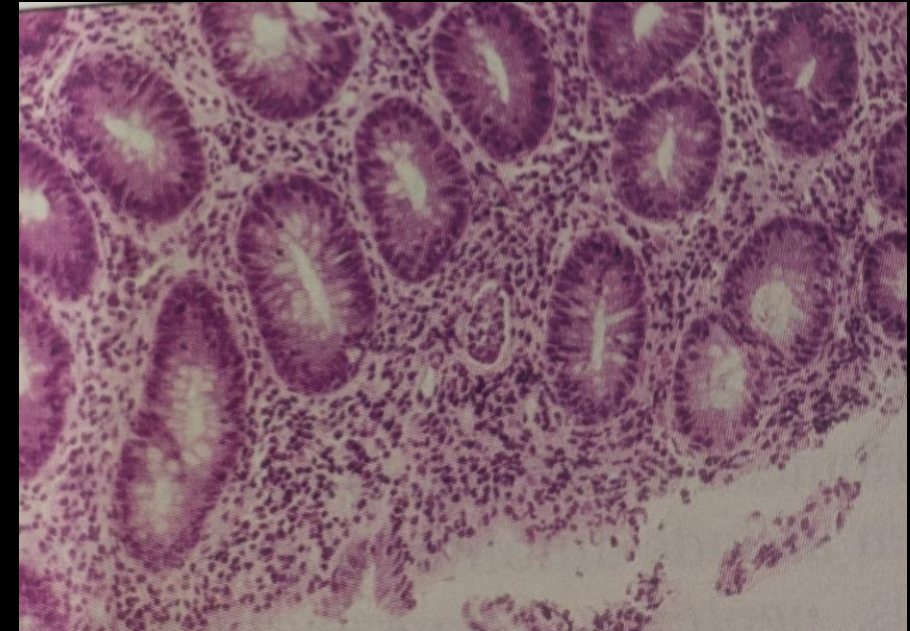
- Minimal Residual Disease
- Paroxysmal Nocturnal Hemoglobinuria

Case 1: Infantile Onset Type 1 Diabetes

- 4 mo old male presents to the clinic with an intractable diarrhea and failure to thrive. Antibiotic treatment was administered, but watery stools persisted.
- At 6 mo old, he developed high glucose levels and glucose in the urine.
 - Diagnosed with Type 1 Diabetes and referred to endocrinology.
- Further work-up revealed enlarged lymph nodes and spleen, but his white blood cell count (7300 / μ l), hemoglobin (11.3 g/dl), and platelet count (435,000/ μ l) were normal.
- Autoantibodies to both glutamic acid decarboxylase (GAD65 antigen) and pancreatic islet cells were detected.

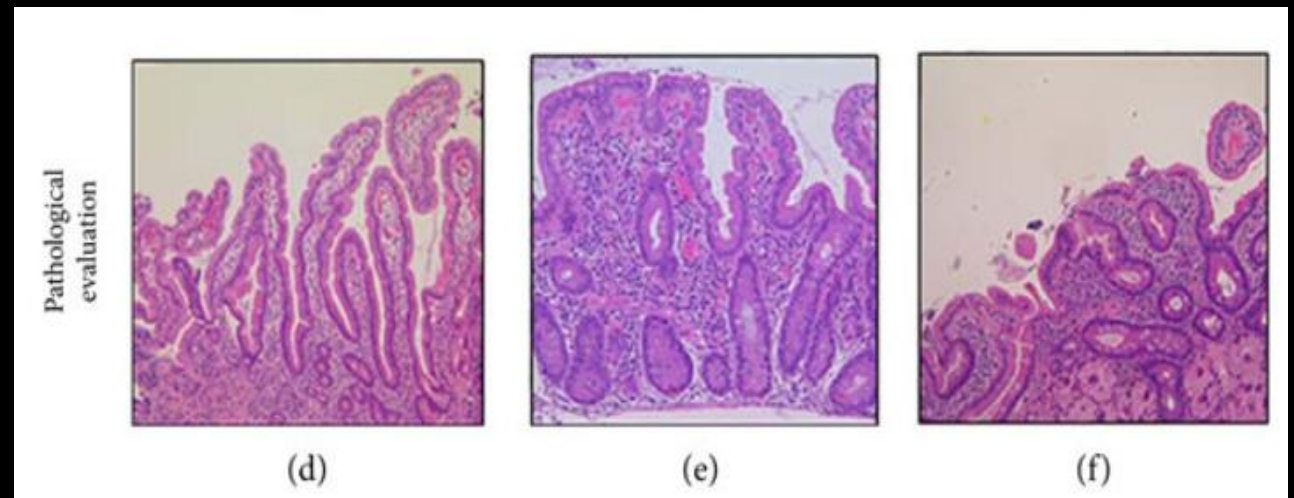
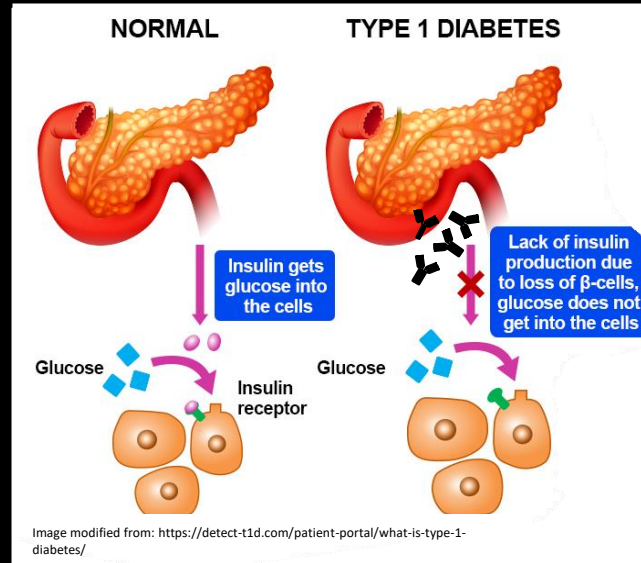
Case 1: Cont'd.

- Despite insulin treatment, diarrhea persisted, as well as continued failure to thrive, prompting a duodenal biopsy.
 - Biopsy revealed almost total villous atrophy, with dense infiltration of T cells and plasma cells.
- Follow up discussion with the family revealed no major health complications excluding a case of atopic dermatitis shortly after birth of the patient.
- Mother revealed that another son had perished in infancy with severe diarrhea and a low platelet count.



