

# Novel Approaches to ID Testing Using NGS-Based Metagenomics

Robert Schlager, MD, MPH

# Disclosures

## Commercial Interest

Roche Diagnostics

Roche Diagnostics

Hologic

Hologic

Epoch Biosciences

Sanofi Pasteur

IDbyDNA

## What was Received

Honorarium

Research Grants

Contract Research

Honorarium

Contract Research

Contract Research

Stock, Honorarium

## For What Role

Advisor

PI

PI

Advisor

PI

Co-PI

Co-Founder, CMO,  
Board Member

# Conventional Infectious Disease Testing

## Multimodal

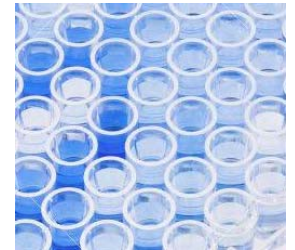
Culture



PCR

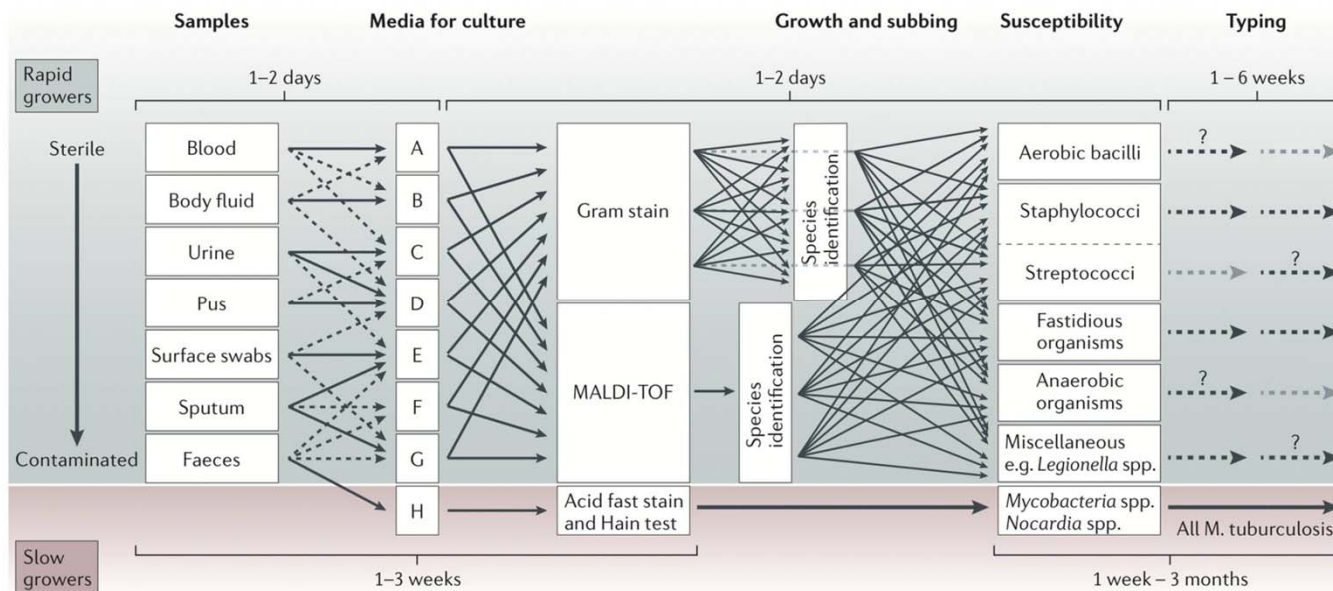


Ag Detection  
Serology



# Conventional Infectious Disease Testing

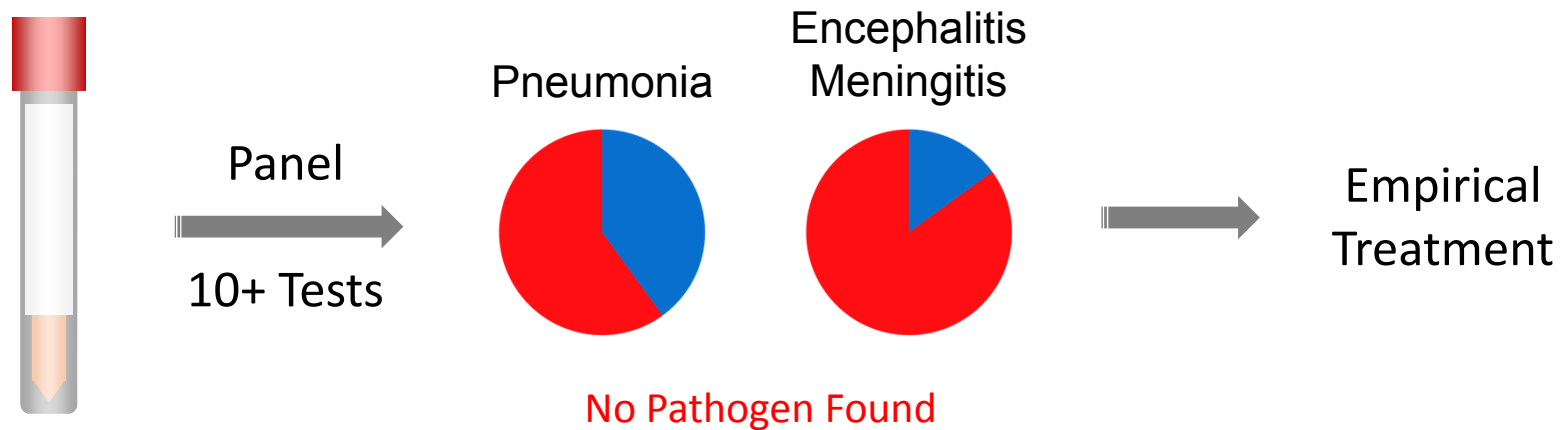
## Complex & Sequential Laboratory Workflows



