

# Clinical Flow Cytometry for the Perplexed

## Part 2: Technical Considerations

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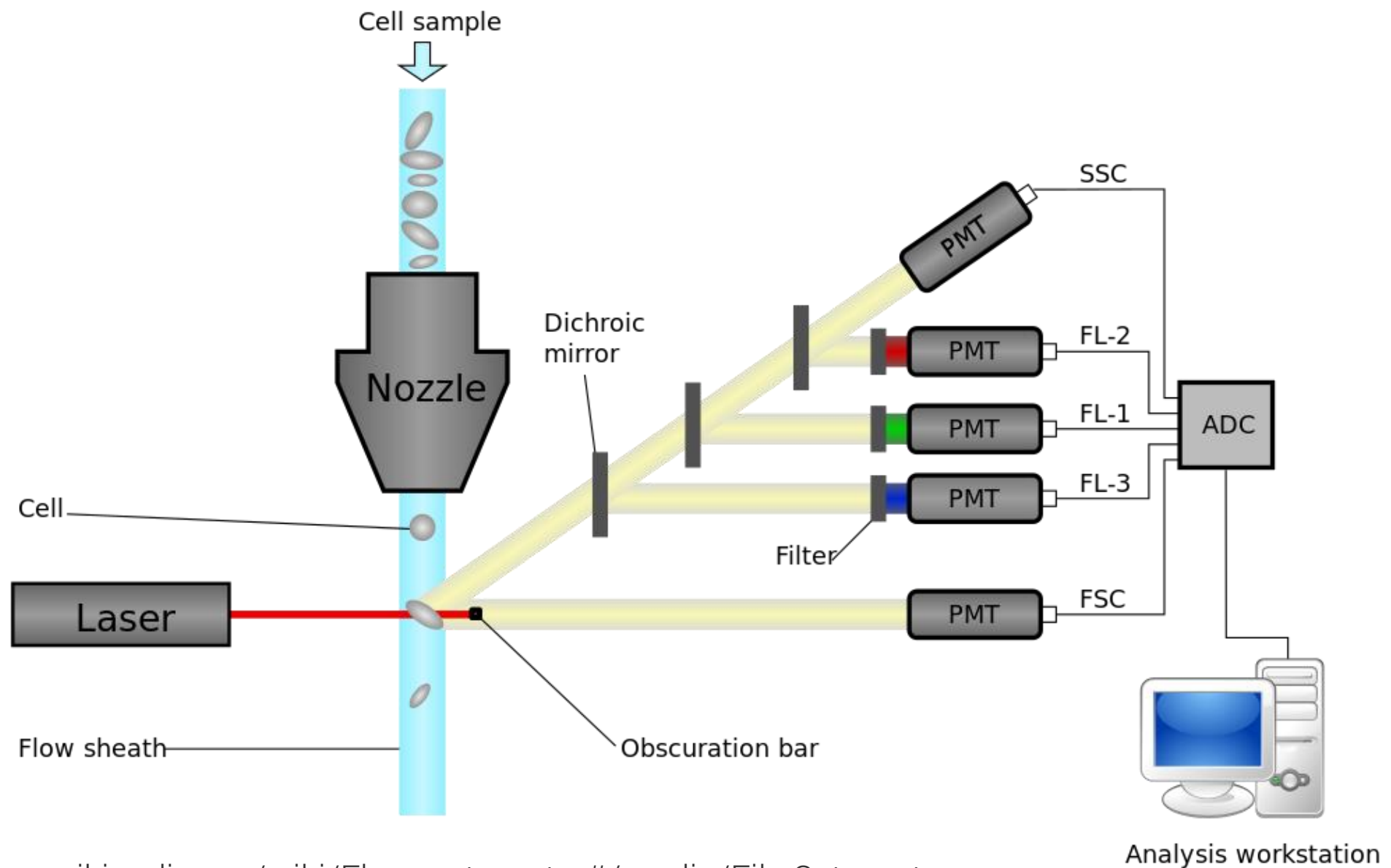
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# Recap from Part 1

- What is Flow Cytometry
- Gating Strategy(ies)

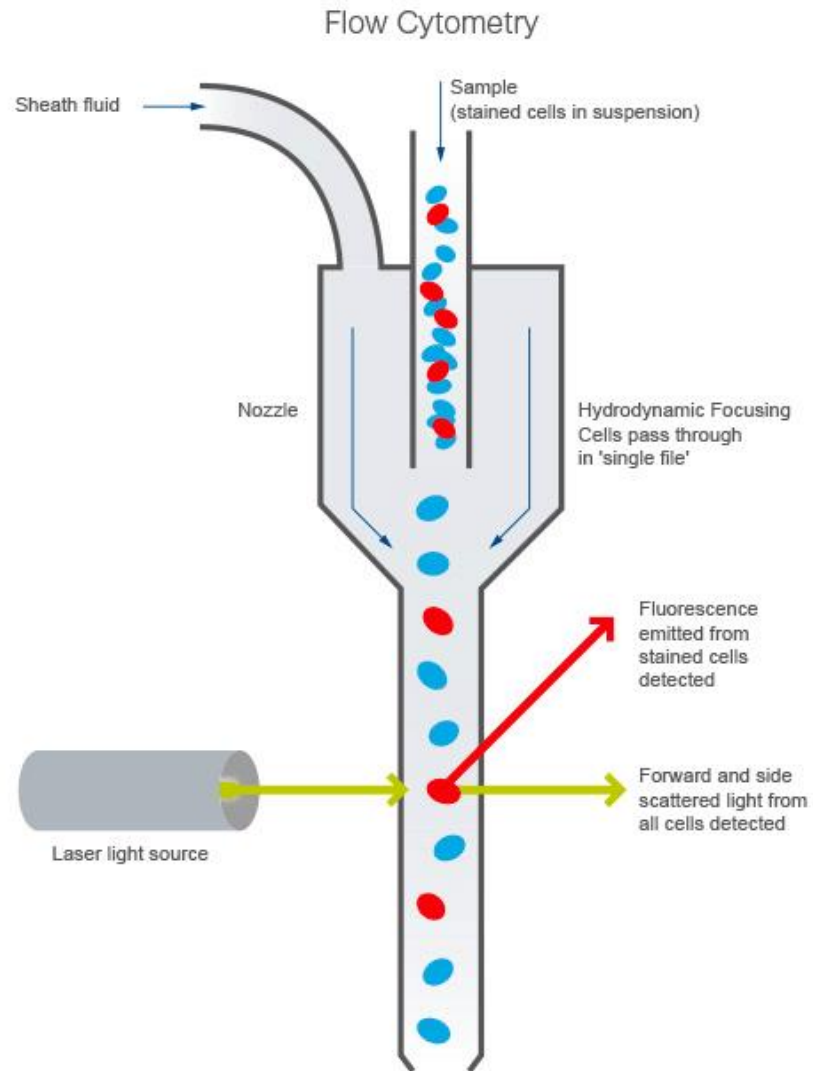
# Goals for Part 2

- How does a flow cytometer work?
- What are the pitfalls and artifacts that you need to know?
  - » Tandem Breakdowns
  - » Spillover/Compensation

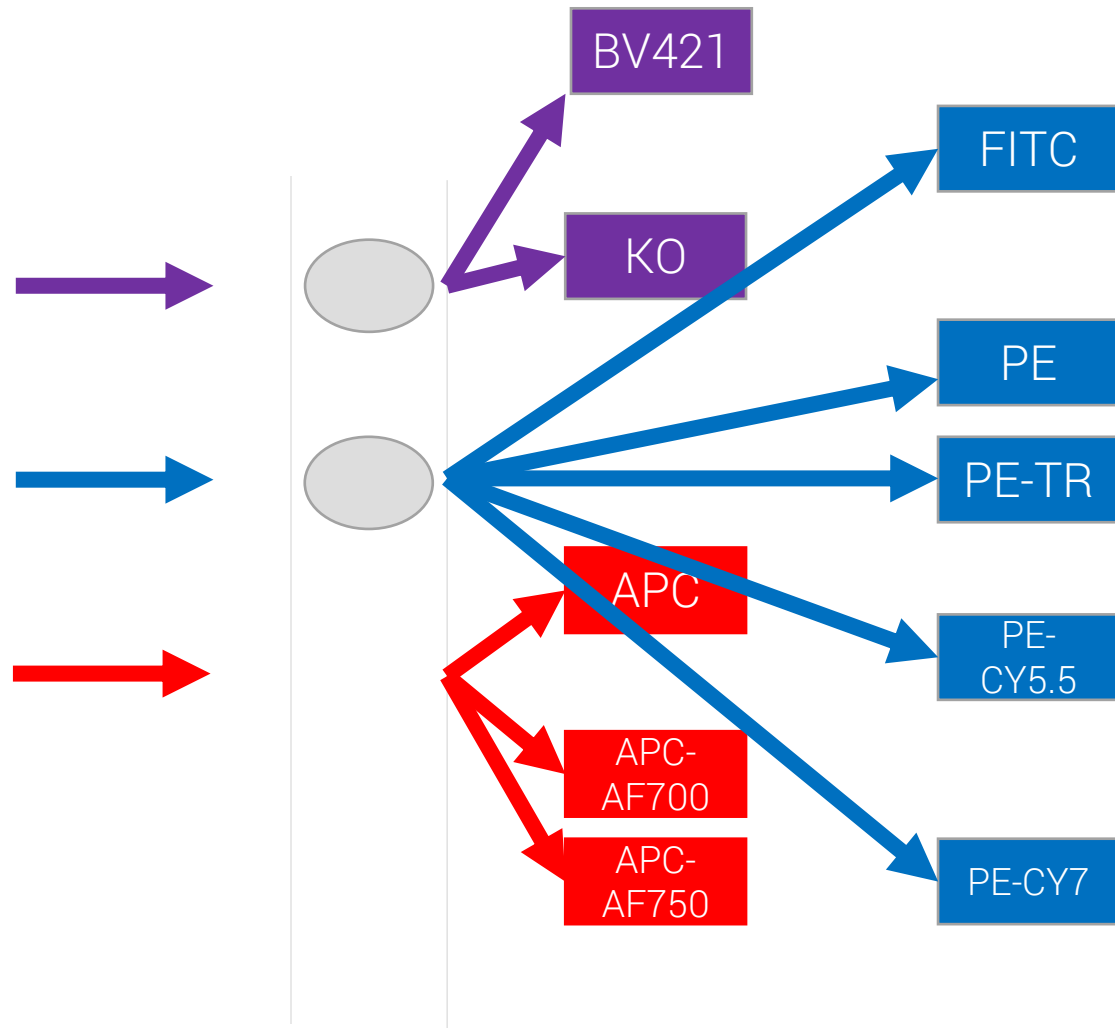


[en.wikipedia.org/wiki/Flow\\_cytometry#/media/File:Cytometer.svg](https://en.wikipedia.org/wiki/Flow_cytometry#/media/File:Cytometer.svg)