Immune Dysregulation, Polyendocrinopathy, Enteropathy, X-Linked (IPEX) Disorder

- Familial autoimmune syndrome with an X-linked recessive pattern of inheritance:
 - Early onset enteropathy
 - Dermatitis
 - Endocrinopathy → Type I Diabetes, thyroid disease
 - Often with autoantibodies

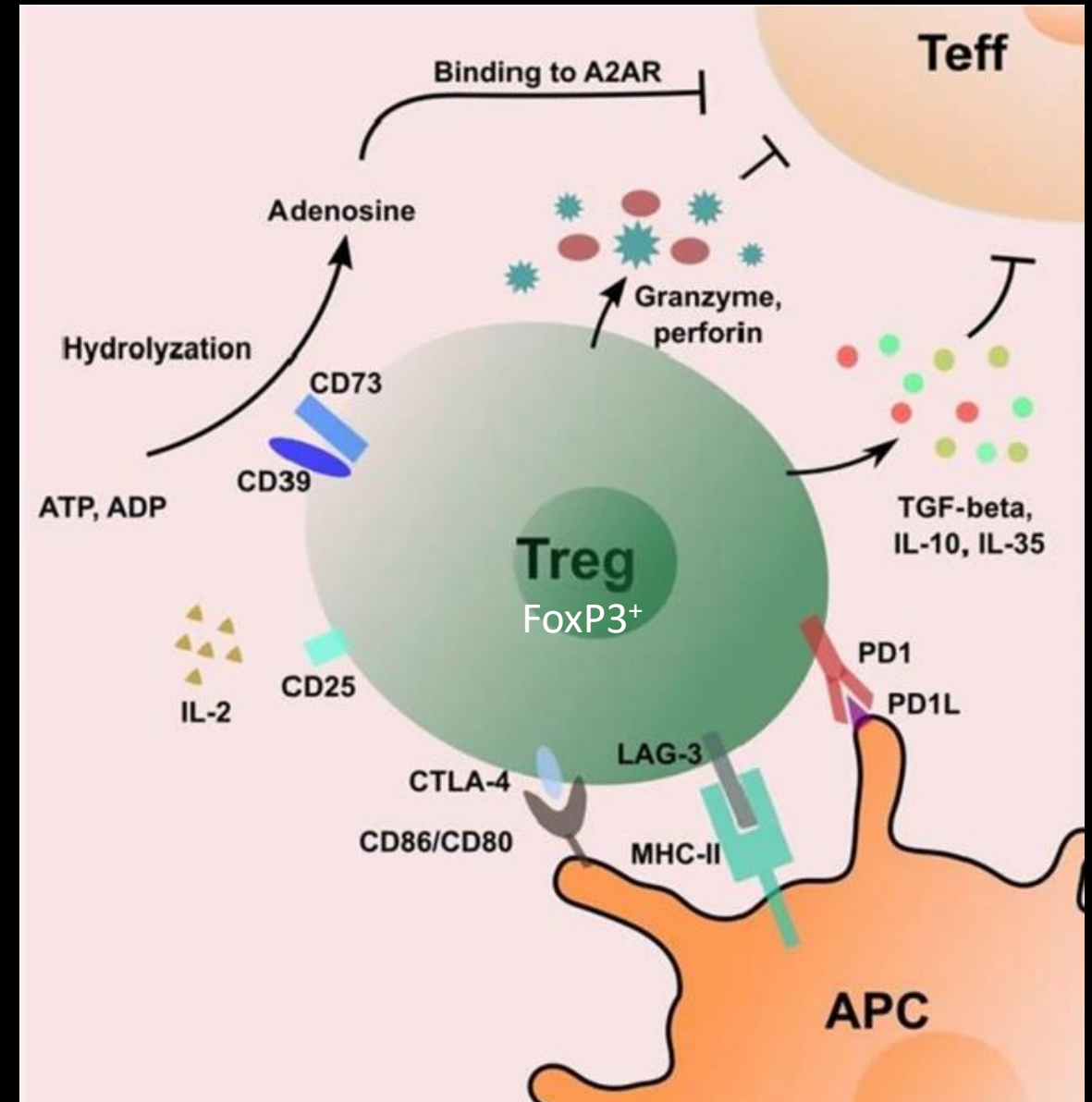


Sakai E, et al. Investigation of Small Bowel Abnormalities in HIV-Infected Patients Using Capsule Endoscopy. *Gastro Research and Practice*. 2017 (5):1-7

Etiology of IPEX → Regulation Breakdown

Dysfunctional or absence of T_{reg} cells leads to IPEX

- T_{reg} cells: subset of CD4 T cell involved inhibiting immune system
- Expression of the transcription factor forkhead protein 3 (FoxP3) commits to T_{reg} phenotype
- Variants or loss of FoxP3 expressing cells leads to autoimmunity