Nat Rev Genet. 2012 Sep;13(9):601-12

# Limited Yield for Many Infectious Syndromes

Current tests often pathogen-specific, combined into panels



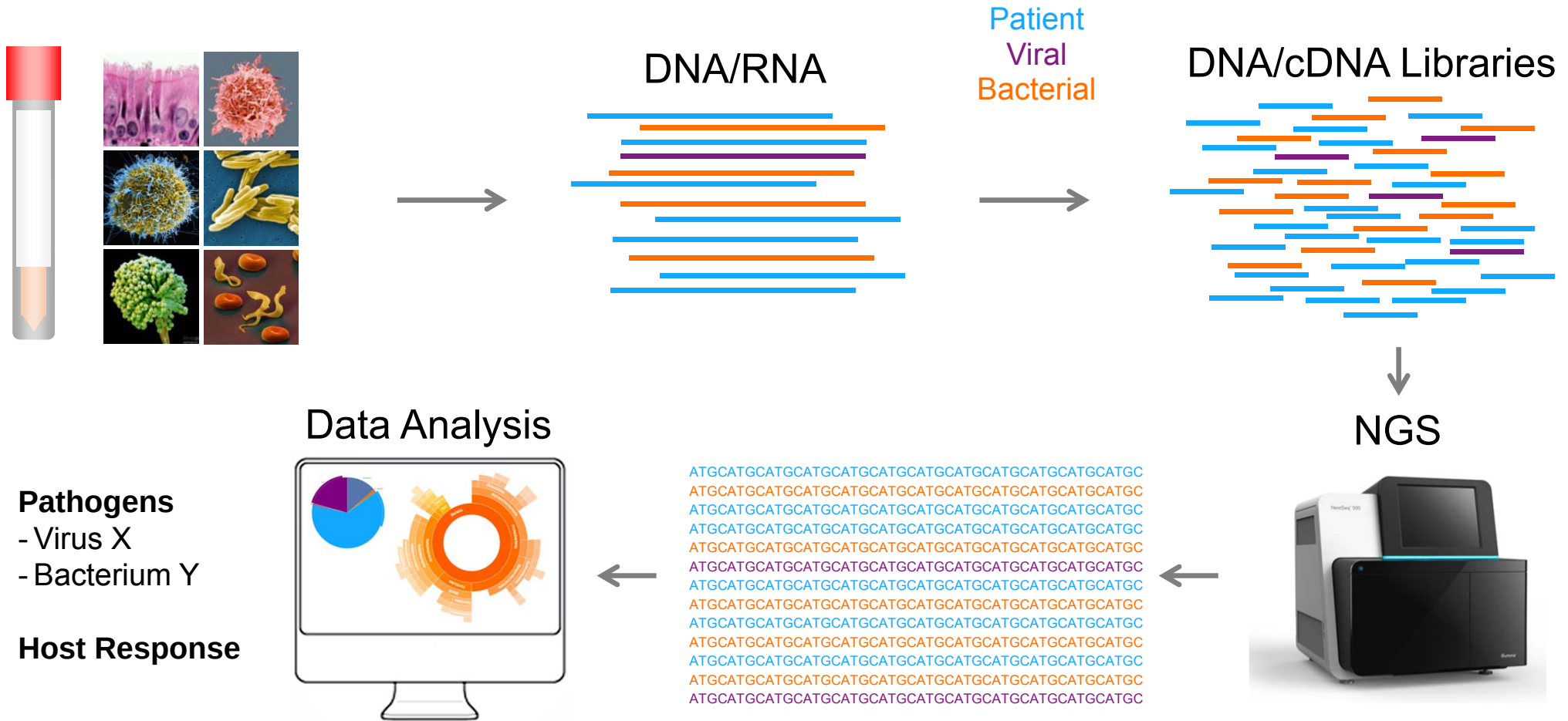
*Glaser, CID;43(12):1565*

*Freifeld, CID;52(4):e56*

*Murdoch, Arch Intern Med.;169(5):463*

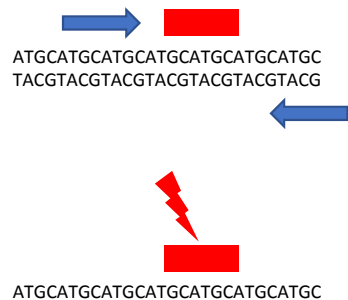
*Jain, NEJM;373(5):415*

# Metagenomics – One-Stop Testing



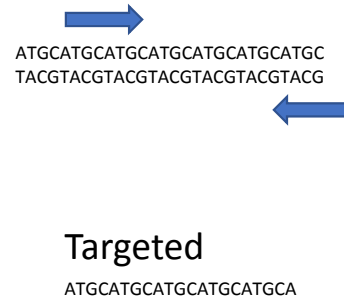
# NAAT, Targeted Sequencing, Unbiased Metagenomics

## NAAT



- Sensitive
- Specific
- Fast
- Can only detect targeted pathogens

## Targeted Sequencing



- Broader scope
- Hypothesis driven
- Classification limited due to genetic snapshot

## Unbiased Metagenomics

### Unbiased

ATGCATGCATGCATGCATGCATGC  
TGATGCATGCATGCATGCATGCATGCA  
GCATGCATGCATGCATGCATGCATGCAT  
CATGCATGCATGCATGCATGCATGCATG  
ATGCATGCATGCATGCATGCATGCATGC  
TGATGCATGCATGCATGCATGCATGCA  
GCATGCATGCATGCATGCATGCATGCAT  
CATGCATGCATGCATGCATGCATGCATG

- Broadest method
- Hypothesis-free
- Unlimited multiplexing
- All classes of pathogens

Why Do I Think This Is Worthwhile?



# First Applications: Pathogen Discovery

## Three SOT recipients (same donor)

- Fever, encephalopathy
- Died 4 to 6 weeks after transplantation

## Donor

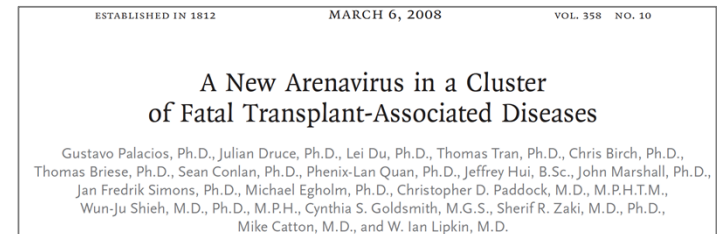
- Cerebral hemorrhage 10 d after travel to rural areas of Eastern Europe

## All tests negative

- Bacterial and viral cultures, pathogen-specific PCR's
- Viral and panmicrobial oligonucleotide microarray

## Detected by metagenomic cDNA sequencing

- 454 sequencing (~100,000 total sequences)
- LCMV-like arenavirus (protein level)



*Palacios et al., NEJM. 2008 Mar 6;358(10):991-8*

# From Case Reports to Analytical Performance and Diagnostic Yield

NP Swabs (n=109)



PCR Panel (RVP)