# The Flow Cell

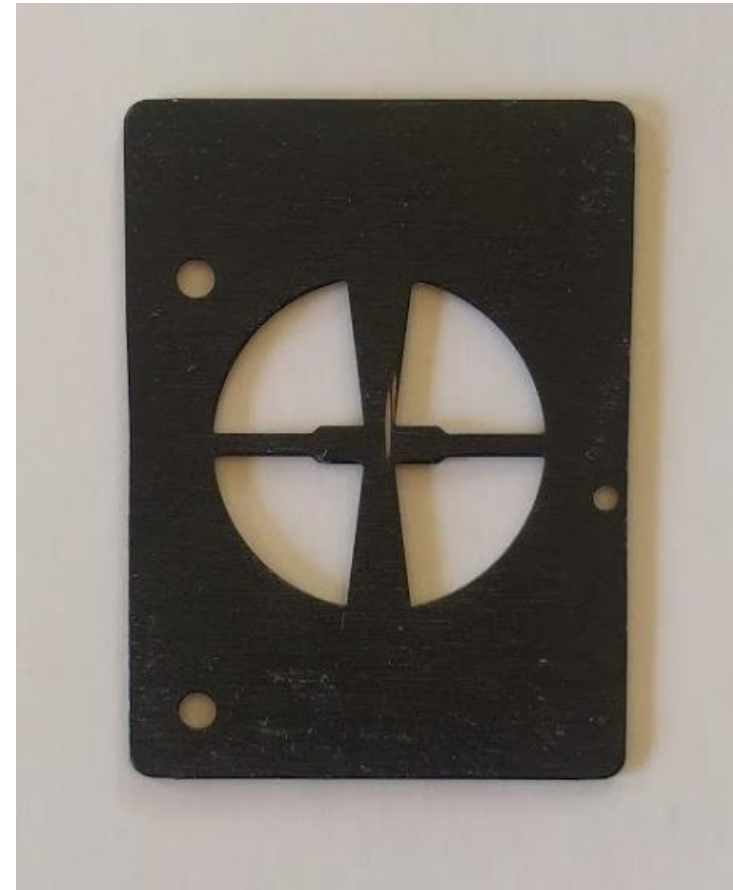


# Spatial Separation

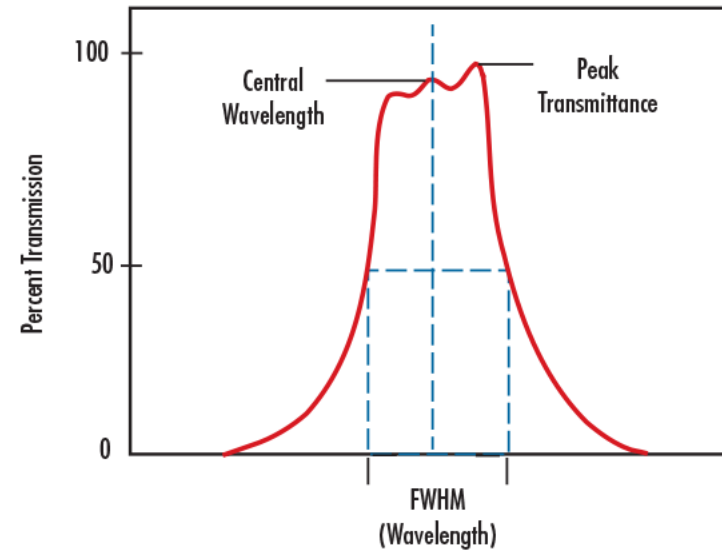
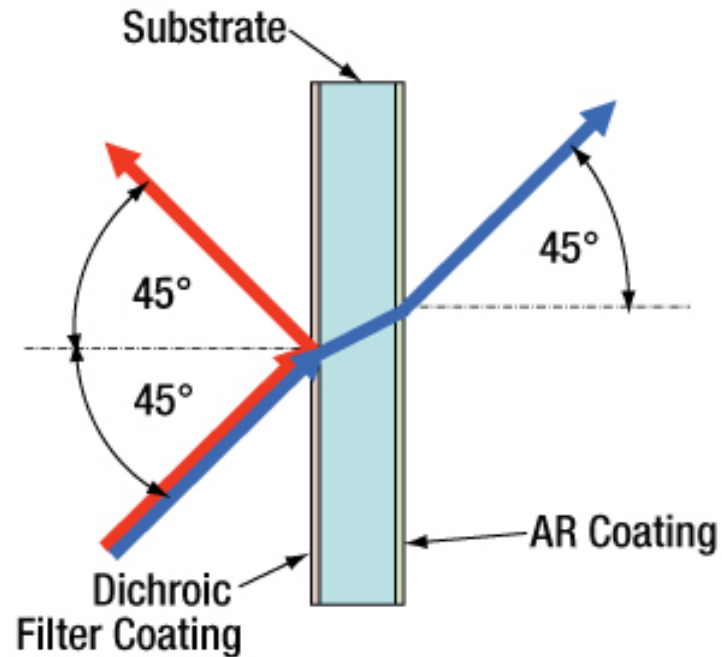


# Forward Scatter

- Roughly proportional to cell size

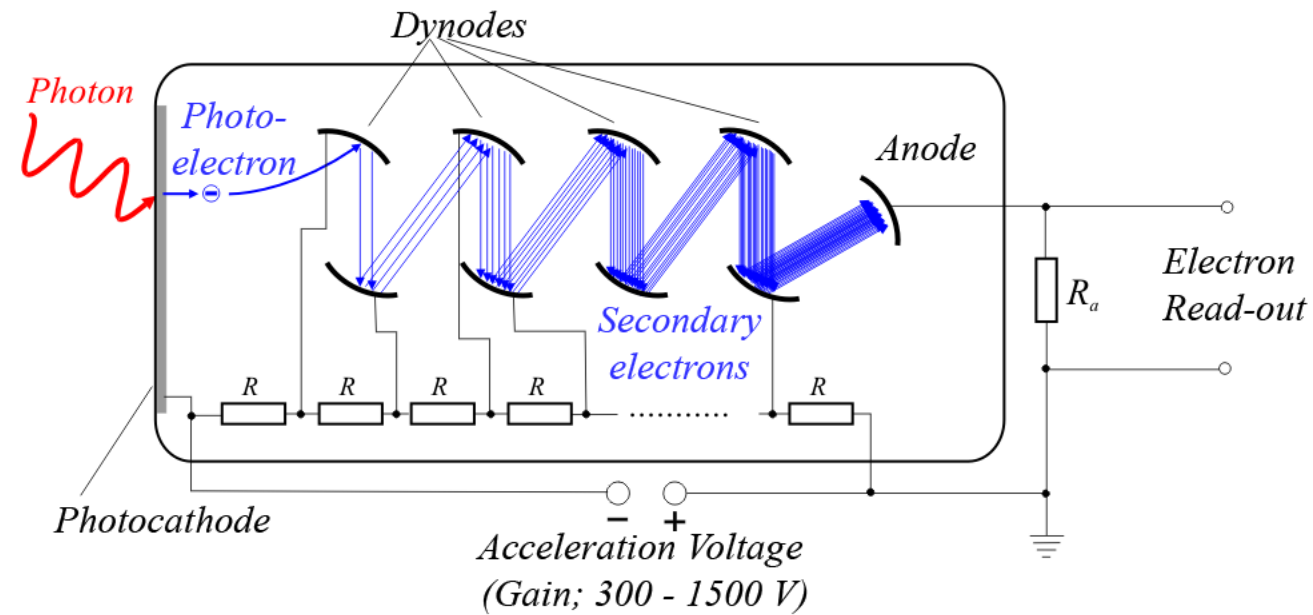


# Dichroic Mirrors / Bandpass Filters

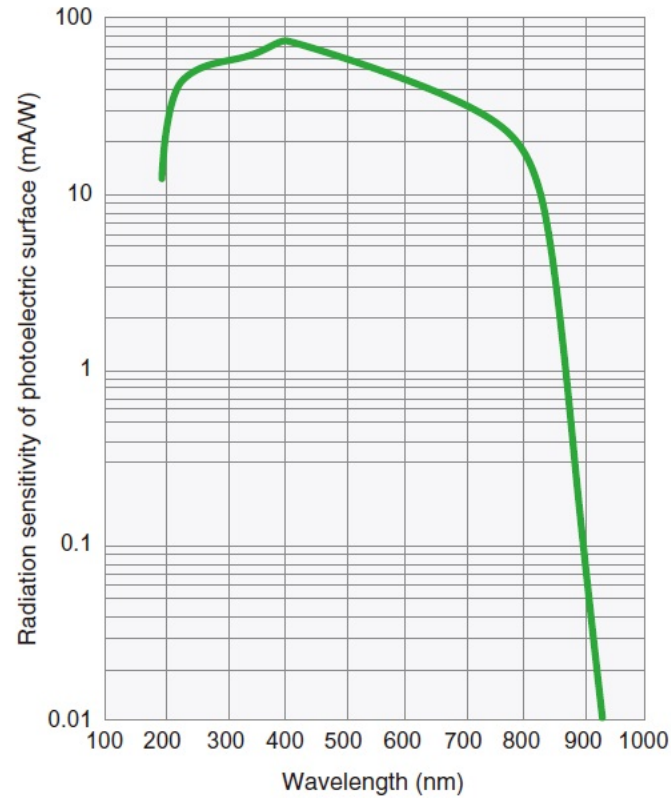




# Photomultiplier Tube

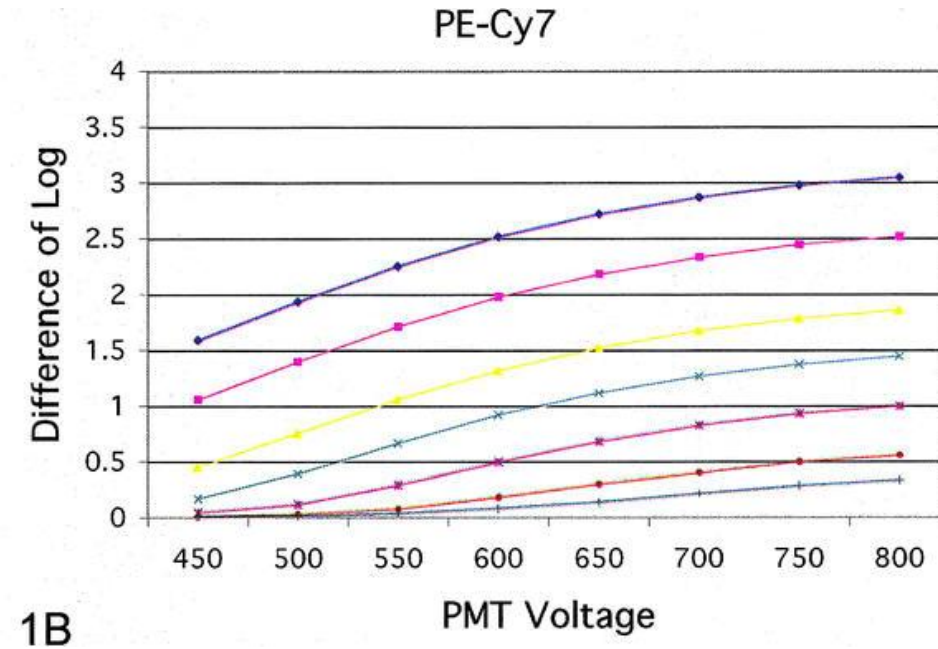
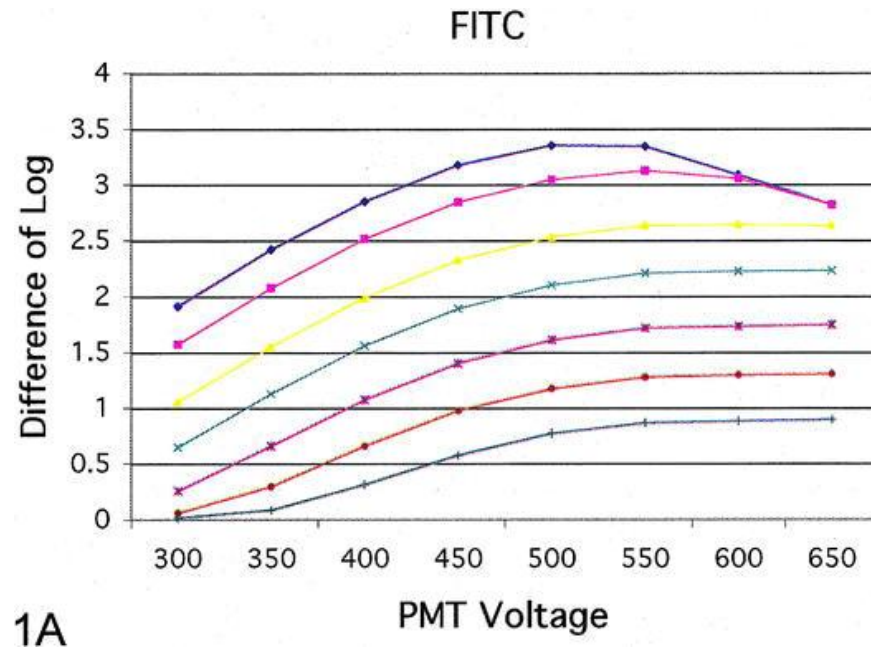