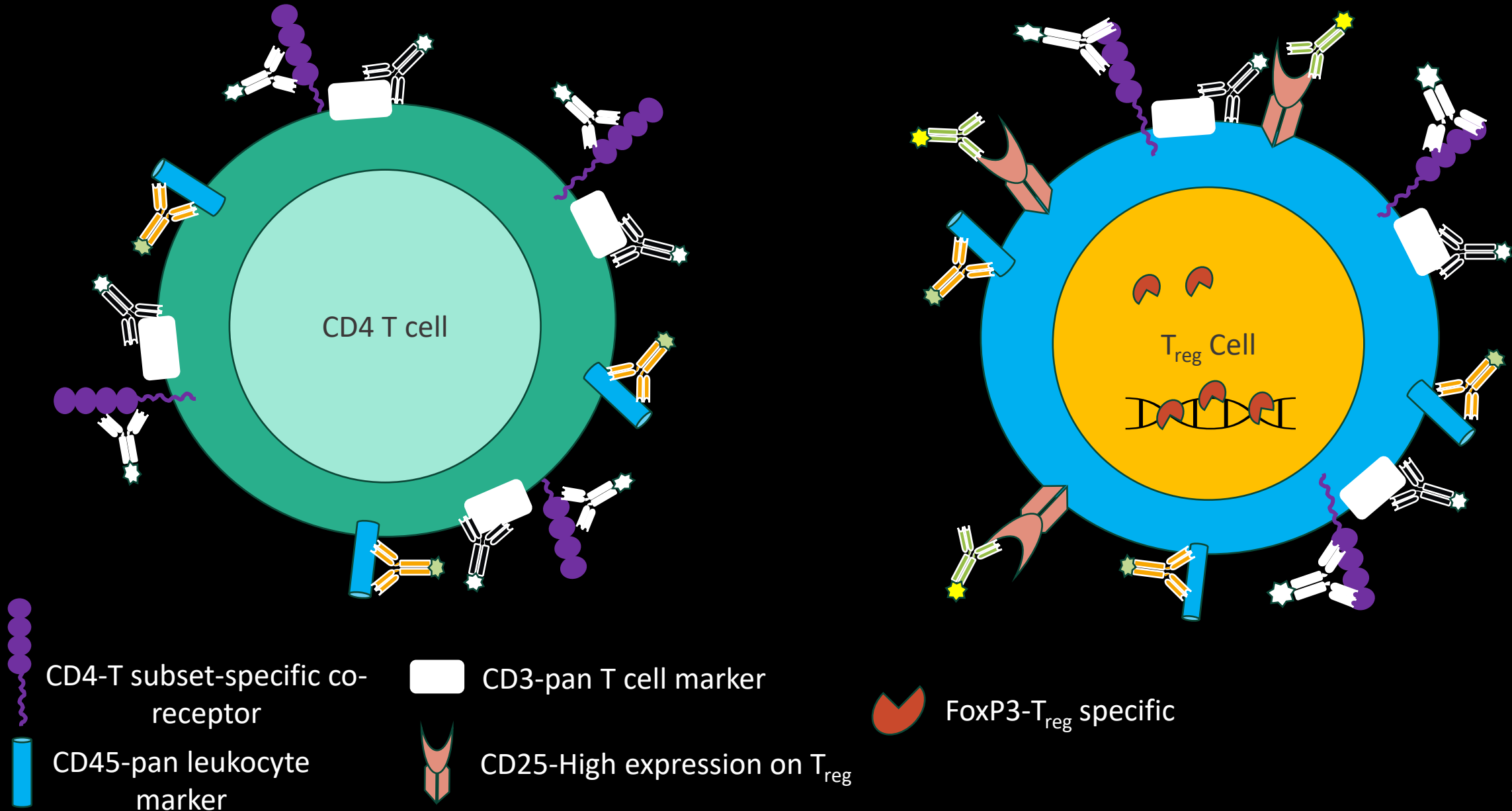




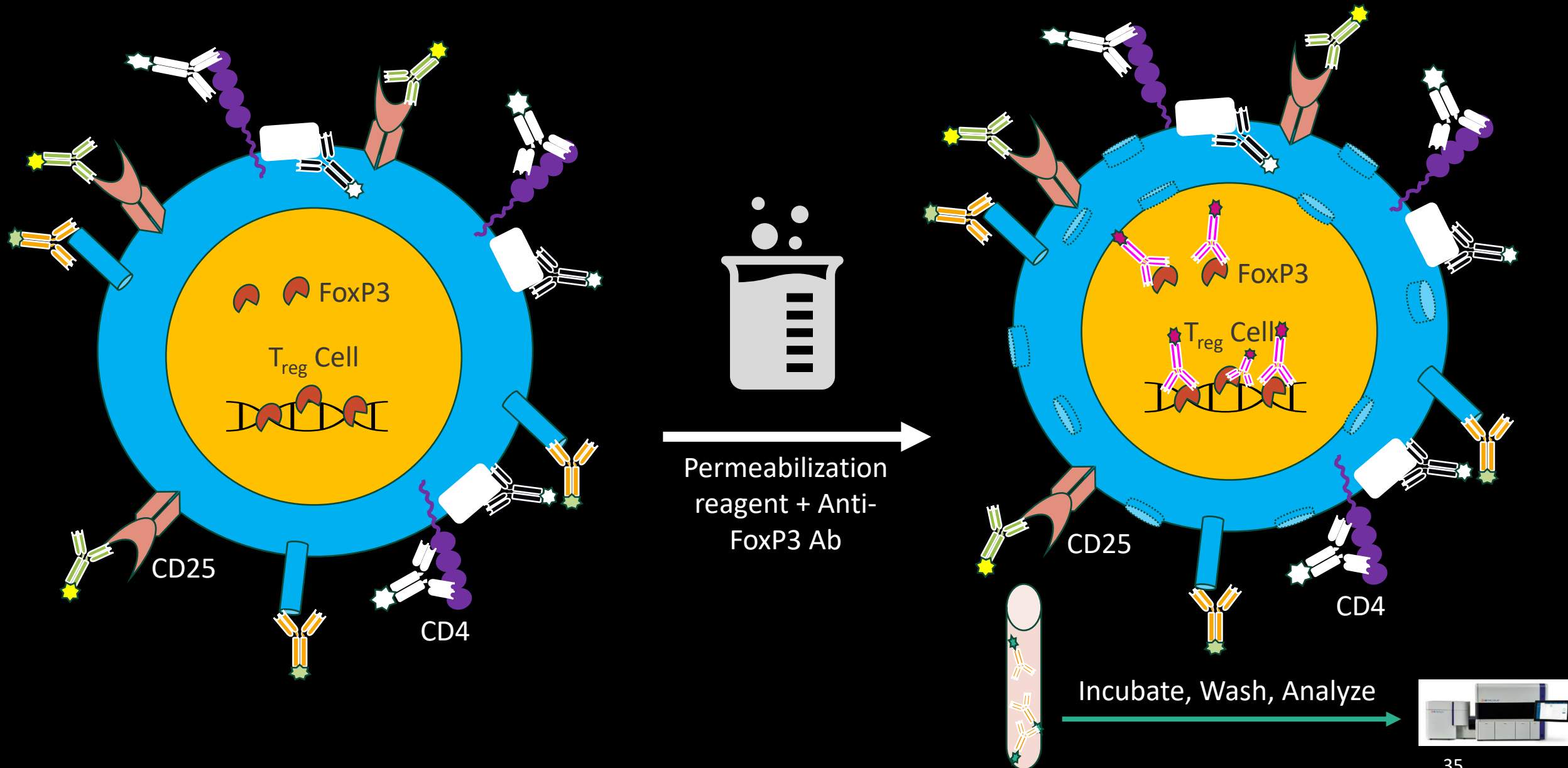
Diagnosis of IPEX

- Currently, identification of a pathogenic variant in *FOXP3* is considered the gold standard for establishing a diagnosis of IPEX
- Identification of variants can be difficult → non exon variants, deletion/duplication, variants of unknown significance etc.
- Adjunct assays targeting detection of T_{reg} cells and/or FoxP3 expression can supplement, confirm pathogenicity of genetic testing.