I. Positives (n=42)

II. Unselected (n=67)

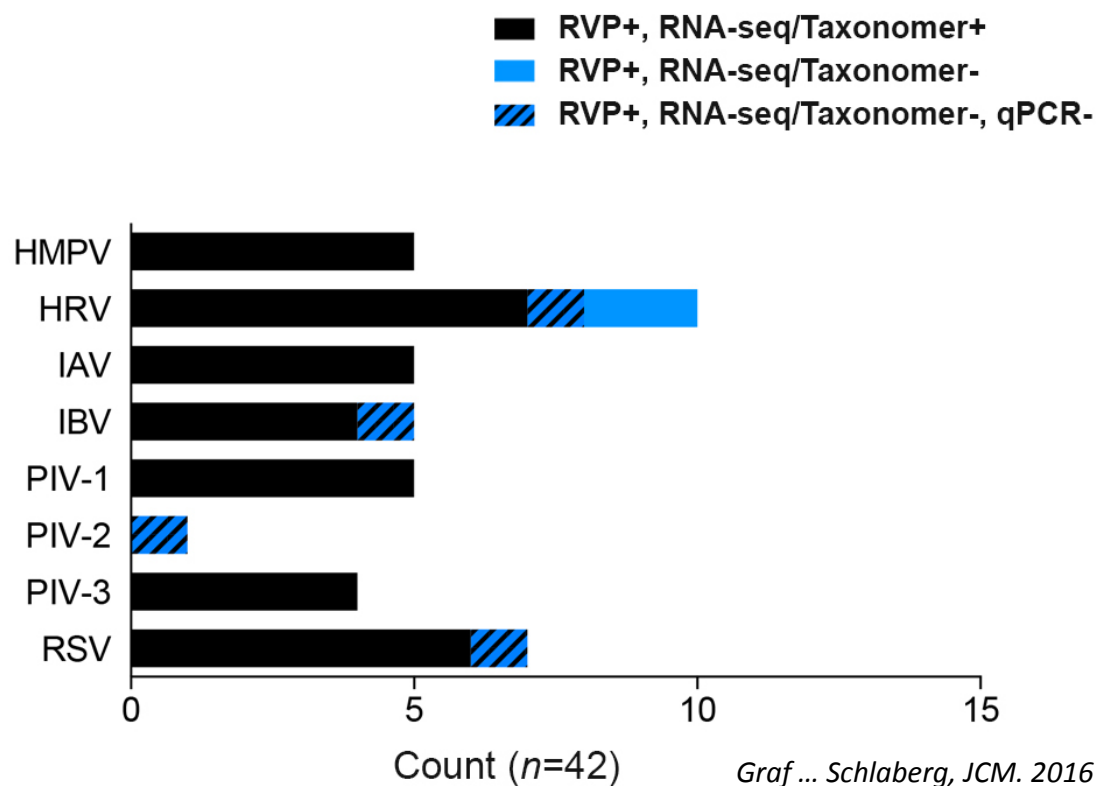
RNA-seq (HiSeq)



5-10 M reads/specimen

## >95% Positive Agreement with PCR Panel

NP/OP Swabs (Viral Pathogens), RNA-seq & Taxonomer



Graf ... Schlager, JCM. 2016 Apr;54(4):1000-7

NP Swabs (n=109)



PCR Panel (RVP)



I. Positives (n=42)

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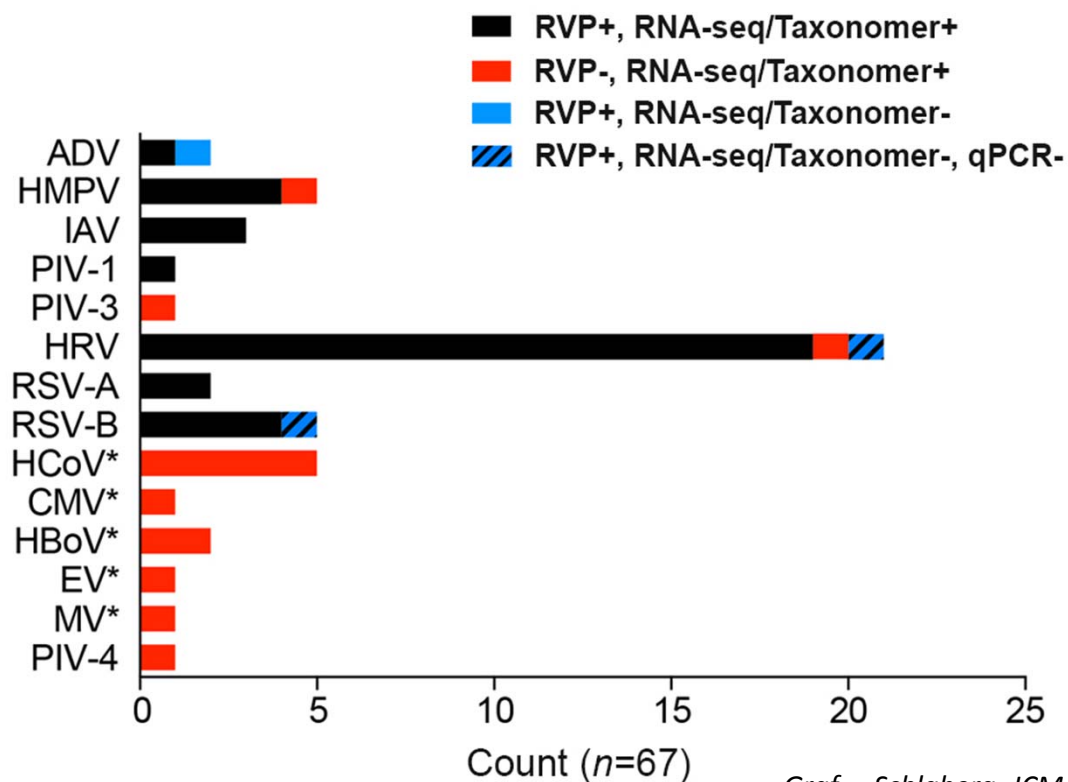
RNA-seq (HiSeq)



5-10 M reads/specimen

## >90% Overall Agreement with PCR Panel

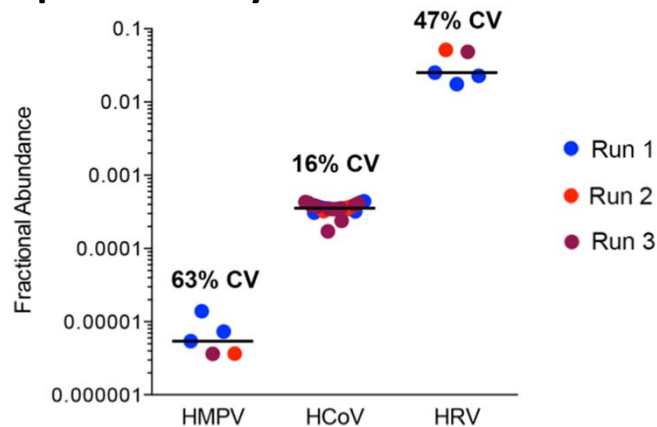
NP/OP Swabs (Viral Pathogens), RNA-seq & Taxonomer



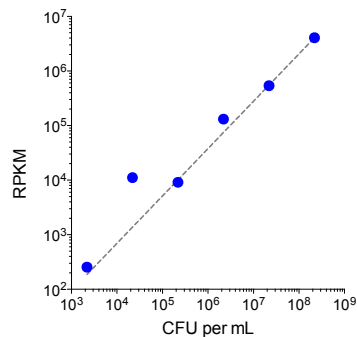
Graf ... Schlager, JCM. 2016 Apr;54(4):1000-7