# Implications (sensitivity)

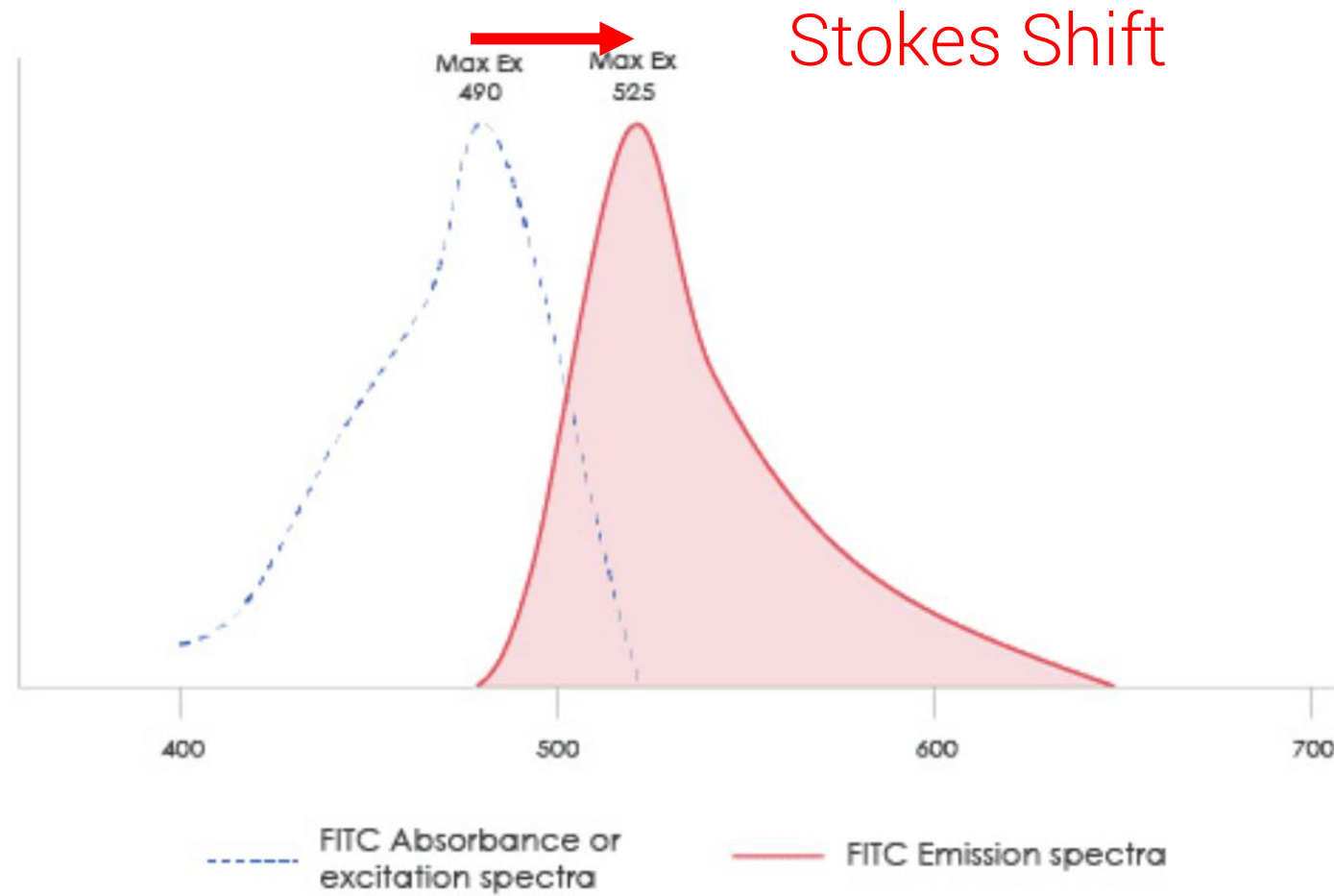


# Implications (gain)

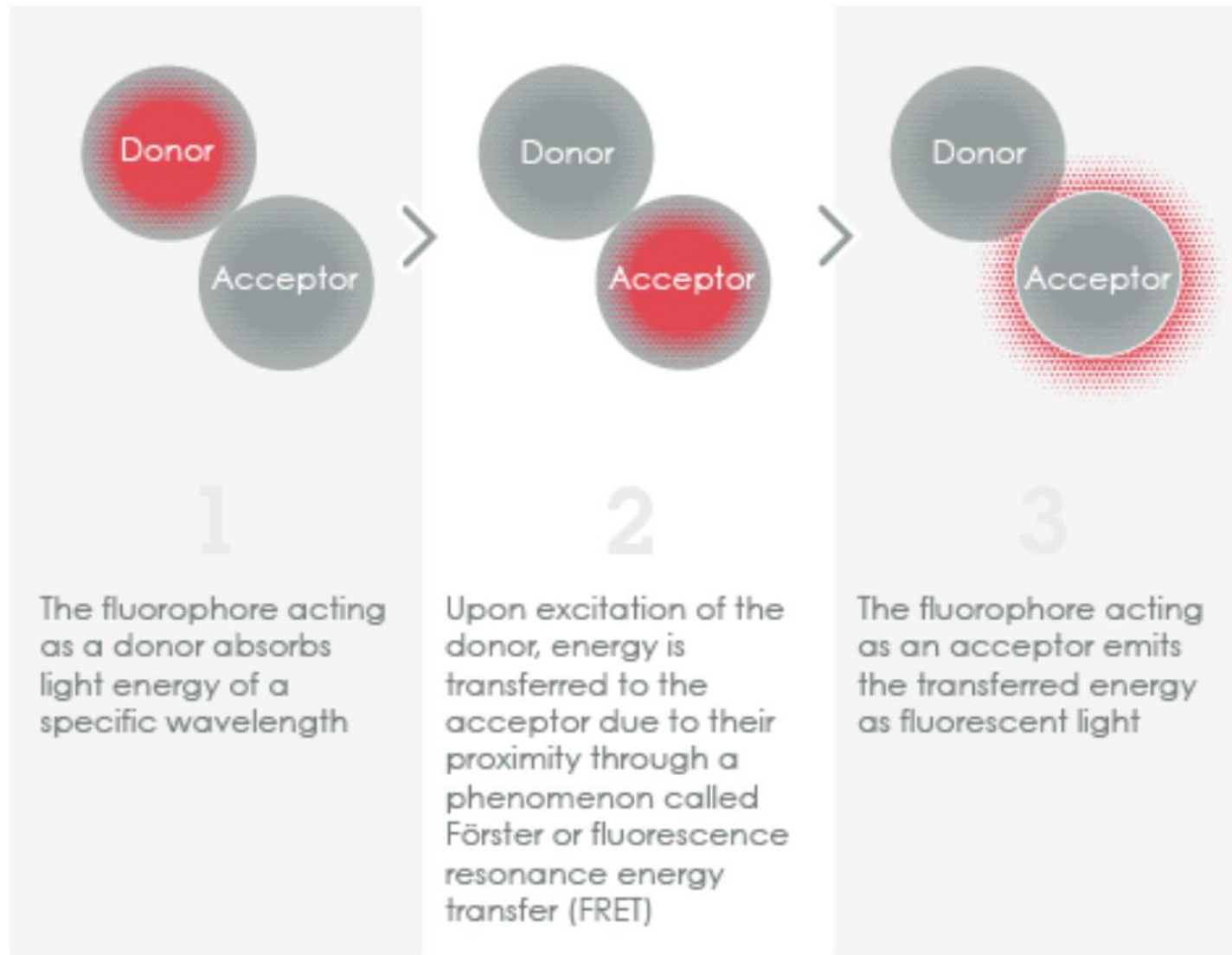
- Limited linear range of gains



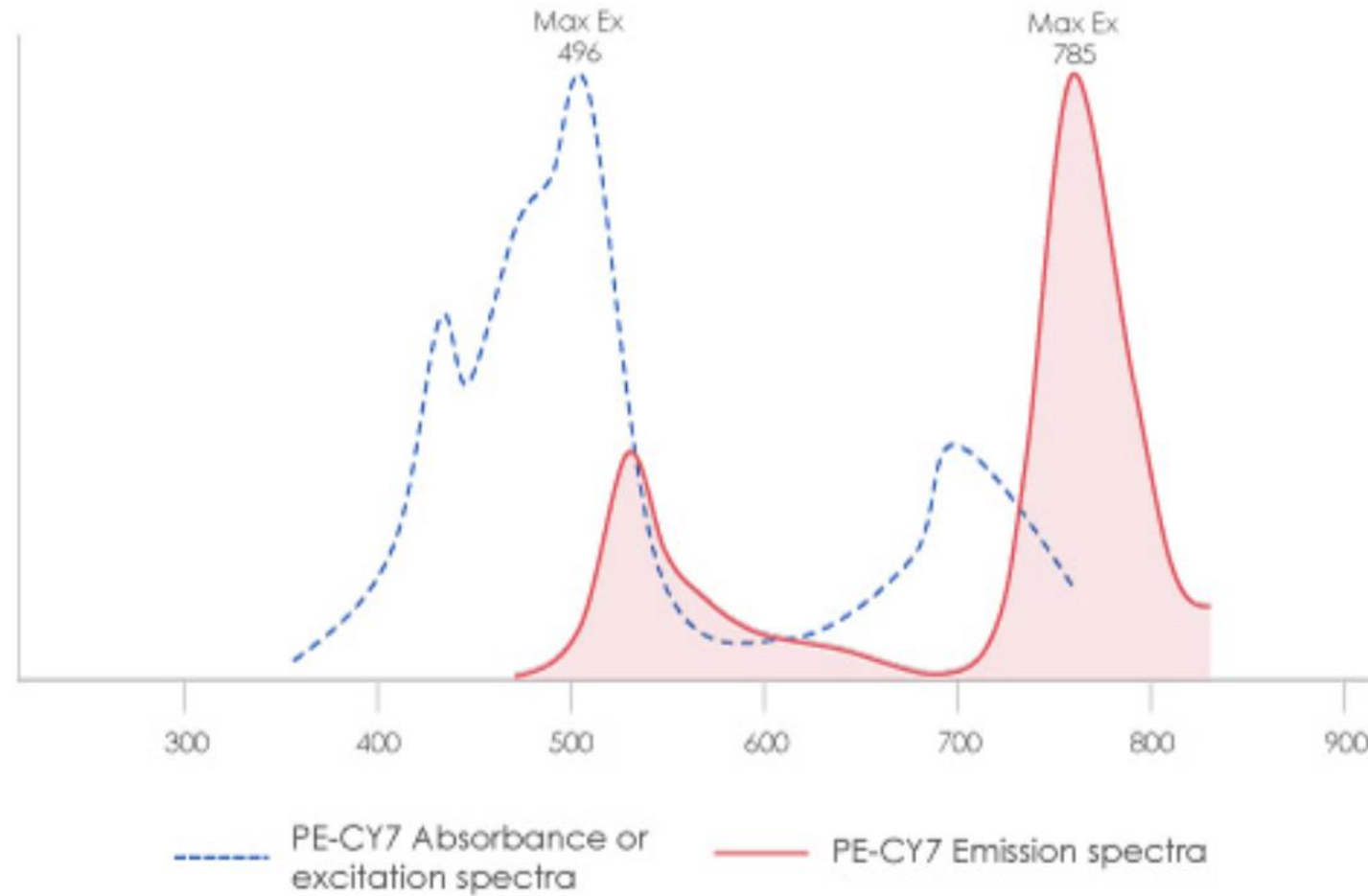
# Fluorophores



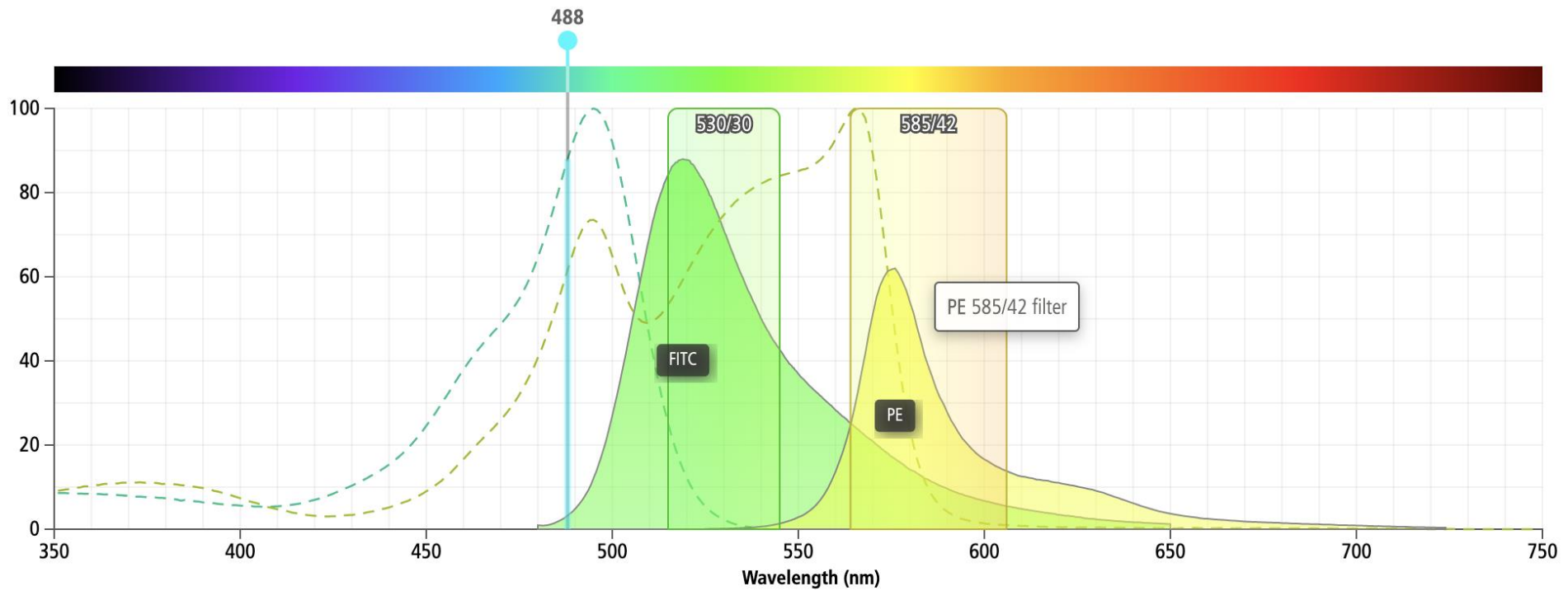
# Tandem Fluorophores



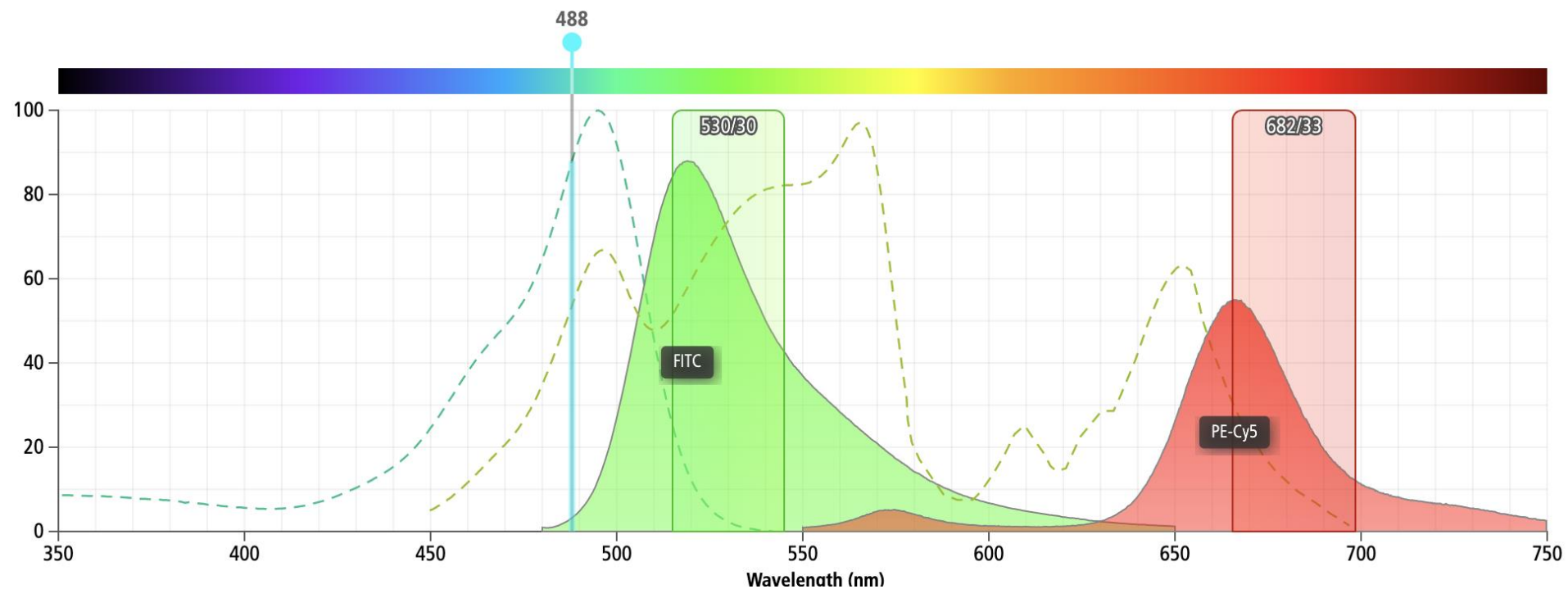
# Tandem Fluorophores



# Spillover

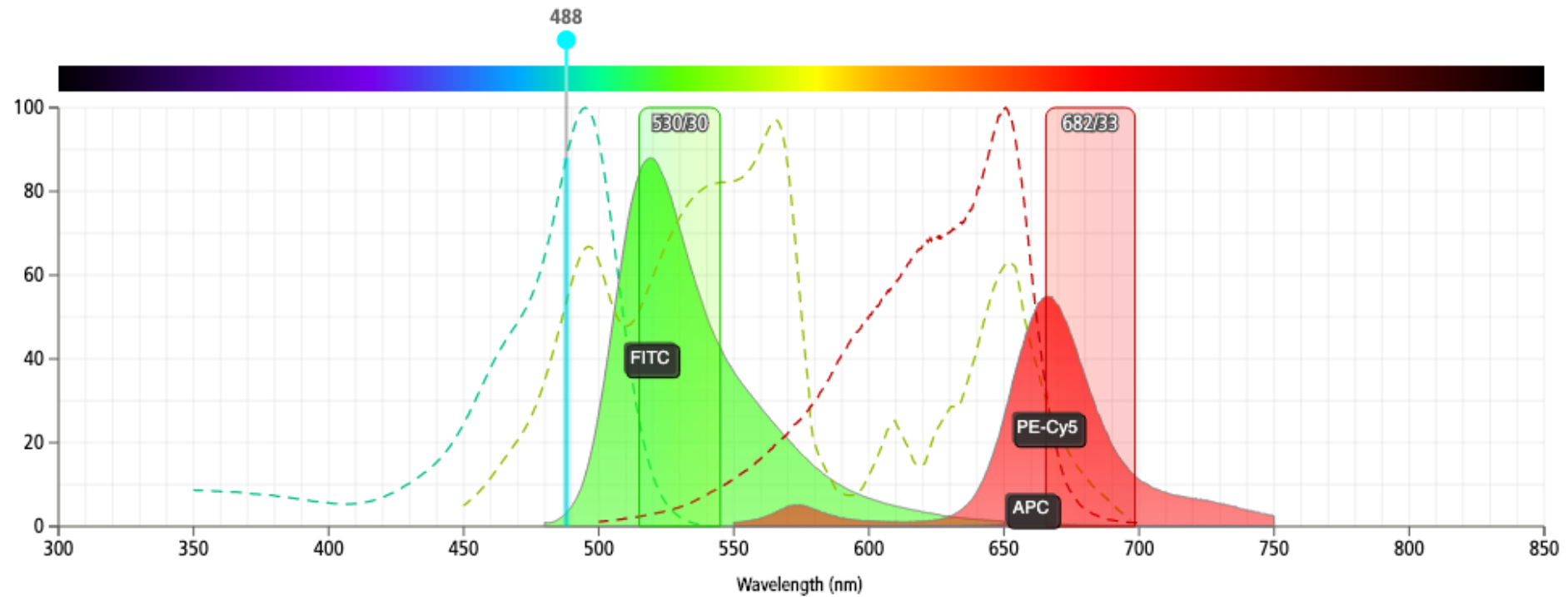


# Spillover Tandems

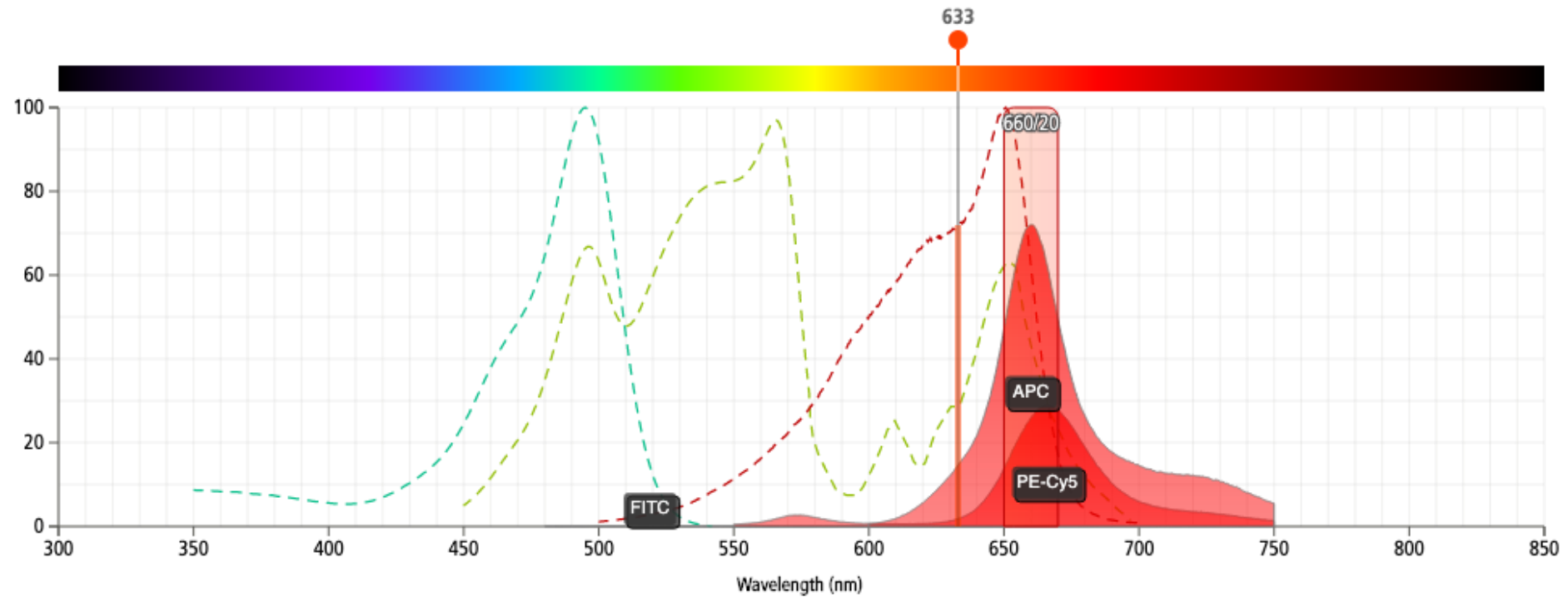


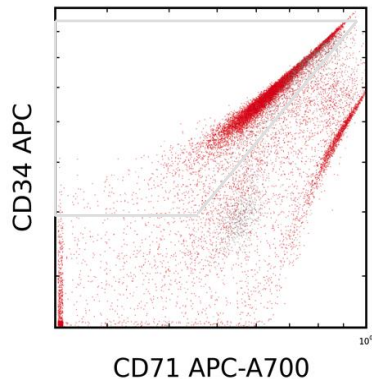


# Spillover - Crosslaser

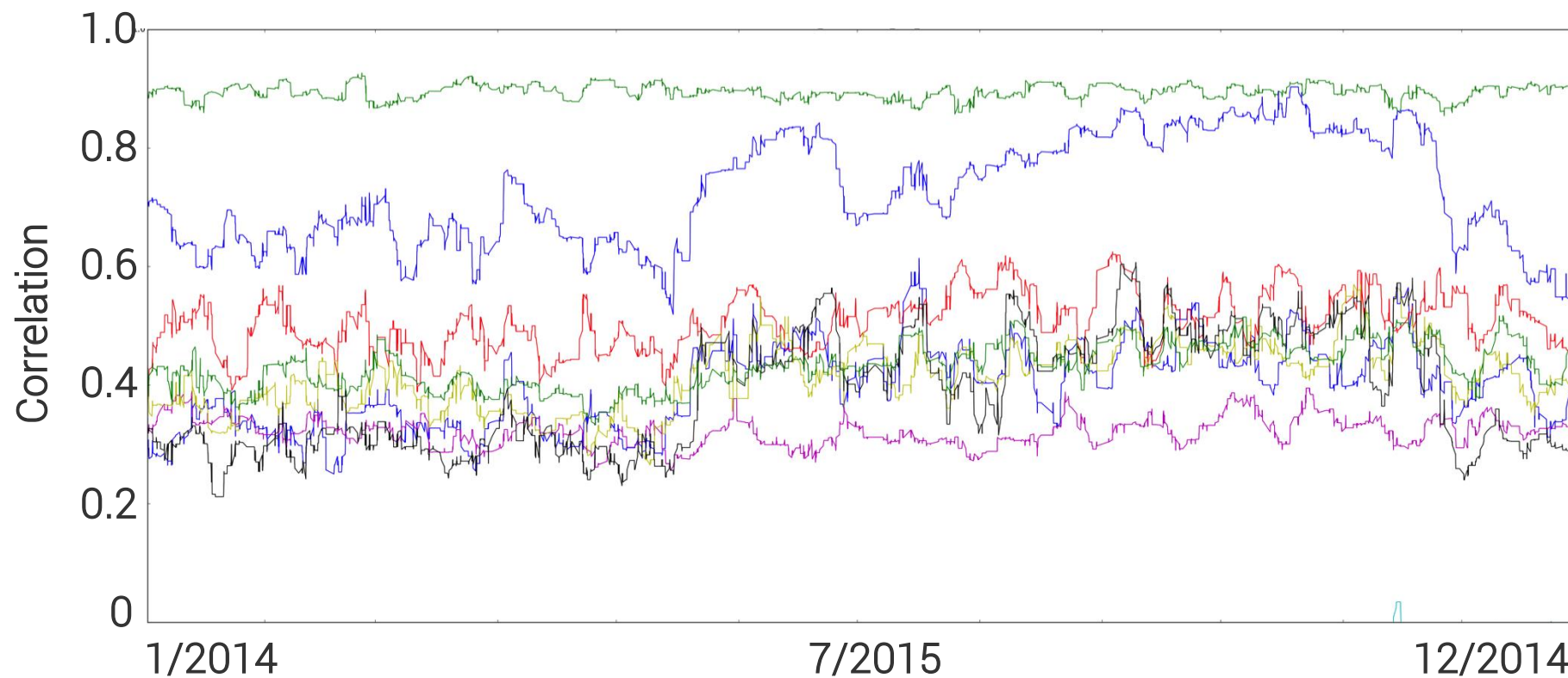
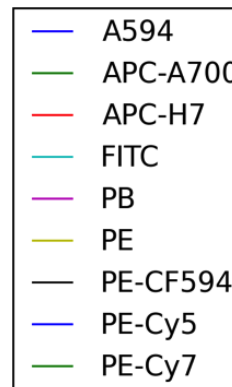


# Spillover - Crosslaser





# Spillover is not constant



APC, Myeloid 1, CD34

# How do I avoid diagnostic errors?

- Under Calls
  - » Flow is an ancillary study...
  - » “Its not your fault” – Sampling Issues
  - » “Its entirely your fault” – Perceptual Issues
- Over Calls
  - » Is that abnormal population real?

# Flow is an ancillary study...

- HG follicular lymphoma vs DLBCL
- Peripheral blood contamination vs tissue involvement

# Sampling Issues

- A single follicle center can be light chain skewed...
- Plasma cell myelomas are patchy
- Poisson Statistics in paucicellular samples
  - » Population SD =  $\sqrt{\# \text{ of } \textit{positive events}}$
  - » Precision drops as the number goes down!