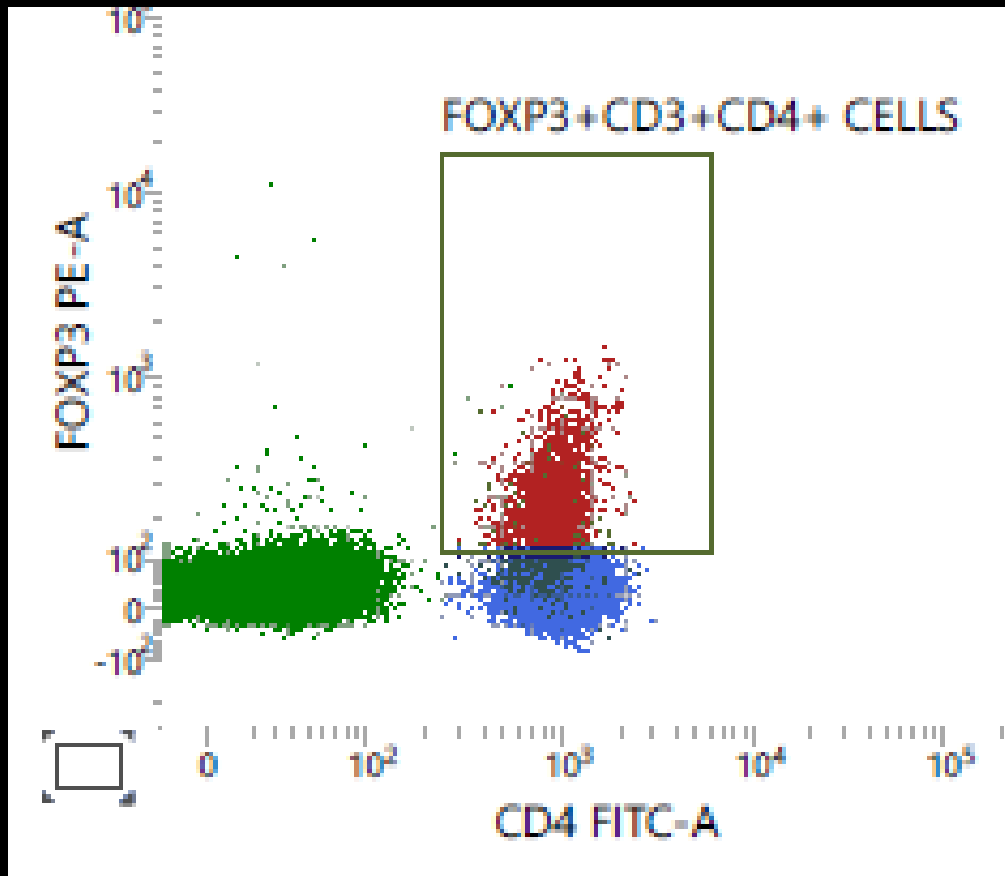
Regulatory T Cell Panel, FoxP3-Quantitative Assay