# Reproducible & Quantitative Results

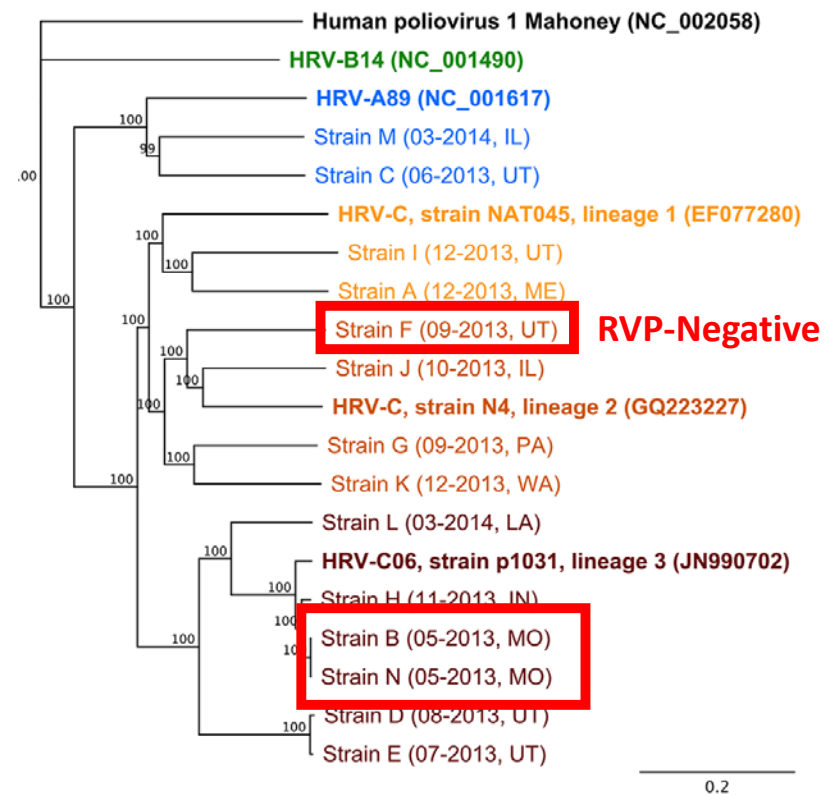
## Reproducibility



## Bacterial Load *H. influenzae*

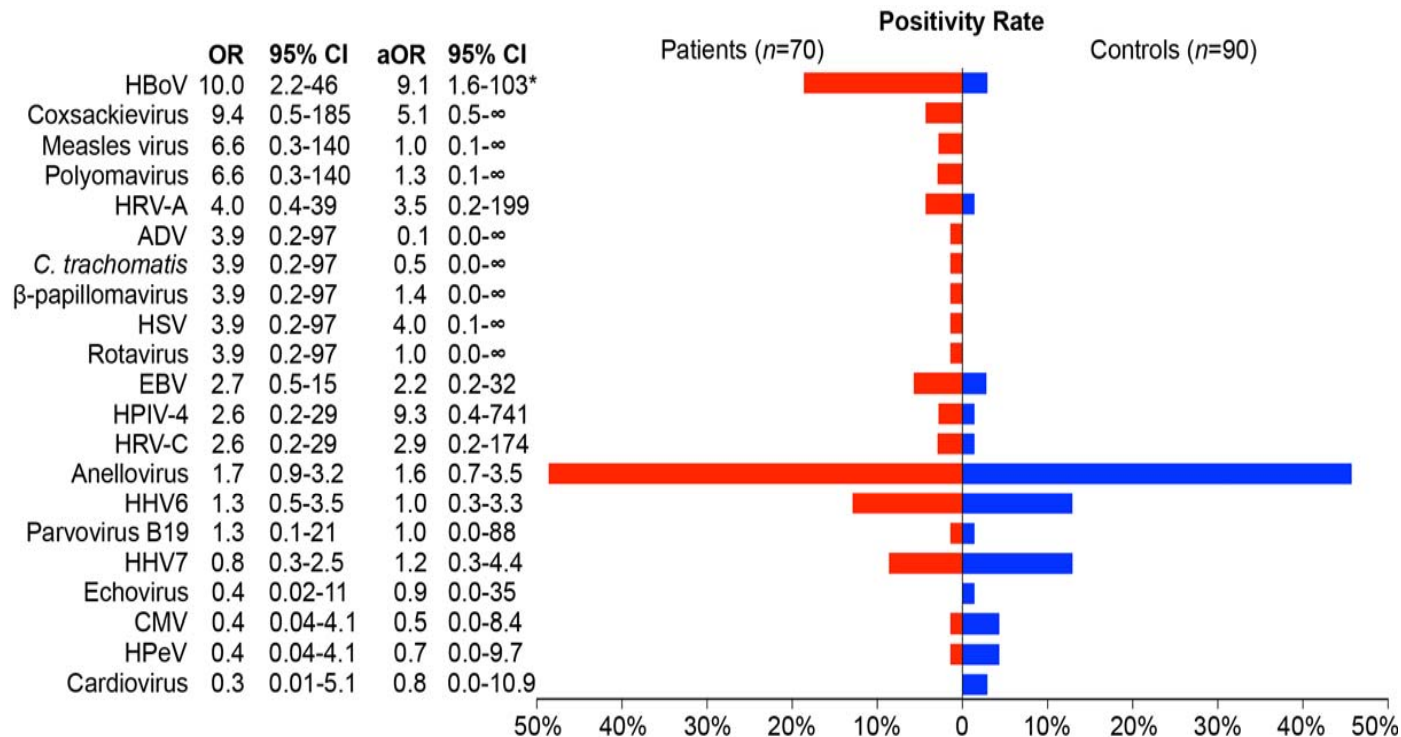


## Sequence Agnostic



Graf ... Schlager, JCM. 2016 Apr;54(4):1000-7

# CAP of Unknown Etiology: Putative Pathogens in 30% NP/OP Swabs (Viral Pathogens), RNA-seq & Taxonomer and Panviral Group PCR

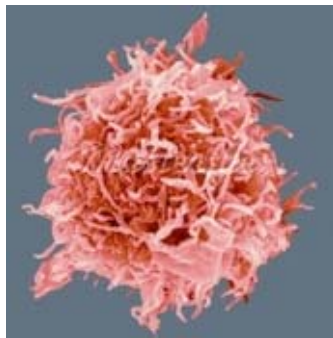


Schlager et al., JID;215(9):1407-1415

Can This Be Implemented In Routine Practice?