# Real vs Fake Populations

- Non specific binding of antibodies
  - » Isotype controls
  - » FMO controls
- Compensation issues

# Why are isotype controls generally stupid?

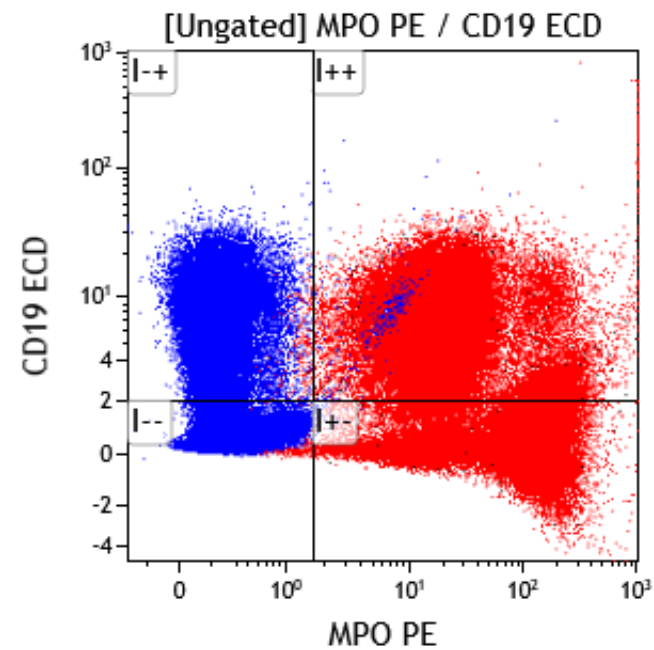
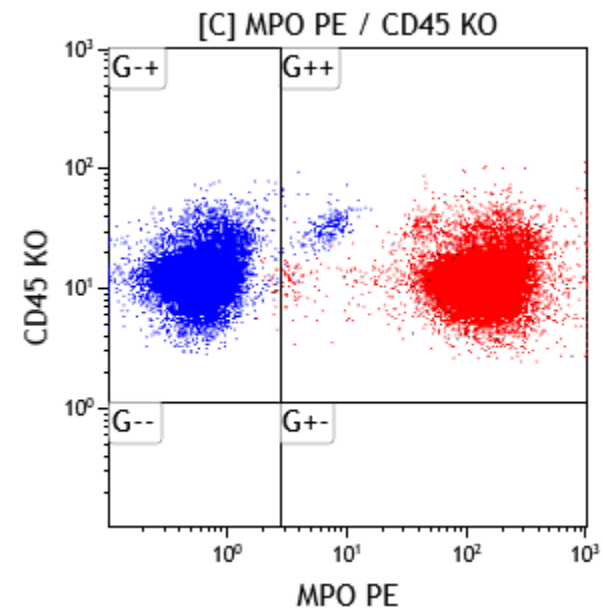
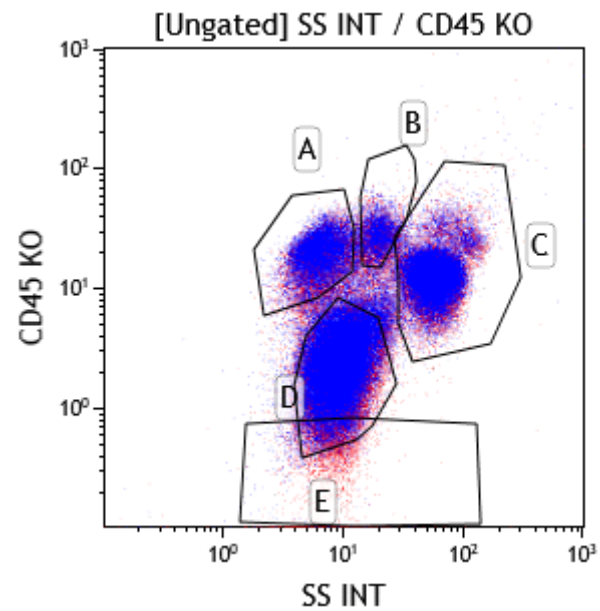
## PRO

- A non-specific antibody to measure Fc interactions
- Gives you a 'negative' population to work with

## CONS

- Its not really the same antibody
  - » There is no such thing as a true isotype
- False sense of security
- Doesn't solve problems about autofluorescence



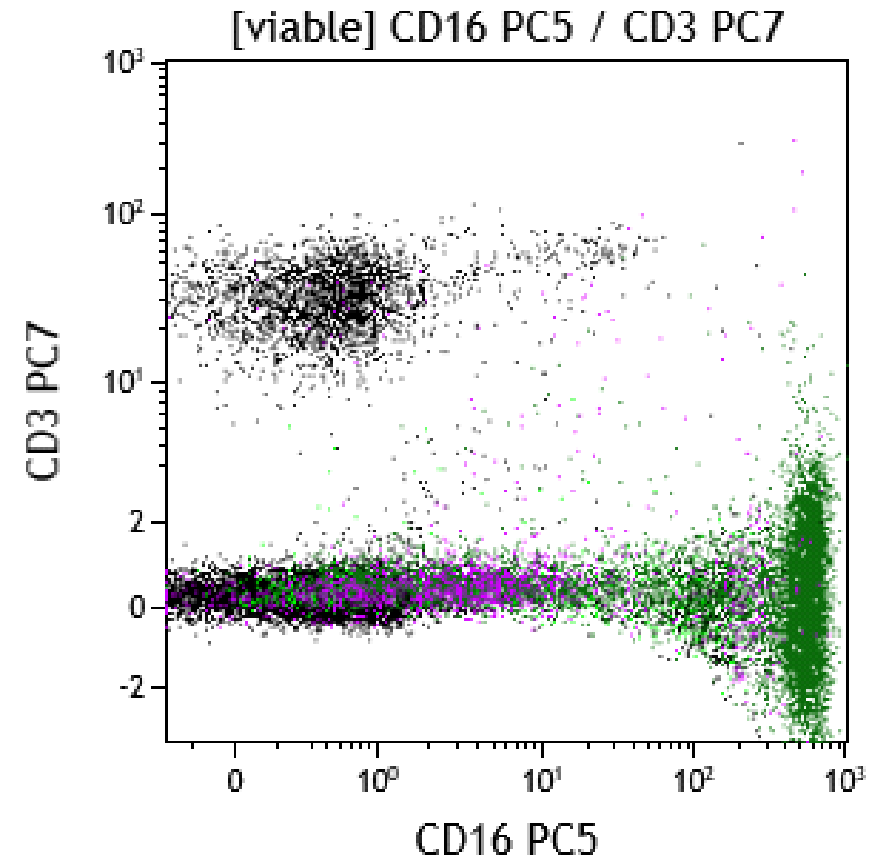
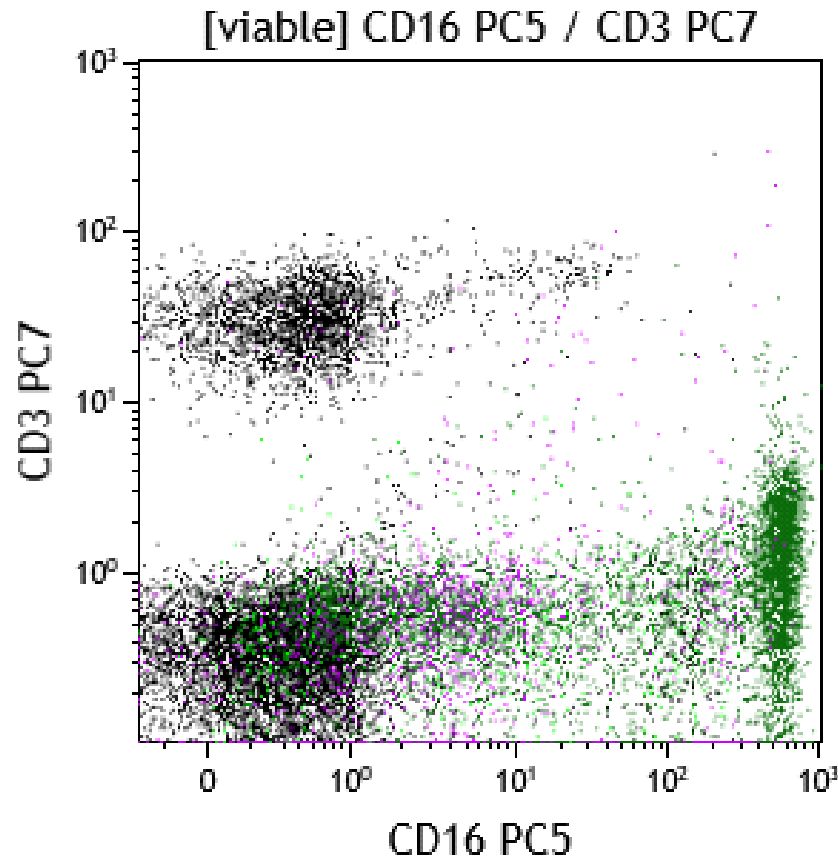


# Isotype controls (con't)

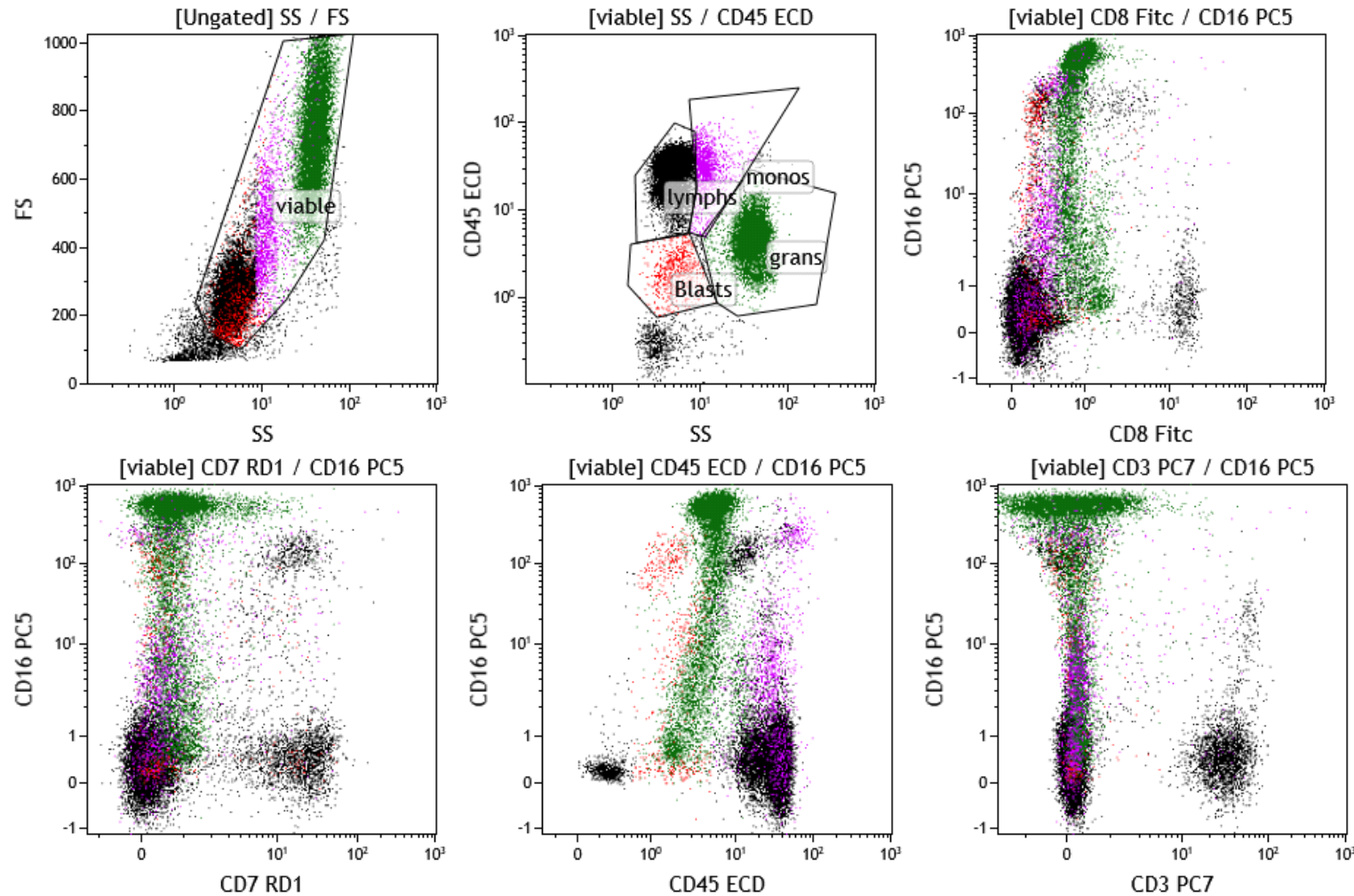
Rule: Don't use them

- » In the modern era of specific monoclonal antibodies its not needed
  - » Autofluorescence should be measured by FMO control
- 
- May have applications in the monocytes, some immune subsets (Fc binding) and cytoplasmic reagents (neo-epitope formation?)