Regulatory T Cell Panel FoxP3 Staining



Normal Control

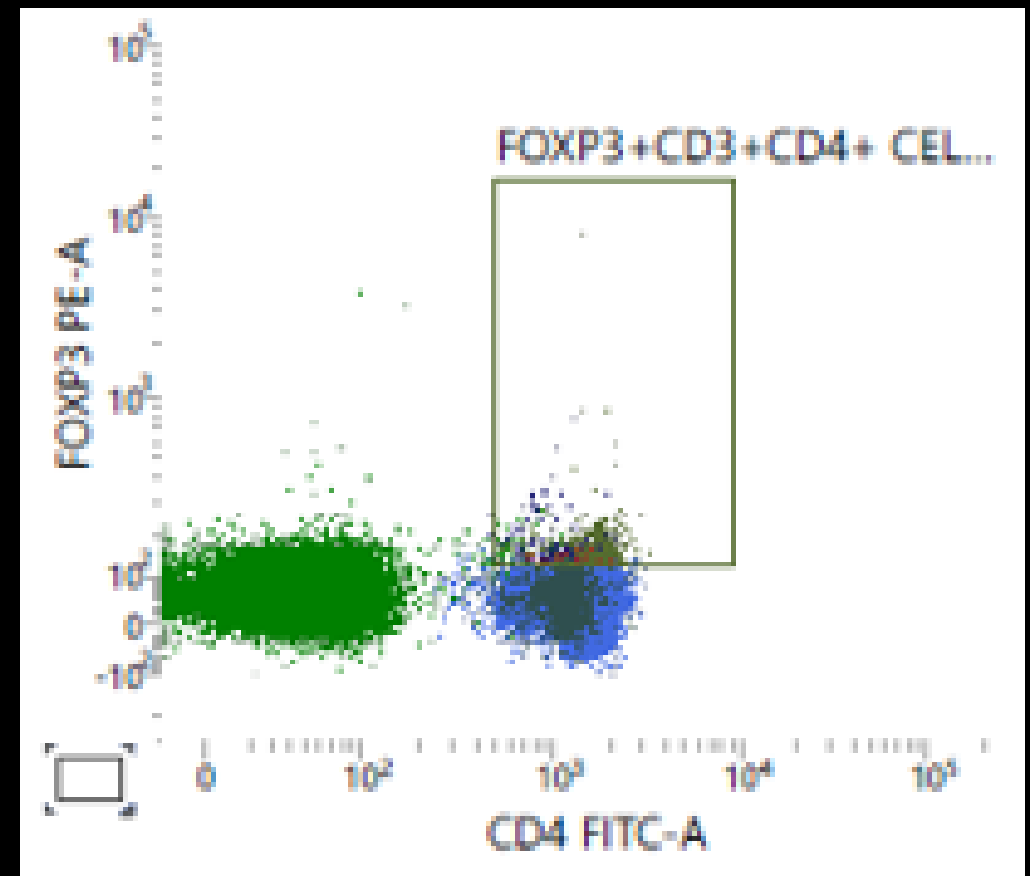


RESULTS

FOXP3TREGS %: 11.9

FOXP3TREGS Abs: 154

Suspect IPEX patient



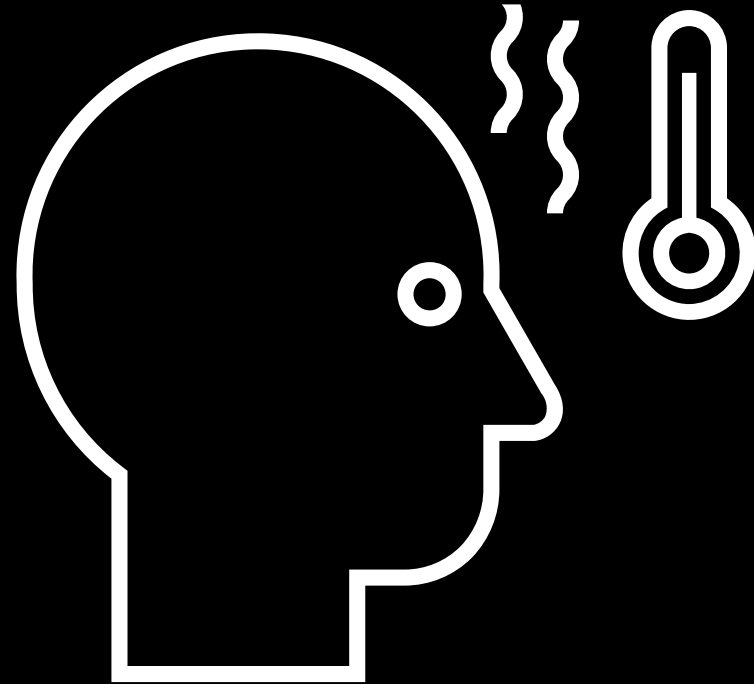
RESULTS

FOXP3TREGS %: 0.6

FOXP3TREGS Abs: 6

Case 2: A Curious Case of Neutropenia

- A 67-yr old male with hyperlipidemia (high cholesterol) presents to the clinic with a fever lasting > 3 days.
- Aside from fever, the patient's physical condition was unremarkable.
- Initial blood test revealed leukocytopenia (low white blood cell count).



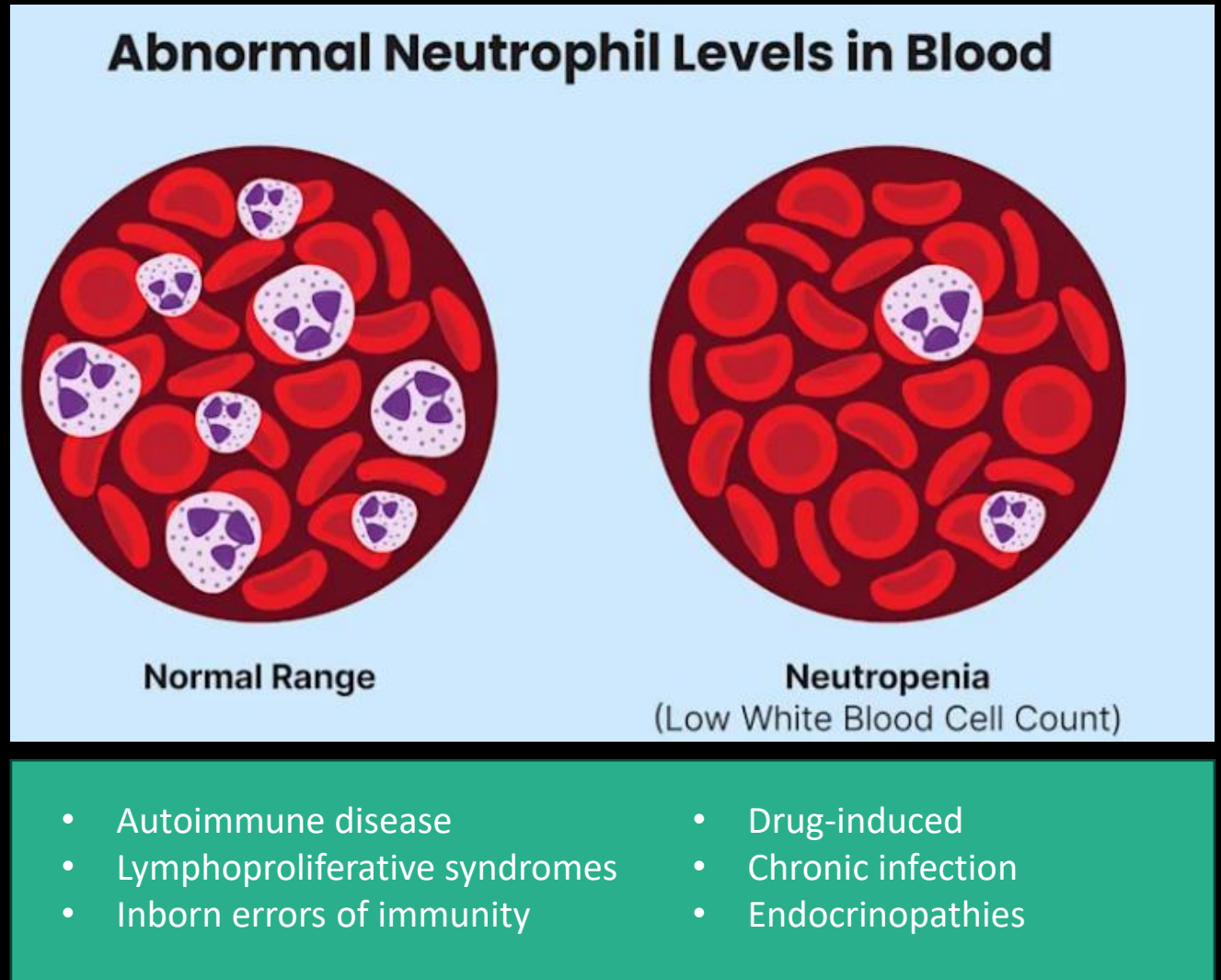
Case 2: Cont'd

- Total WBC count was 1100/ μ l, with a neutrophil count of 33/ μ l (**LOW**)
- Bone marrow aspiration revealed no morphological dysplasia in any of the hematological cell lineages, nor abnormal cell populations
- A CT scan displayed no signs of infection, swollen lymph nodes, or hepatosplenomegaly

What could be mediating neutropenia in this patient?!