# Specimen-Dependent Performance (Depletion, Enrichment)

Abundant, Unwanted Host Nucleic Acid



>>



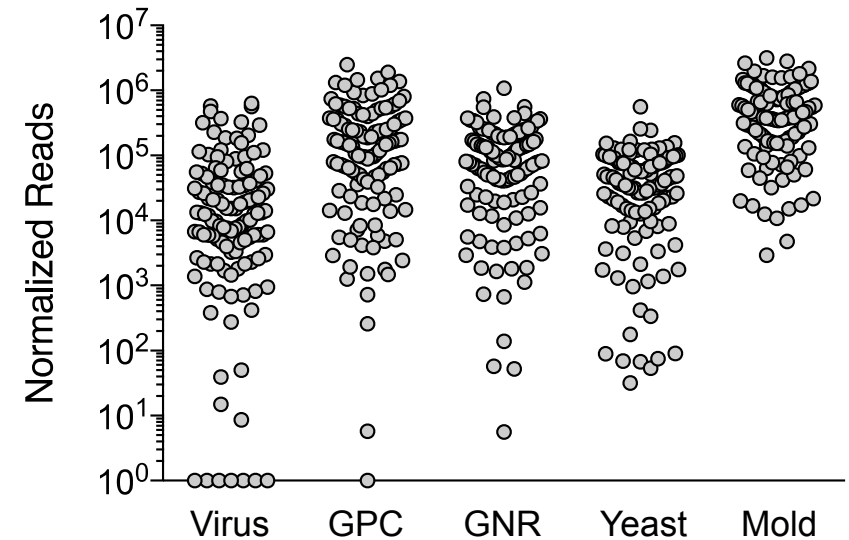
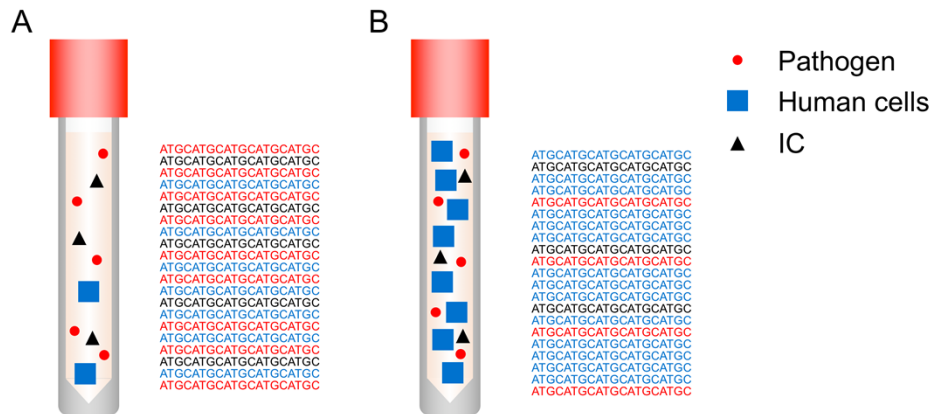


# Specimen-Dependent Performance (Depletion, Enrichment)

## Analytical sensitivity is variable

- Specimen composition/cellularity
- Sequencing depth
- Genome size

## Internal controls to monitor

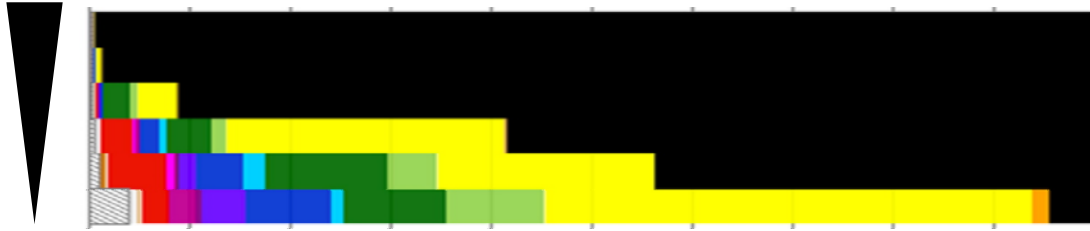


Schlager R, Chiu CY, Miller S, Procop G, Weinstock G, Arch Pathol Lab Med. 2017 Jun;141(6):776-786

# Contamination Risk (Bacteria), Need for Controls

Reagents contain microbial nucleic acid

- Low DNA Yield -> High Contamination Risk

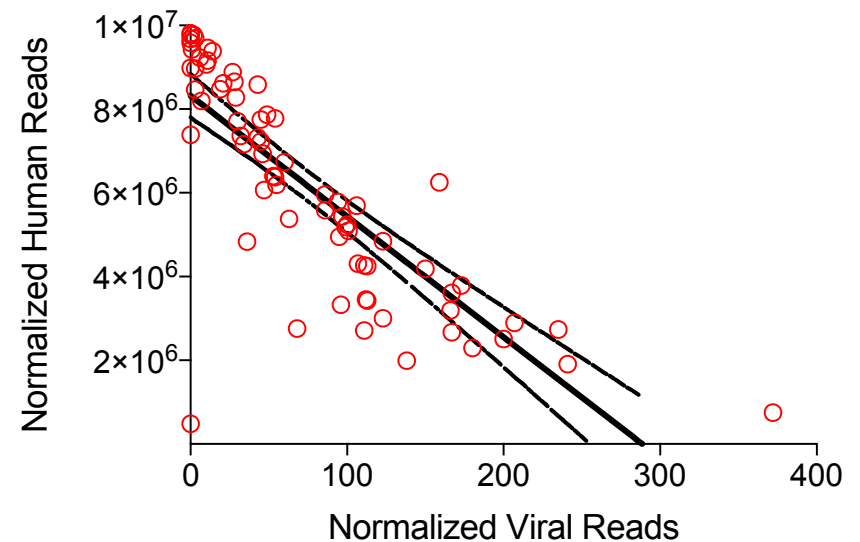
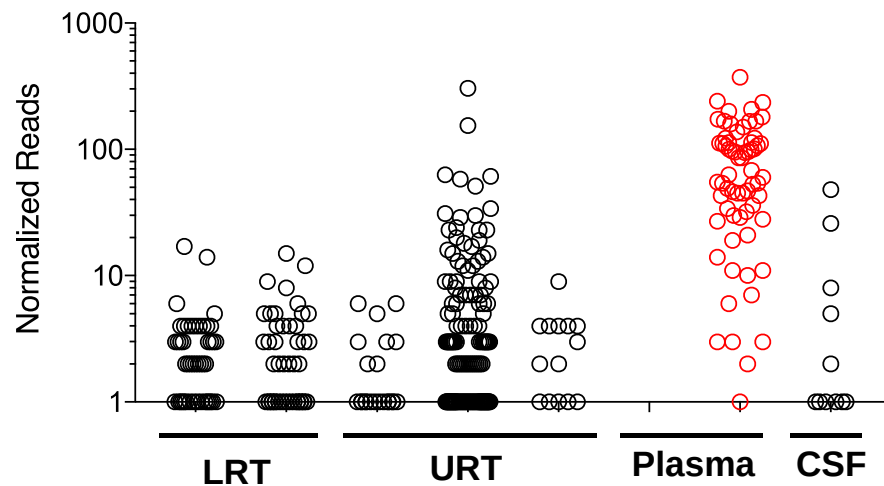


- Highly variable between lots/batches (many reagents, frequent lot changes)

Salter ... Walker, *BMC Biol.*;12:87  
Naccache S ... Chiu CY, *J Virol.*;87(22):11966-77  
Thoendel Patel, *J Clin Microbiol.* 2017 Mar 29

# Contamination Risk (Viruses), Need for Controls

## Kit & NA-Yield Dependent Detection



Salter ... Walker, *BMC Biol.*;12:87  
Naccache S ... Chiu CY, *J Virol.*;87(22):11966-77  
Thoendel Patel, *J Clin Microbiol.* 2017 Mar 29

# Laboratory Workflow Complex to QC

## **Guaranteeing Consistent Performance**

- Involves dozens of steps
- Several steps involve >1 kits/reagents
- Some kits involve reagents from different lots

## **Rapidly-evolving field**

- New, improved reagents & protocols
- Performance comparisons can be complex and expensive
- Update testing protocols frequently

# Rapid, Accurate Classification Needed

## Millions of DNA Sequences

[illegible]