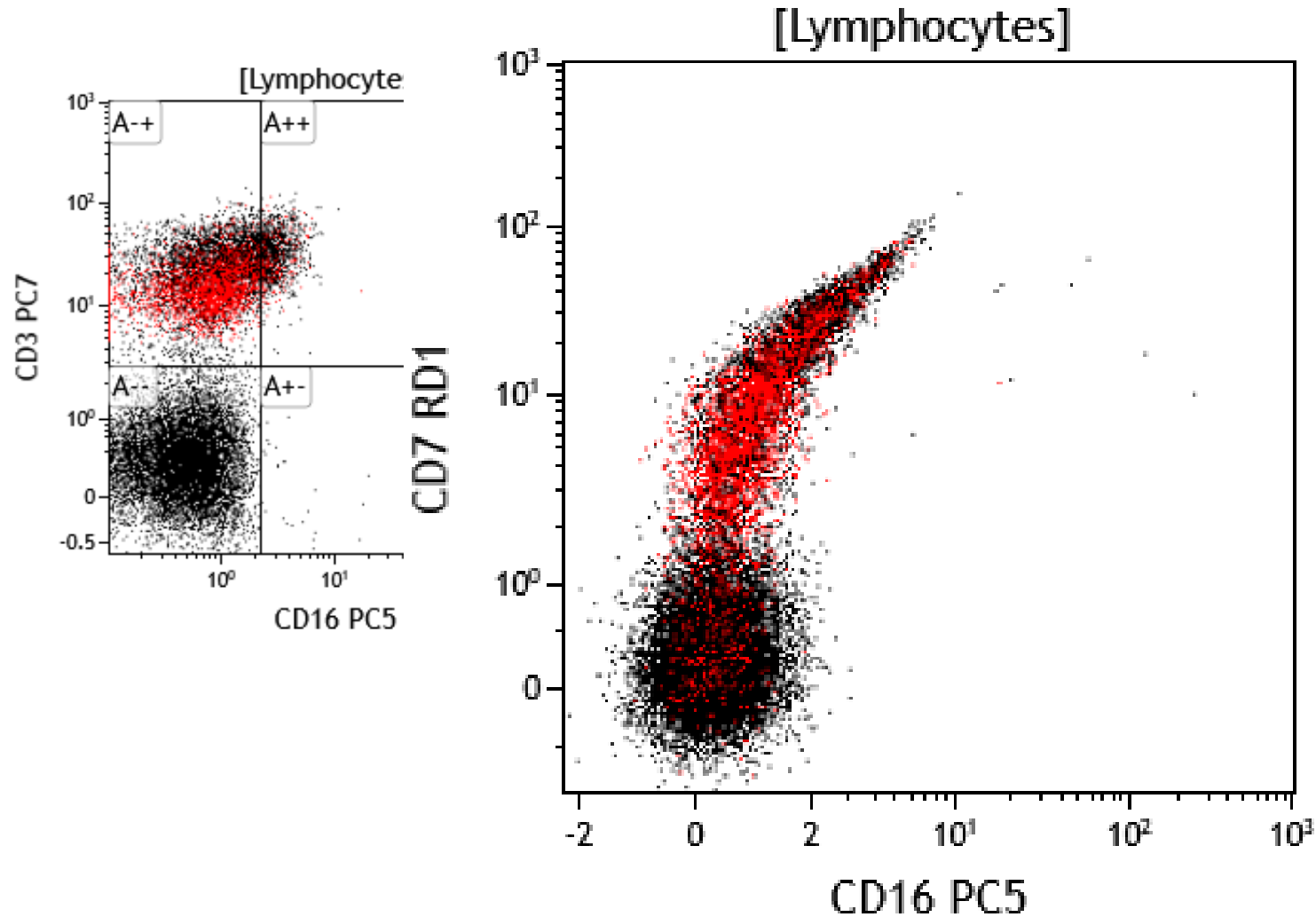
# Biexponential/Logicle Display

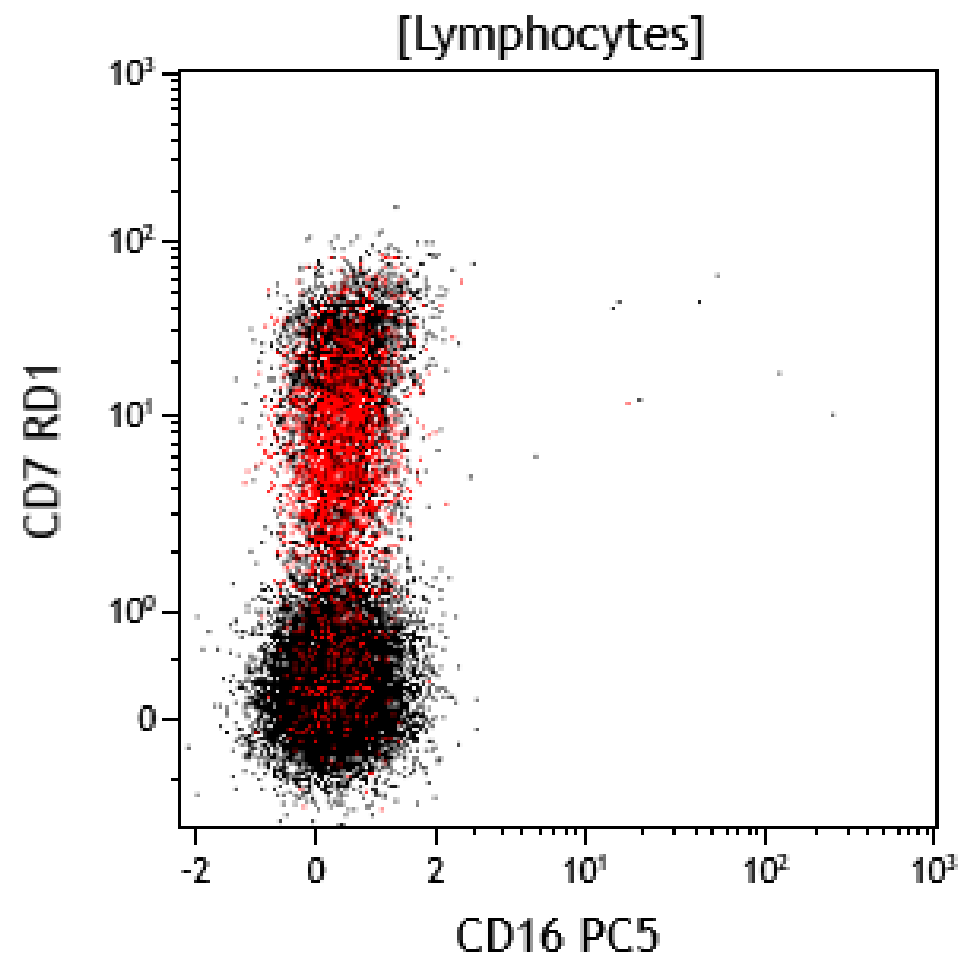


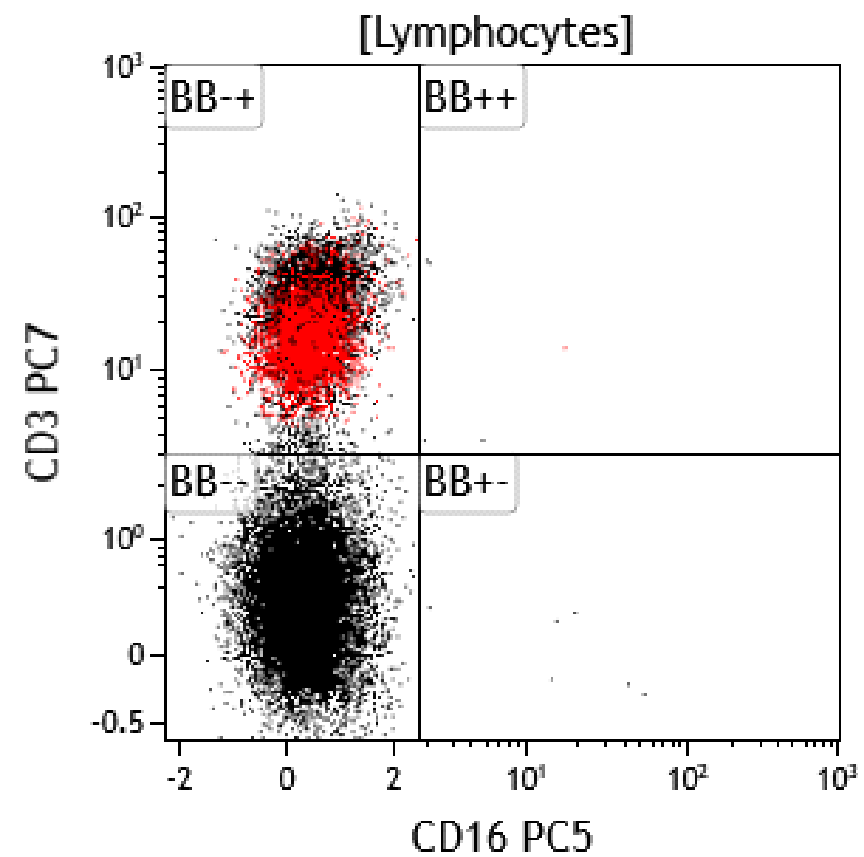
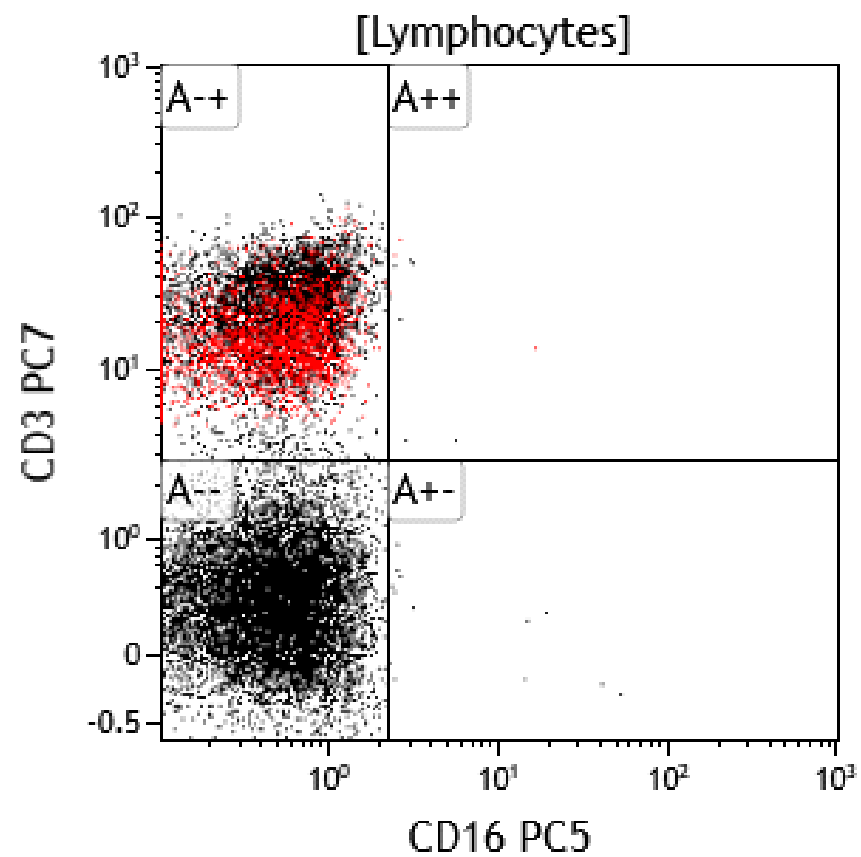
# Issues with high expressors