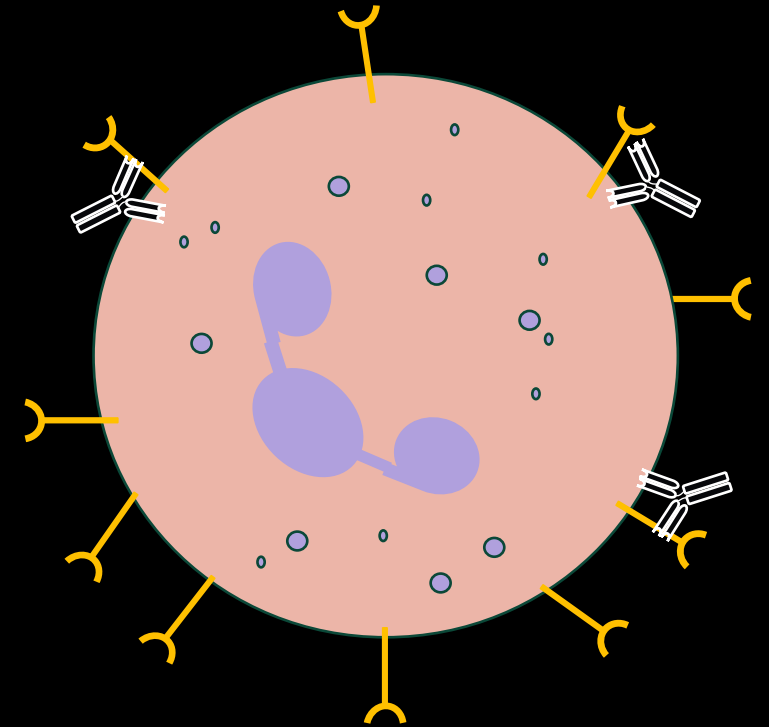
Secondary Neutropenia

- Neutrophil: primary mediator of rapid innate immune response
- Neutrophil count below 500 cells/ μ L greatly increases risk of infection
 - Mild: 1000-1500 cells/ μ L
 - Moderate: 500-1000 cells/ μ L
 - Severe: <500 cells/ μ L
- Common manifestation of severe disease → unspecific (broad differential diagnosis)

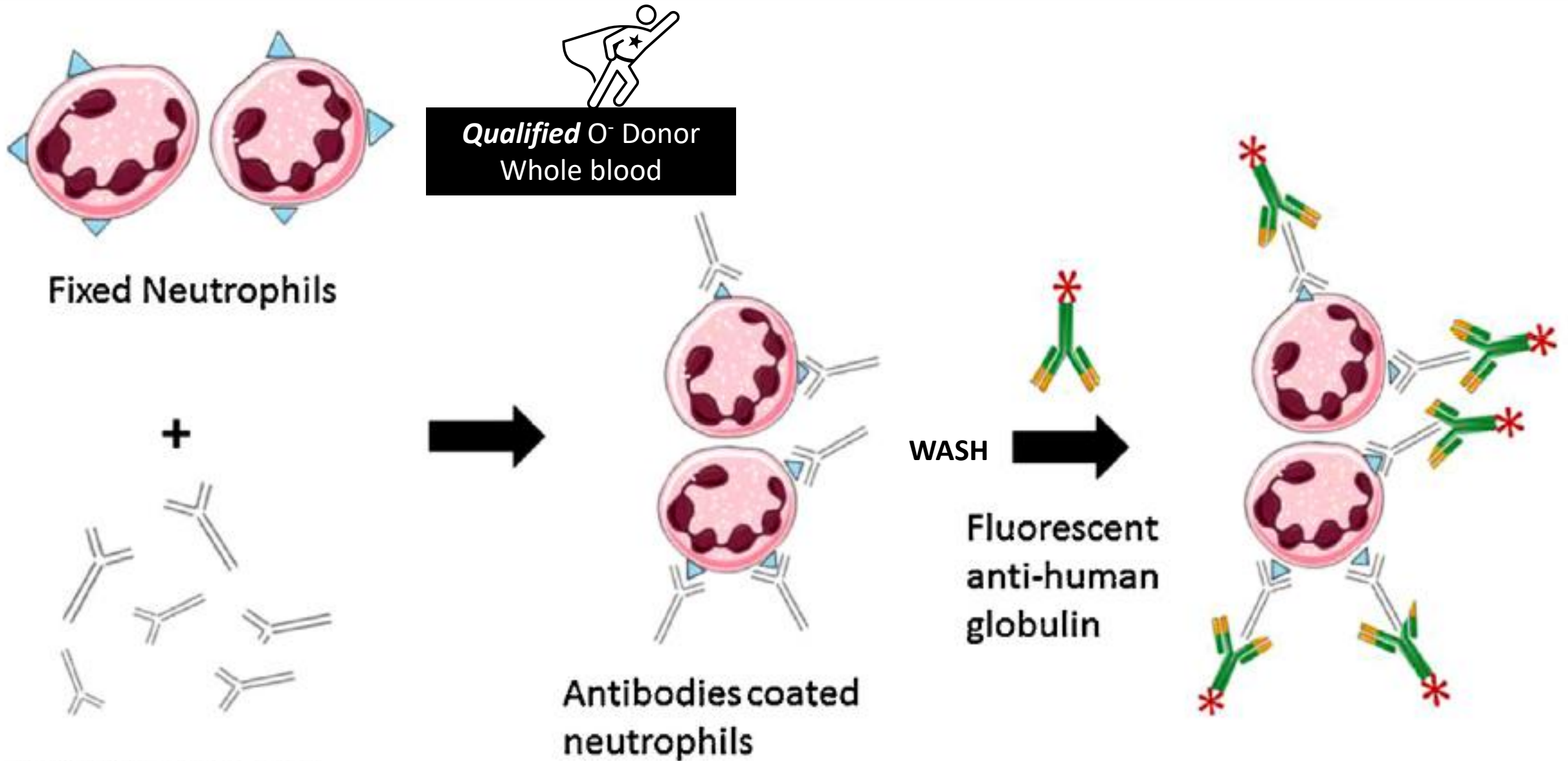


Anti-Neutrophil Antibodies and Neutropenia

- Autoimmune neutropenia (AIN): hematological diseases caused by autoantibody induced destruction of neutrophils
 - Detection of neutrophil autoantibody can help exclude other causes of neutropenia, and inform treatment strategies
 - Granulocyte-colony stimulating factor (G-CSF)
 - Immunosuppressive drugs (prednisolone)
 - IVIG, rituximab, alemtuzumab



-opsonization = increased clearance
-agglutination and phagocytosis
-complement-mediated destruction