## Millions of Microbial References

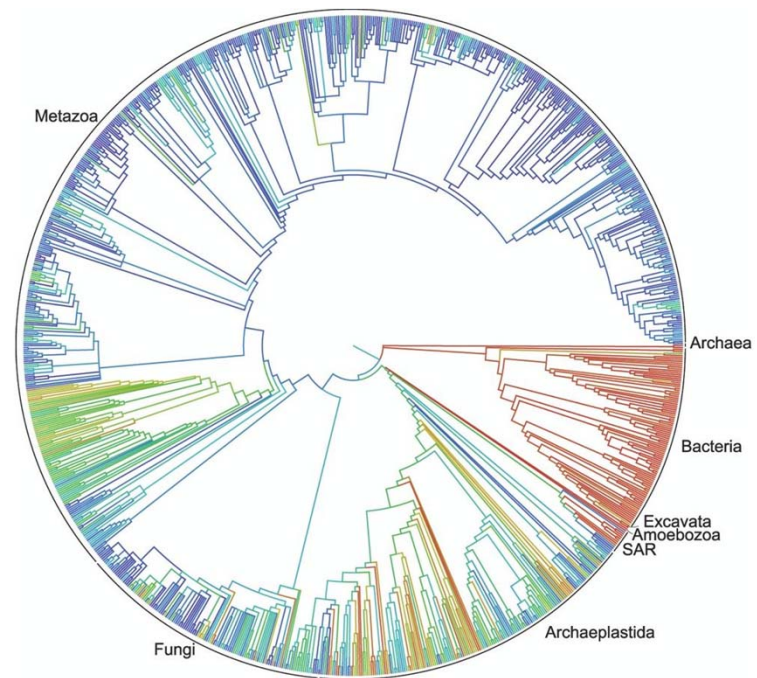
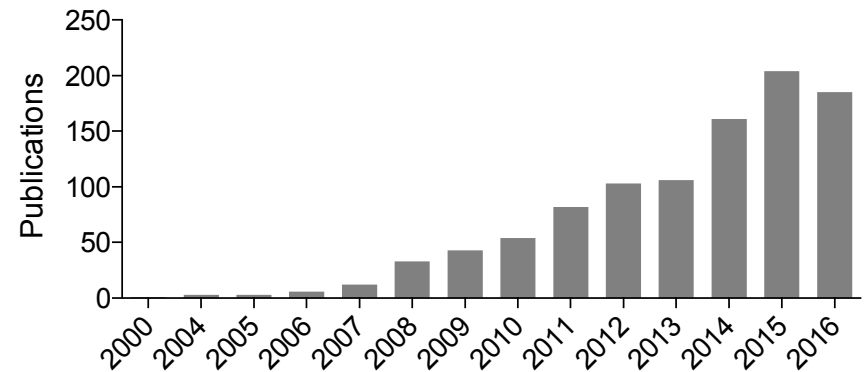


Image from: PNAS;112(41):12764-9

# User-friendly Data Analysis Solutions Needed

## Diagnostic data analysis

- Rapid (minimal impact on TAT)
- Easy to use
- Consistent performance
- Validated
- Easily updatable and expandable



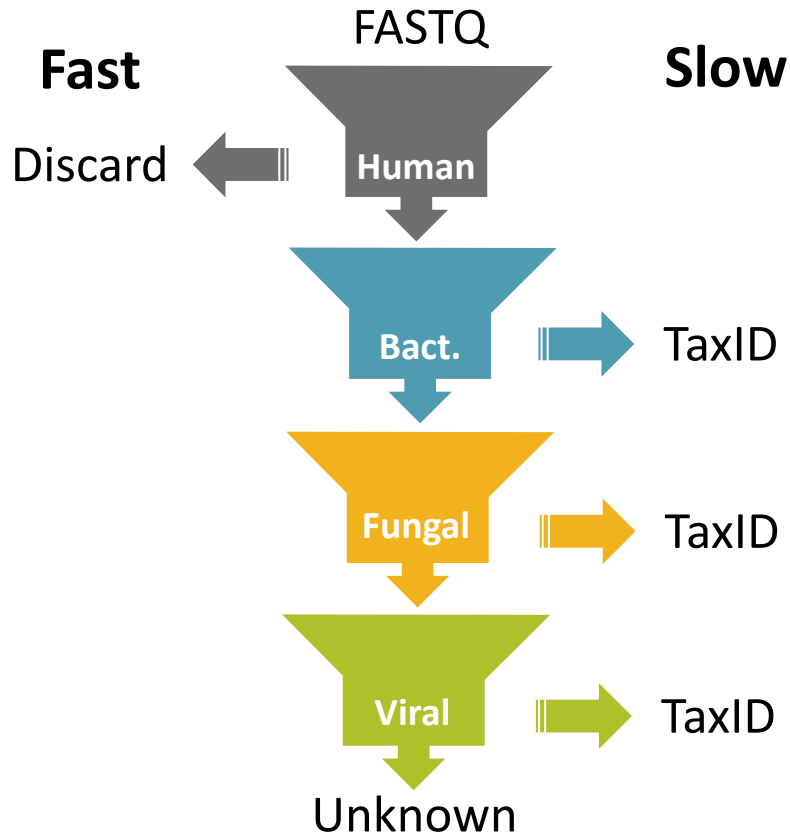
**Kraken** (Wood, Salzberg; Genome Biol 3;15(3):R46)