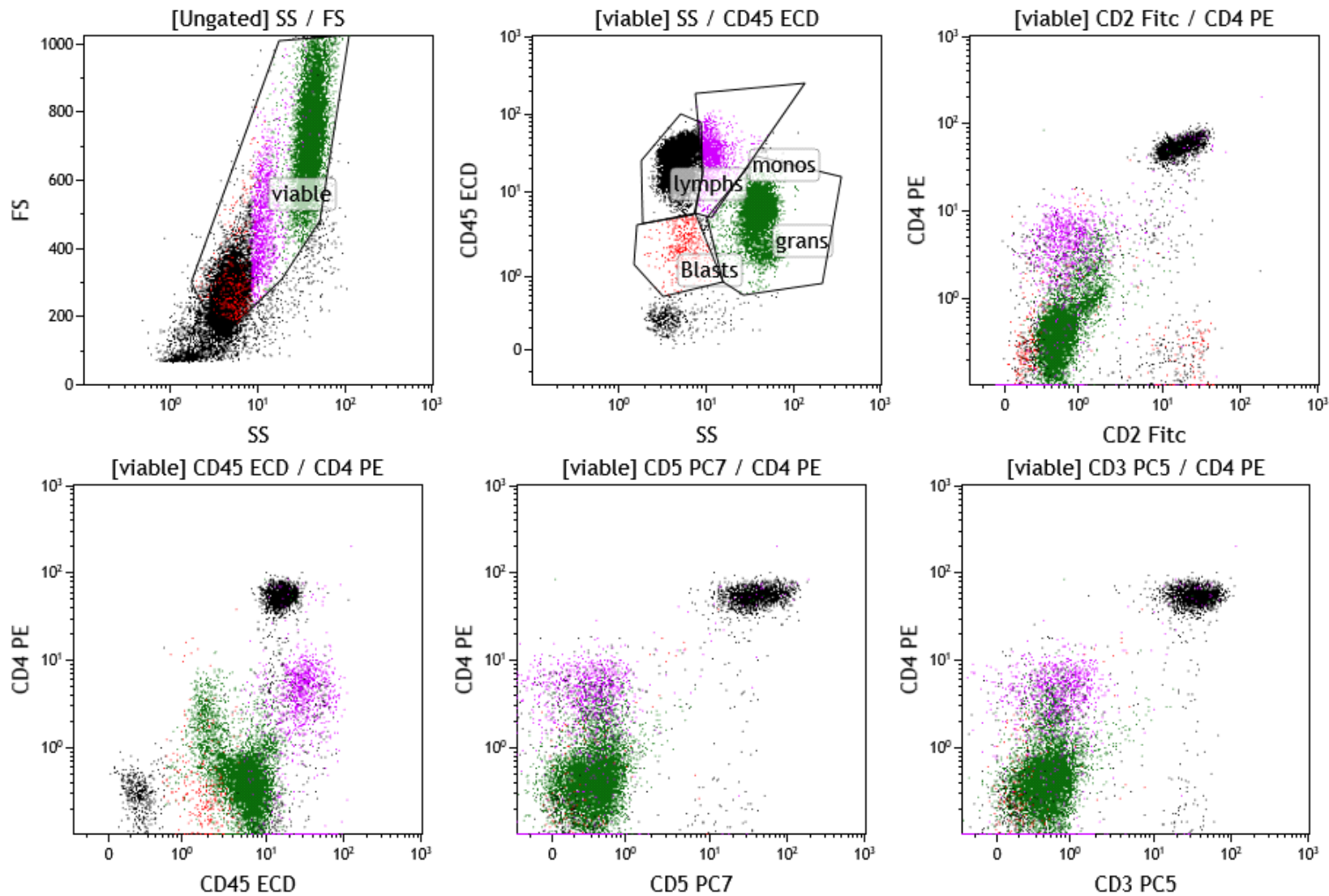
# Comp issues are hard to see





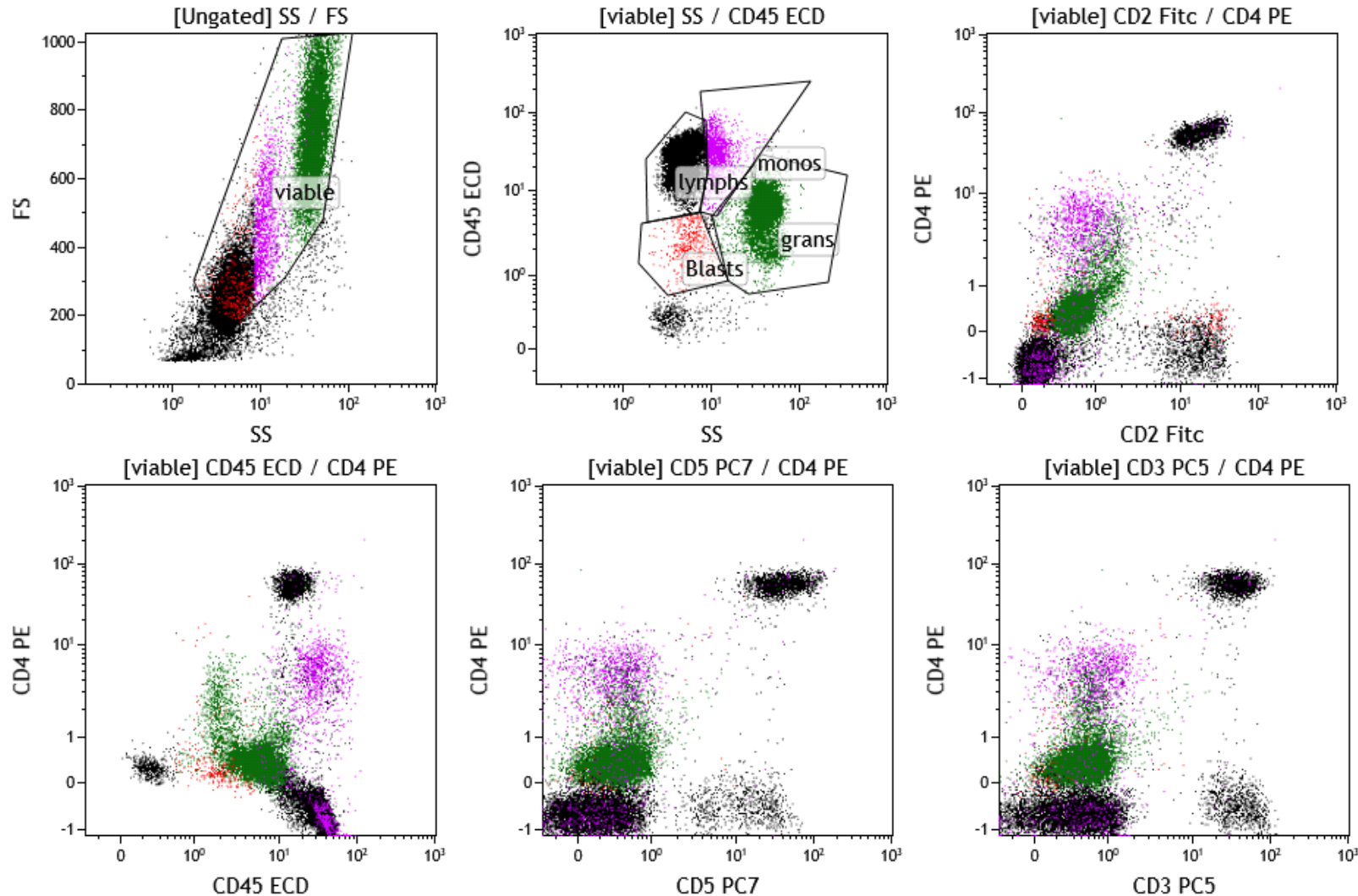


# Where did my CD4<sup>+</sup> T cells go?

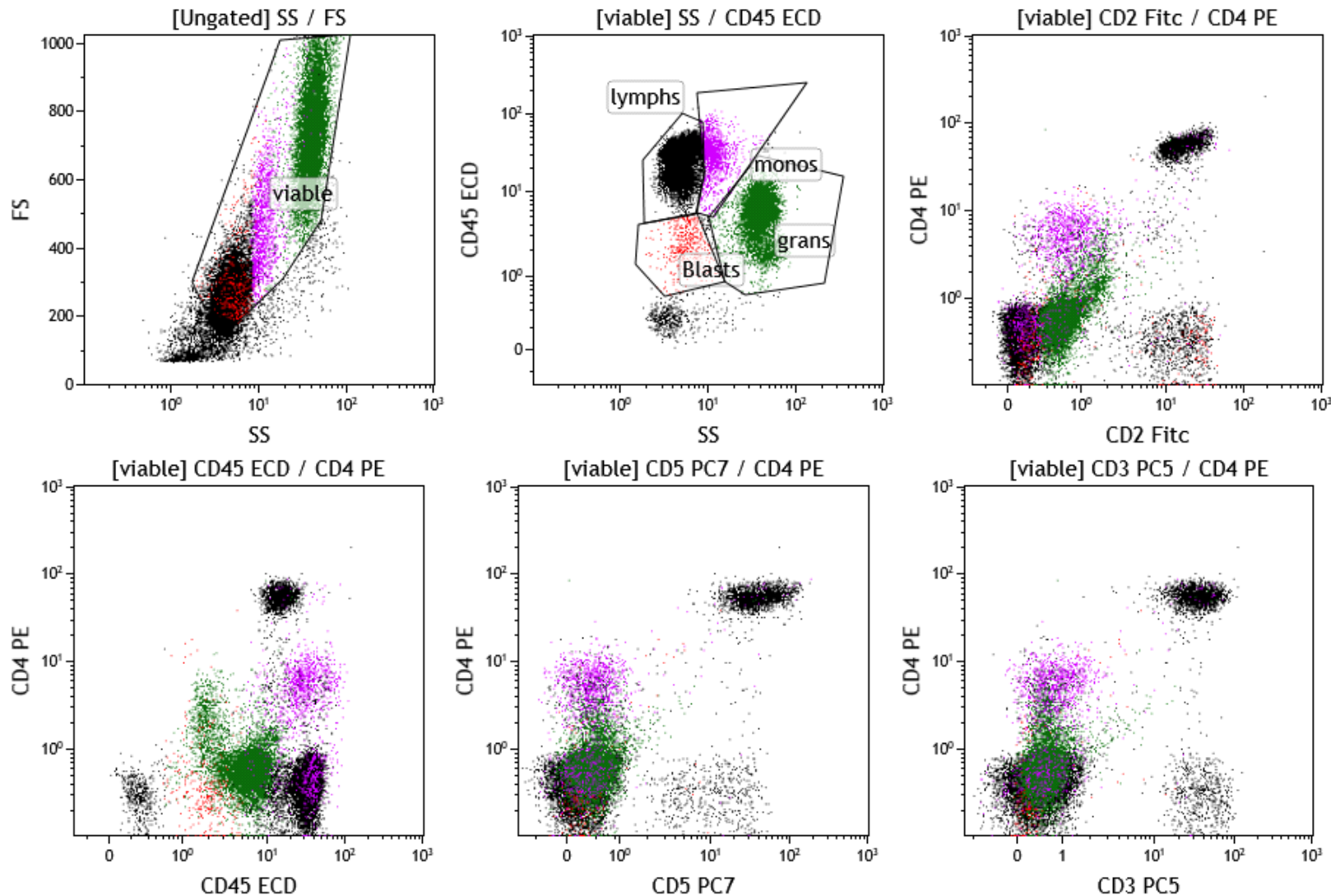




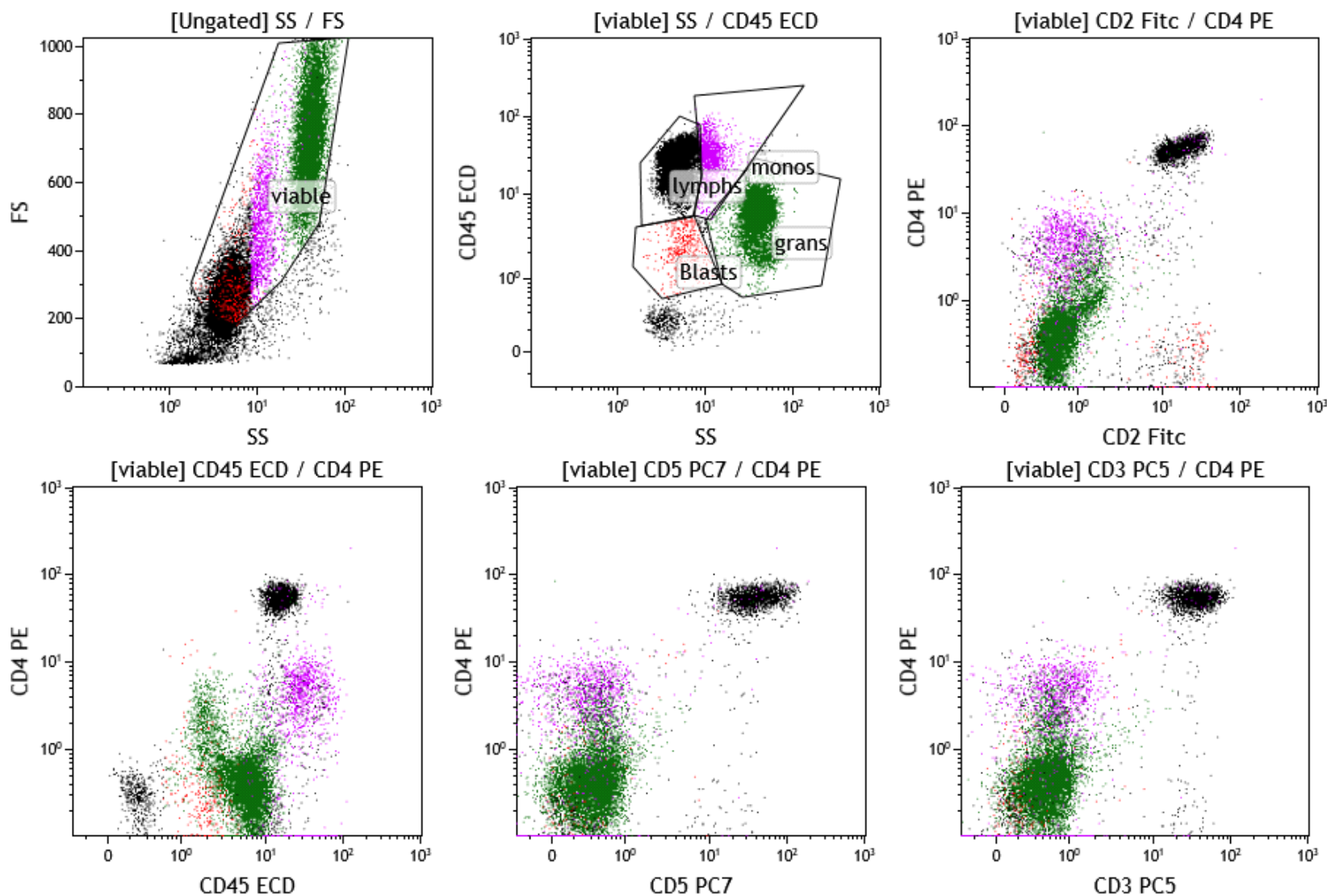
# Oh there they are...



# What it should look like...



# Compare and Contrast...



# Schlemiels and Schlimazels

- Fluorophores are like people:
  - » Some fluorophores spill over into other fluorophores
  - » "The *schlemiel* spills his soup on the *schlimazel*."
- Its important to know who is who
  - » Design around the issue
  - » Recognize the artifact
  - » Adjust the spillover estimate

Spillover (%)										
	FL1	FL2	FL3	FL4	FL5	FL6	FL7	FL8	FL9	FL10
FL1		1.10	0.40	0.30	0.30	0.10	0.30	0.30	0.20	0.70
FL2	22.00		9.60	3.30	2.70	0.10	0.20	0.00	0.10	0.50
FL3	8.90	49.10		1.60	1.00	0.10	0.20	0.30	0.10	0.30
FL4	0.80	5.30	17.70		0.10	0.30	0.80	0.30	0.10	0.00
FL5	0.00	1.00	2.90	35.50		0.00	0.80	2.00	0.00	0.00
FL6	0.00	0.10	0.20	1.70	0.10		10.20	7.50	0.10	0.20
FL7	0.00	0.00	0.00	14.10	0.30	19.30		6.60	0.00	0.00
FL8	0.00	0.00	0.00	9.70	10.00	8.30	47.70		0.00	0.00
FL9	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00		4.40
FL10	4.40	4.70	1.10	0.80	0.70	0.70	1.10	0.00	8.70	

	FITC	PE	ECD	PE-CY5.5	PE-CY7	APC	APC-A70 0	APC-A75 0	BV 421	KO
Spillover (%)	FL1	FL2	FL3	FL4	FL5	FL6	FL7	FL8	FL9	FL10
FL1		1.10	0.40	0.30	0.30	0.10	0.30	0.30	0.20	0.70
FL2	23.00		9.60	3.30	2.70	0.10	0.20	0.20	0.10	0.50
FL3	8.90	44.20		1.60	1.00	0.10	0.20	0.30	0.10	0.30
FL4	0.50	4.10	15.40		0.10	0.30	0.80	0.20	0.00	0.00
FL5	0.00	0.60	2.90	35.50		0.00	0.80	2.00	0.00	0.00
FL6	0.10	0.10	0.20	1.70	0.10		11.20	10.50	0.10	0.20
FL7	0.00	0.00	0.00	14.10	0.30	19.30		6.20	0.00	0.00
FL8	0.00	0.00	0.00	9.70	10.00	8.30	47.70		0.00	0.00
FL9	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00		4.40
FL10	4.40	4.70	1.10	0.80	0.70	0.70	1.10	0.00	8.70	

# Tandem Breakdown

Tandems – the solution to and cause of all life's problems.