Fixed Neutrophils

+

Patient serum with antibodies

Qualified O⁻ Donor Whole blood

Antibodies coated neutrophils

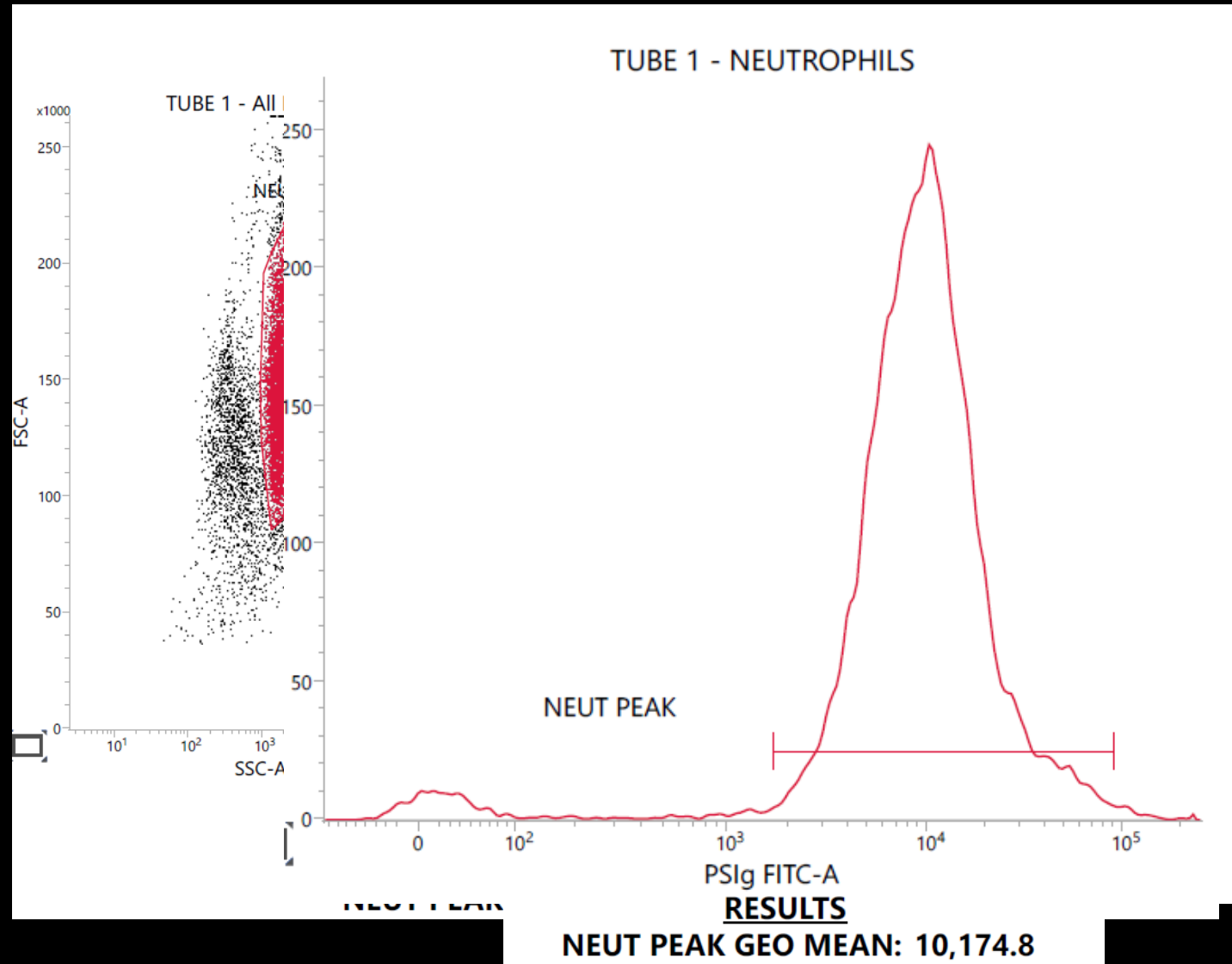
WASH

Fluorescent anti-human globulin

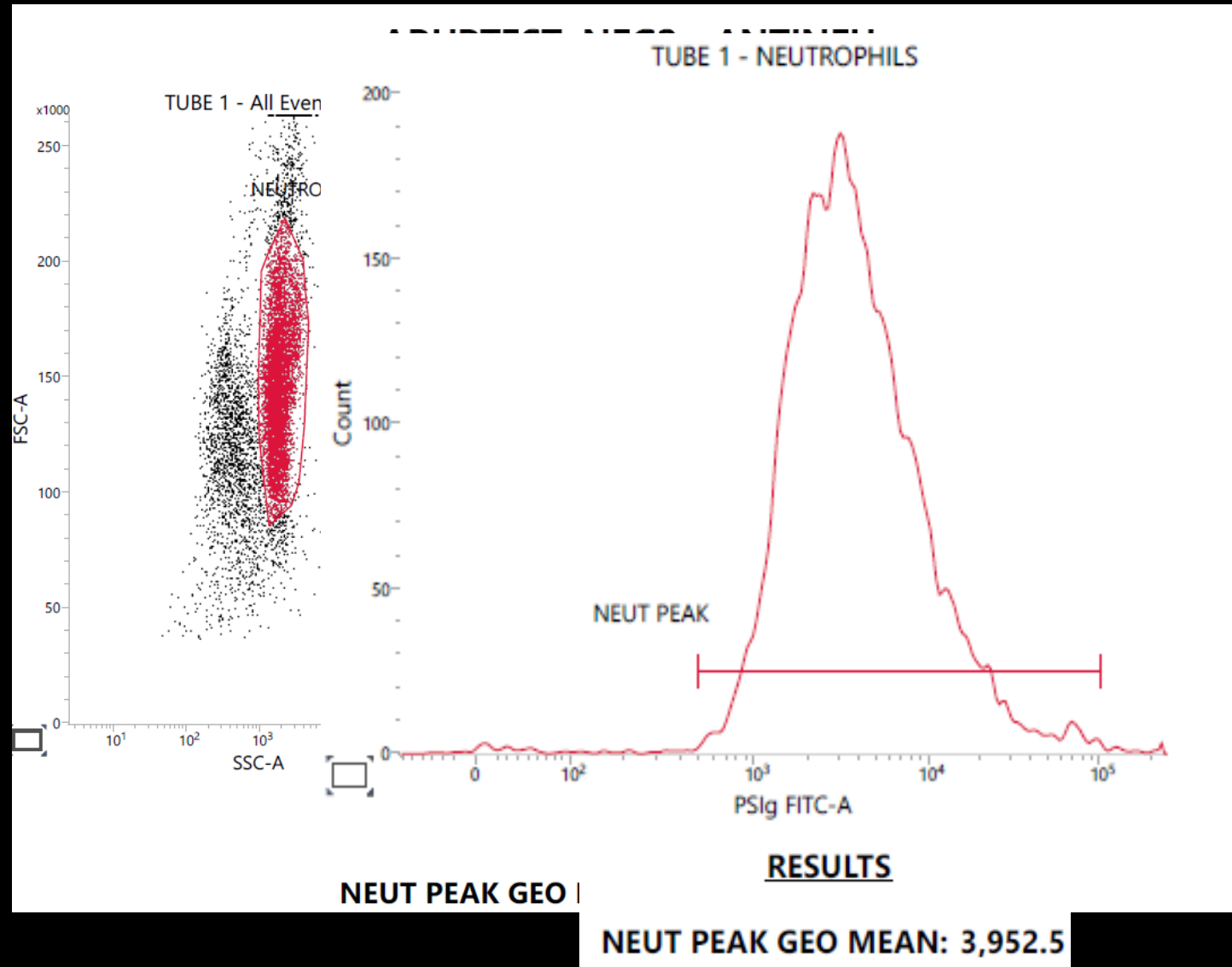
Adapted from Fung YL et al, Vox Sang, 2011 [23]

Image adapted from: Autrel-Moignet et al. Autoimmune Cytopenias Quarterly Medical Review. 2014.