**SURPI** (Naccache, ..., Chiu; Genome Res;24(7):1180-92)

**CLARK** (Ounit, ..., Leonardi; BMC Genomics;16:236)

**Taxonomer** (Flygare, ..., Schlaberg; Genome Biol 26;17(1):111)

# Moving Away from Sequential Subtraction

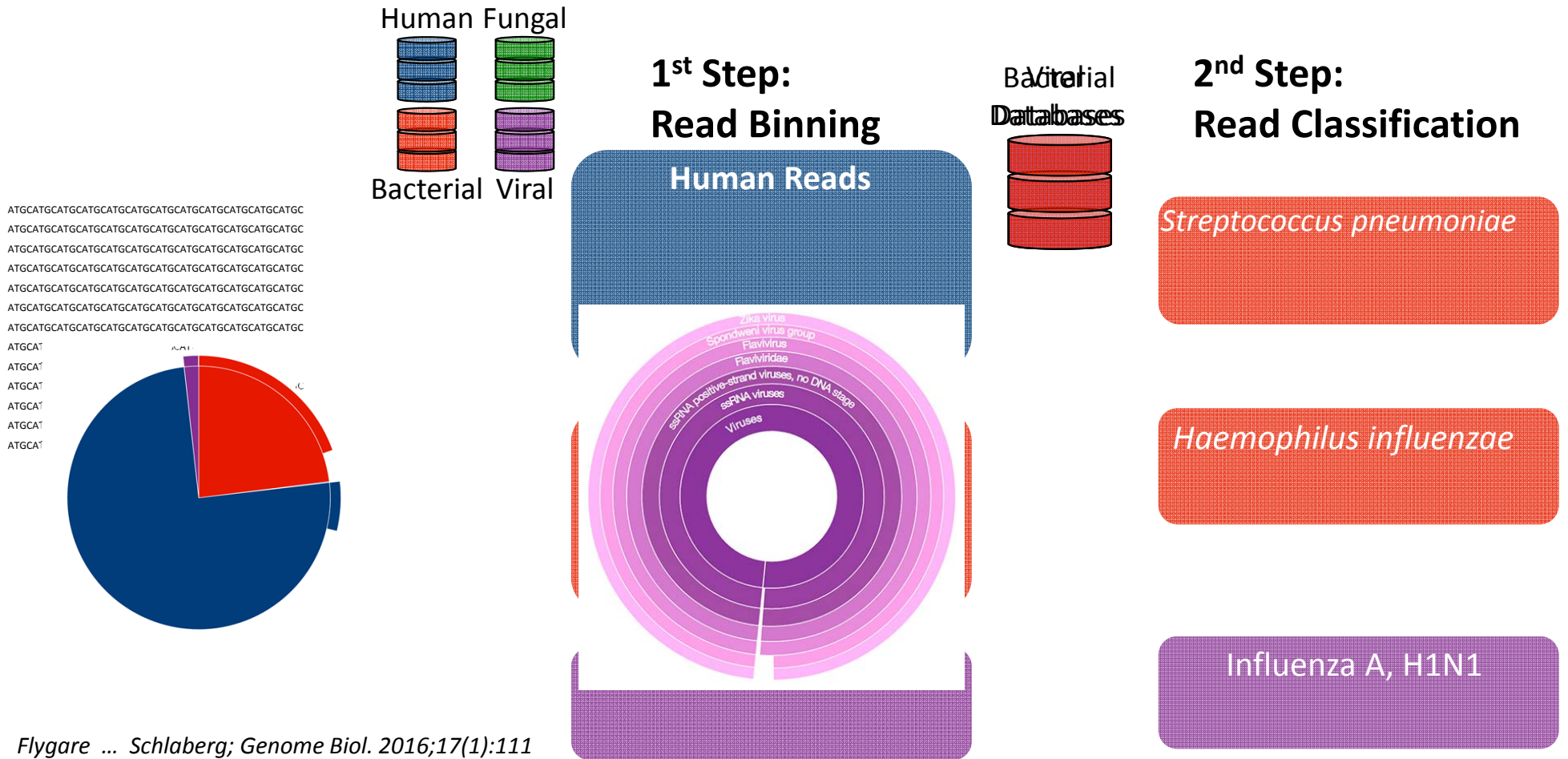


## Disadvantages

- Speed depends on sample composition
- Misclassification
  - Comparison to database subsets
  - Heterogeneous query sequences
  - Difficult to define thresholds



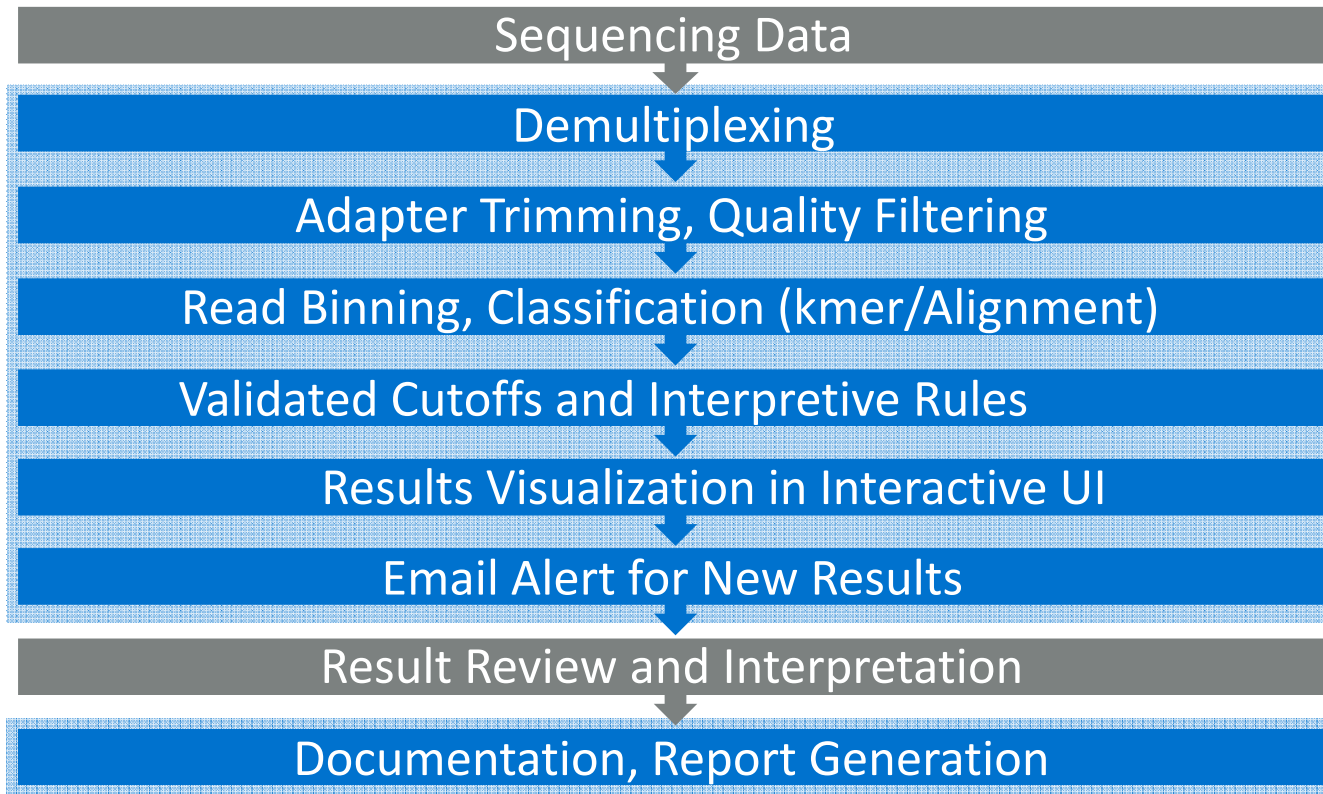
# Rapid, Interactive DNA Search Engine (Taxonomer)