- Ab – Donor – Acceptor
- Antibody with a tandem looks like its donor
- CD34 APC-A700 -> acts like CD34 APC

What happens if you have CD7 on APC?

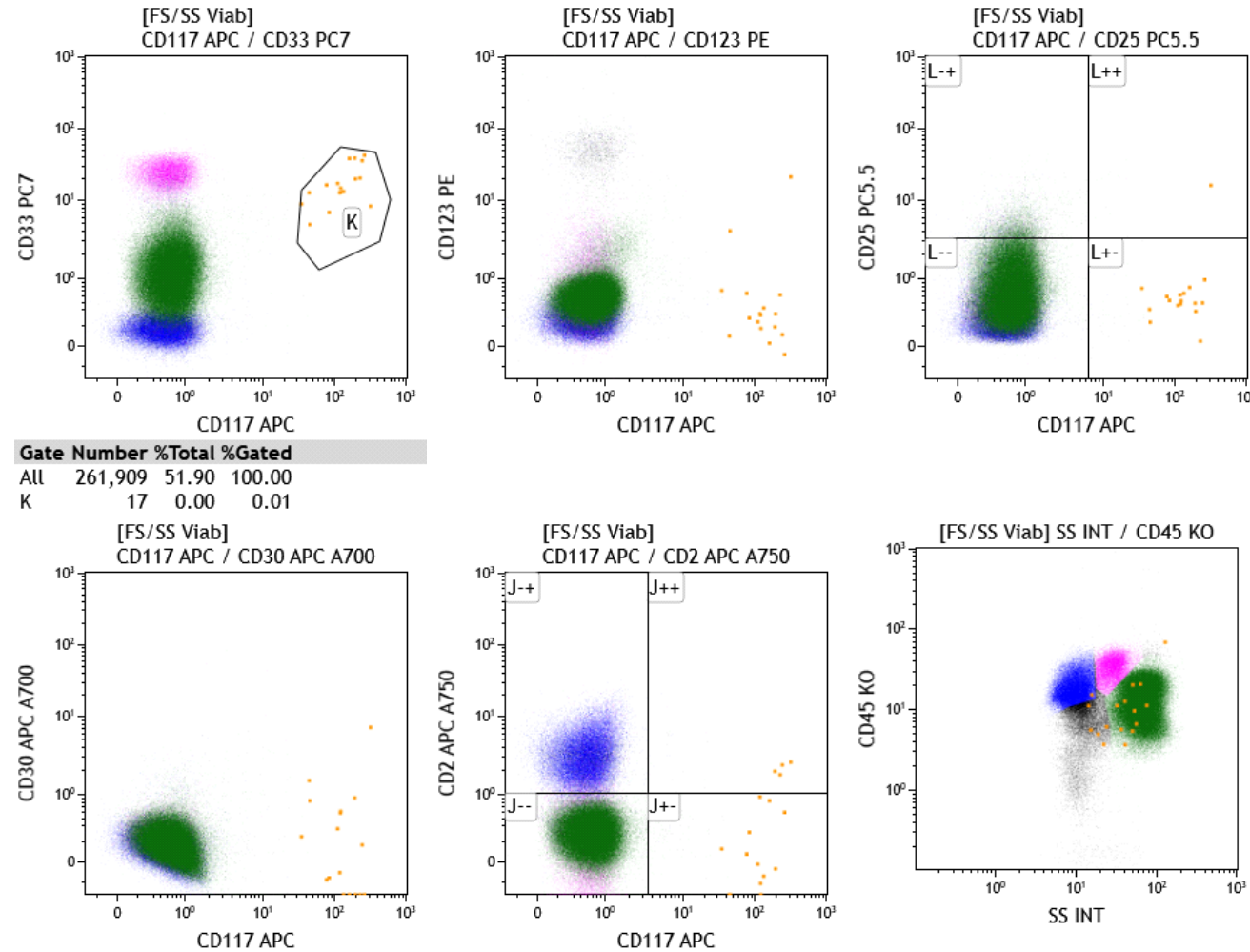


# Panel Design

- What antibody and fluorophore goes where?
- Strong antibody and weak fluorophore (vice versa)
- Strong antibodies/fluorophore can lead to spillover issues, increased “negative” control
- Fit-for-purpose

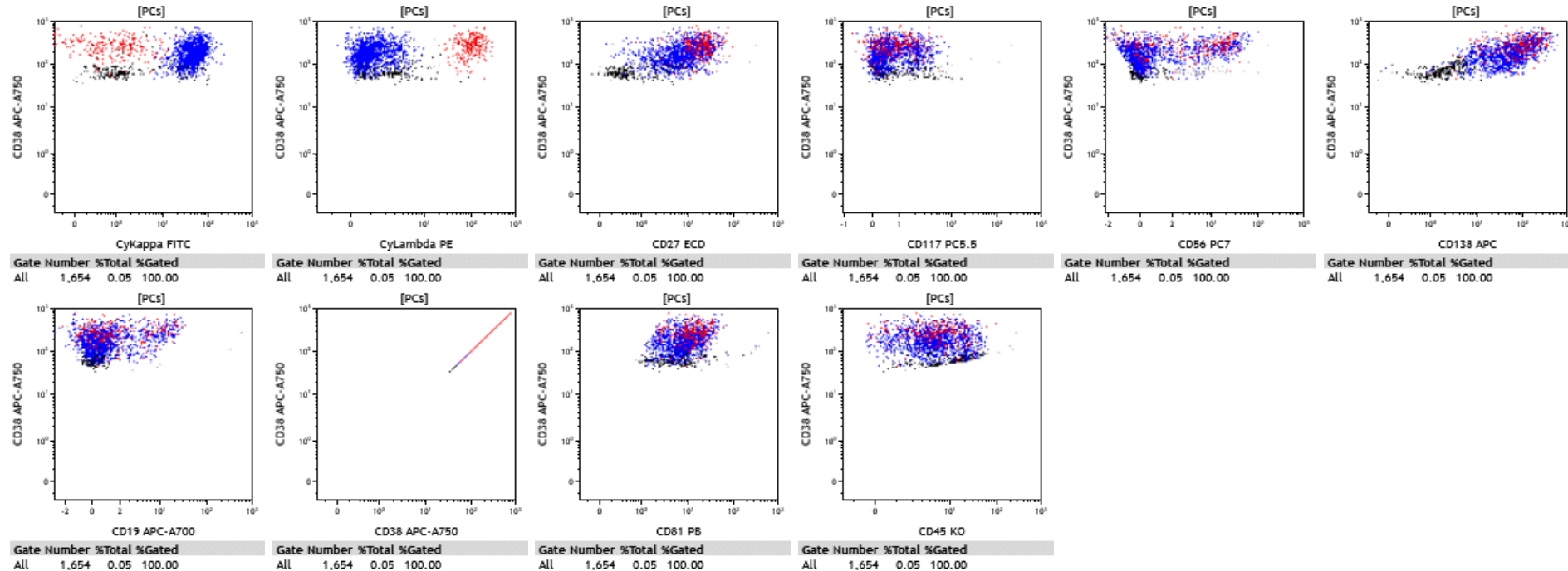


# Panel Design Example/Considerations



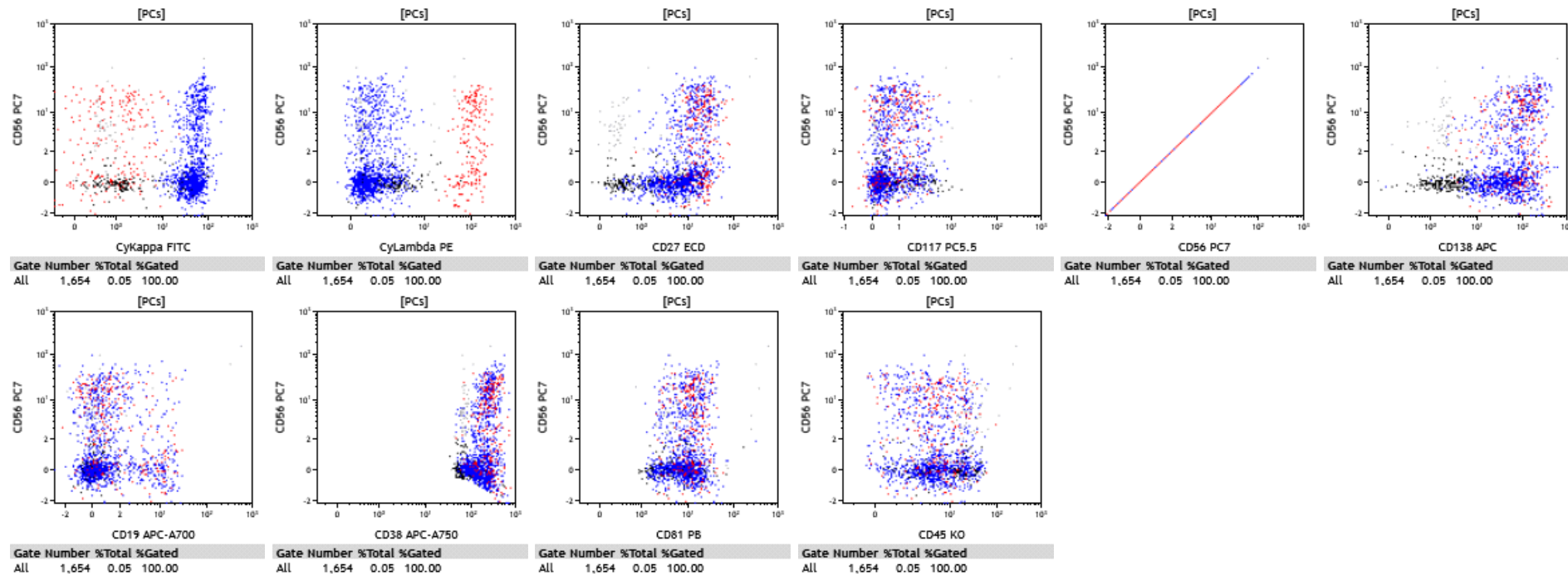
# Panel Design

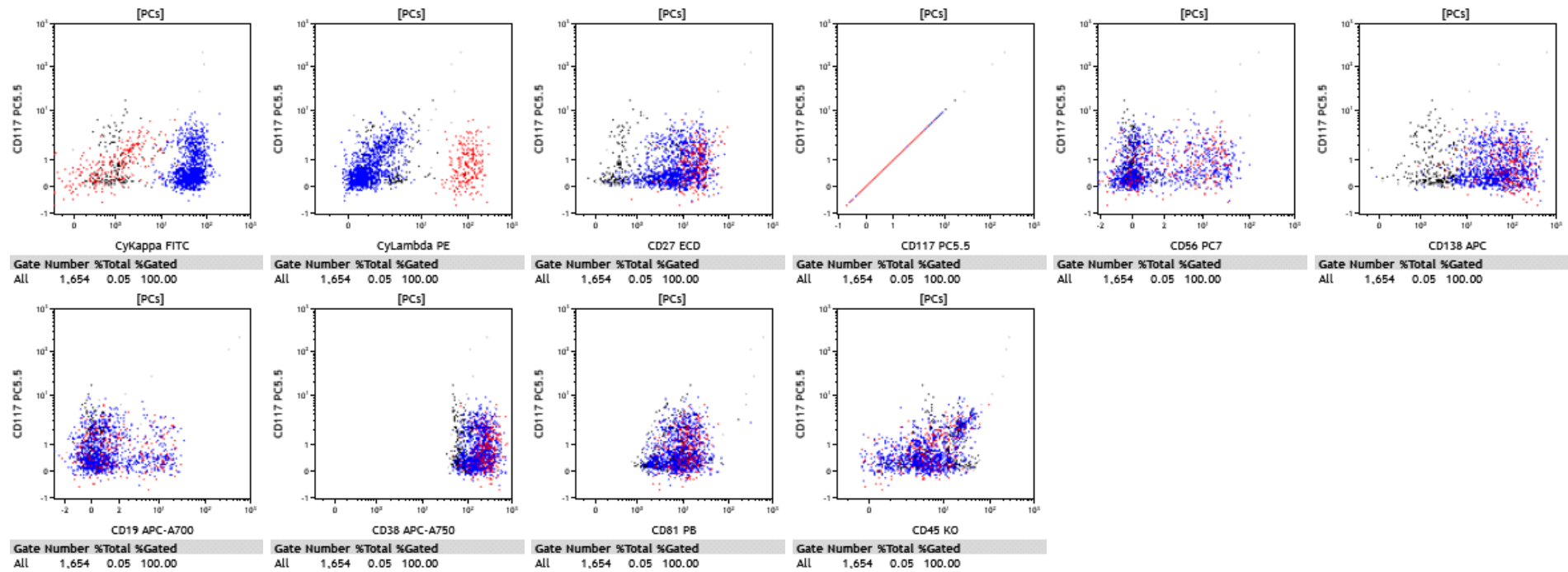
## Example/Considerations



# Additional things to care about

- CD56 and CD117





# Summary

- Technical artifacts need to be recognized
  - » Compensation
  - » Breakdown
  - » Positivity Discrimination
- You can decrease but never eliminate artifacts
  - » Schlemiels are always abound
  - » But bright schlimazels can't be spilled on effectively



*ARUP is a nonprofit enterprise of the University of Utah and its Department of Pathology.*