Anti-Neutrophil Antibody Assay Result Interpretation:

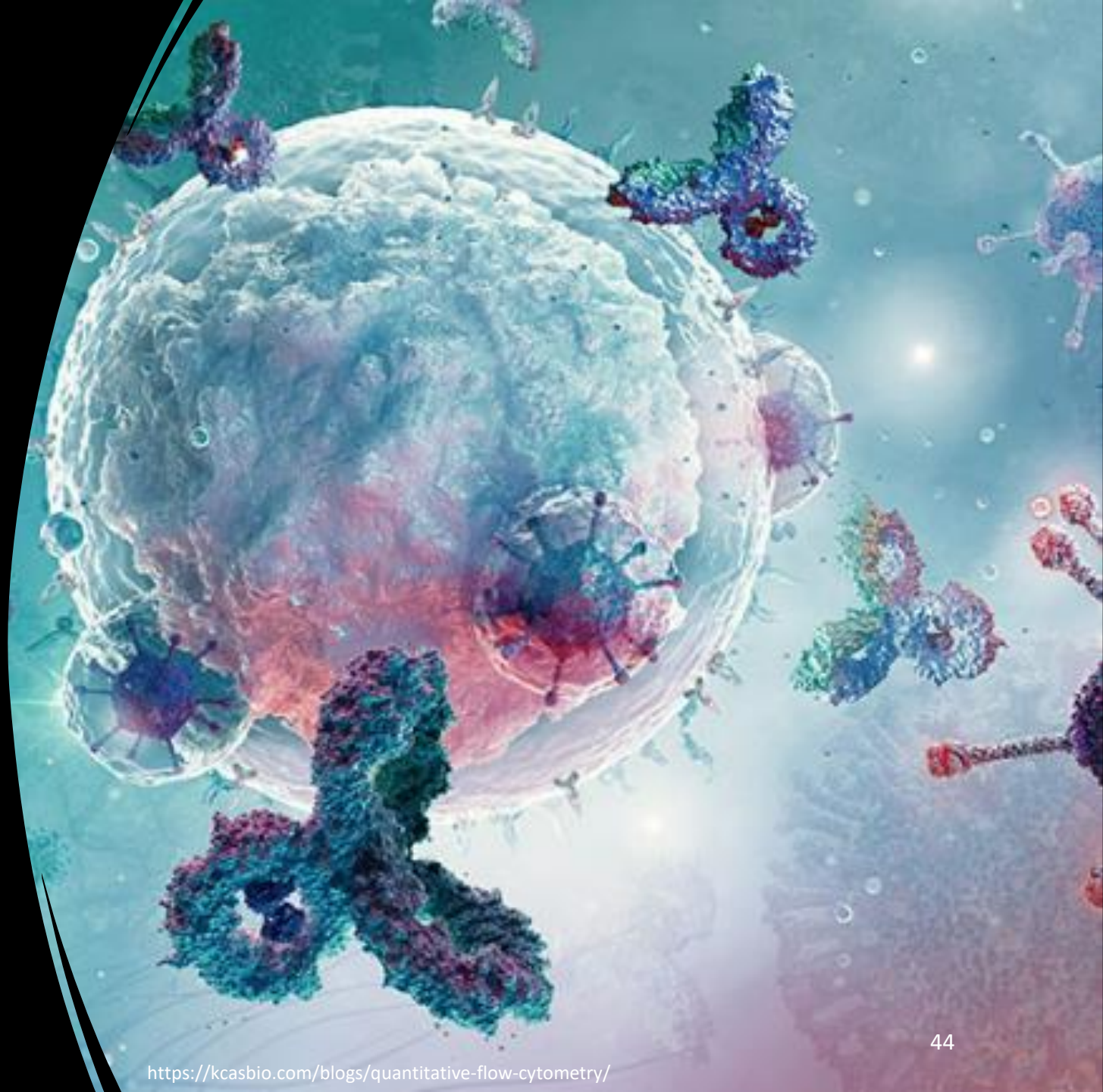


Anti-Neutrophil Antibody Assay Result Interpretation: Weak Positives



Flow Cytometry: Incyte for the Clinic

- Flow cytometry is a powerful tool for analyzing clinical samples.
 - Multiparametric analysis
 - High sensitivity and throughput
 - Versatile: Live, fixed, quantitative, functional assays, rare event analysis, sorting
- Versatility of flow cytometry allows its application to diverse clinical scenarios where it can be a key factor in informing practice, diagnosis, and monitoring of disease.



Thank You!

Knowledge Check

Further work up with a *FoxP3* genetic panel did not detect a variant in the patient. Which of the following flow cytometric approaches would you consider most useful to further investigate the possibility of IPEX?

- A. Total CD4 and CD8 T cell enumeration via flow cytometry.
- B. T cell proliferation assay via flow cytometry.
- C. Quantify total FoxP3 expressing cells.
- D. Intracellular cytokine measurement.

Knowledge Check:

For a quantitative, intracellular flow assay, which step(s) are crucial to include?

A. Fixation

B. Permeabilization

C. Collection of enough target cell events

D. Use of a normal reference control

Knowledge check

Which of the following factors should be considered when designing a panel to detect anti-neutrophil antibodies using flow cytometry?

- A. An assessment of neutrophil viability.
- B. Permeabilization to detect *intracellular* anti-neutrophil antibodies.
- C. Criteria for a minimal number of neutrophils analyzed per sample to meet sensitivity requirements.
- D. All of the above.

Knowledge Check

How can we use fluorescence intensity to resolve borderline results in this flow cytometric assay?

- A. Use a reference material, such as BD TruCount™ beads
- B. “Eye-ball it”
- C. Use negative and normal controls to establish positive cut-off
- D. Borderline results suggest pre-analytical issue, retest