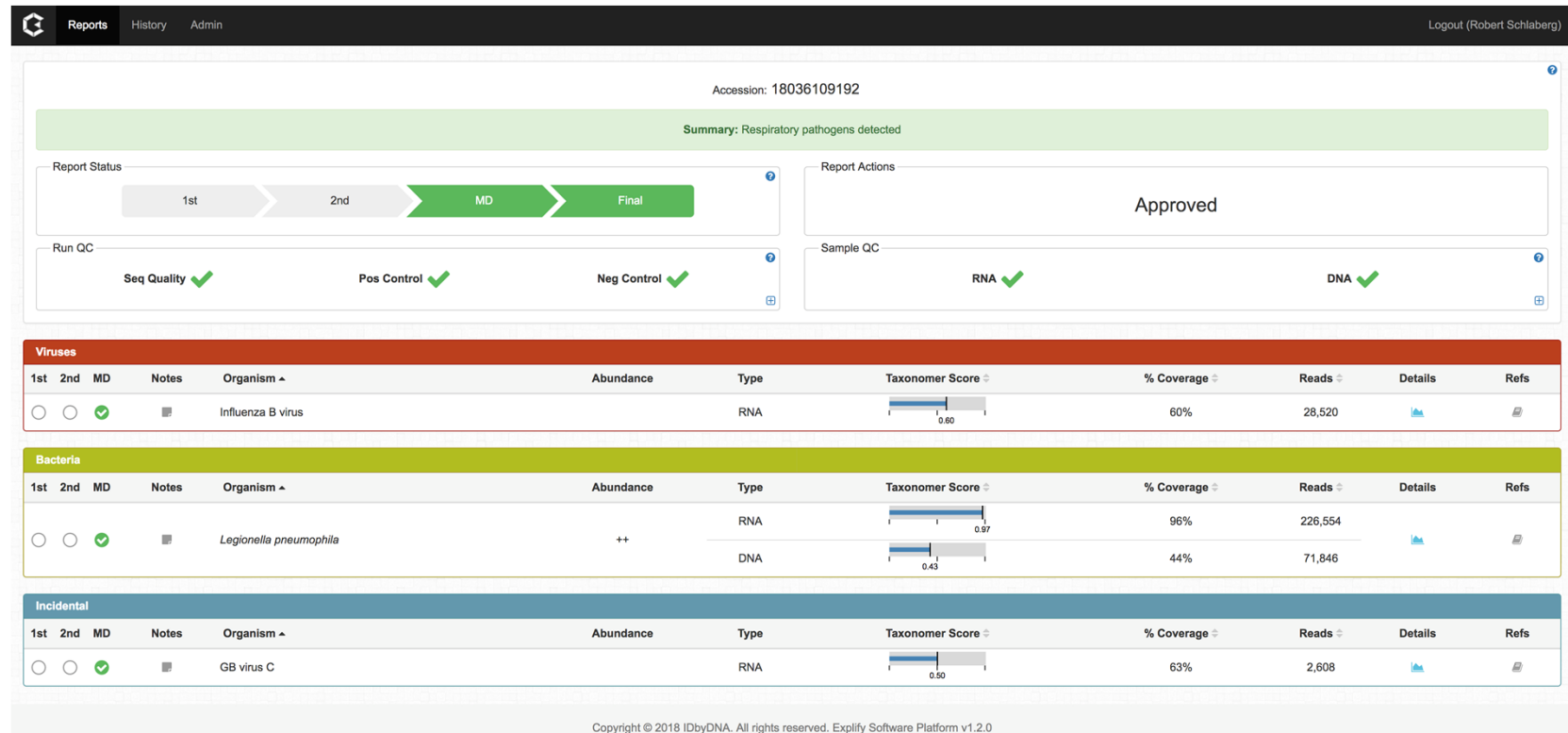
# End-to-End Metagenomics Data Analysis (Explify)

User



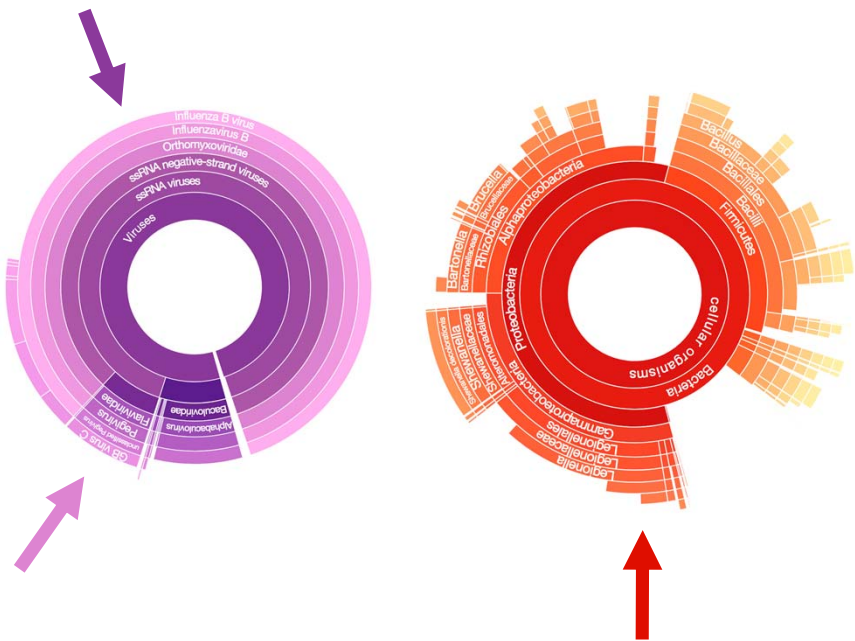
User

# End-to-End Metagenomics Data Analysis (Explify)



# Research vs. Diagnostic Software

## Research



## Diagnostic

Run QC

Seq Quality

Pos Control

Neg Control

Sample QC

RNA

DNA

Viruses

| 1st                   | 2nd                   | MD                                  | Notes | Organism ^        | Abundance | Type | Taxonomer Score ^ | % Coverage ^ | Reads ^ | Details | Refs |
|-----------------------|-----------------------|-------------------------------------|-------|-------------------|-----------|------|-------------------|--------------|---------|---------|------|
| <input type="radio"/> | <input type="radio"/> | <input checked="" type="checkbox"/> |       | Influenza B virus |           | RNA  |                   | 60%          | 28,520  |         |      |

Bacteria

| 1st                   | 2nd                   | MD                                  | Notes | Organism ^             | Abundance | Type | Taxonomer Score ^ | % Coverage ^ | Reads ^ | Details | Refs |
|-----------------------|-----------------------|-------------------------------------|-------|------------------------|-----------|------|-------------------|--------------|---------|---------|------|
| <input type="radio"/> | <input type="radio"/> | <input checked="" type="checkbox"/> |       | Legionella pneumophila | ++        | RNA  |                   | 96%          | 226,554 |         |      |
|                       |                       |                                     |       |                        |           | DNA  |                   | 44%          | 71,846  |         |      |

Incidental

| 1st                   | 2nd                   | MD                                  | Notes | Organism ^ | Abundance | Type | Taxonomer Score ^ | % Coverage ^ | Reads ^ | Details | Refs |
|-----------------------|-----------------------|-------------------------------------|-------|------------|-----------|------|-------------------|--------------|---------|---------|------|
| <input type="radio"/> | <input type="radio"/> | <input checked="" type="checkbox"/> |       | GB virus C |           | RNA  |                   | 63%          | 2,608   |         |      |

# Curated, Comprehensive, Balanced Databases

- Curated
  - Errors: misclassification
- Comprehensive
  - False-negative results if correct match missing
  - False-positive results (near neighbor) if correct match missing
- Balanced
  - Reduce bias

Periodic updates, re-validation, version control

# Curated, Comprehensive, Balanced Databases

**Microbial Universe**  
(Viruses, Bacteria, Fungi, Parasites)

**Known**

**Pathogens**

**Human Genome**



# A First Diagnostic Test

# Considerations Before Making the Leap



Known Pathogens

Standardization

Intended Use

Confidence



Discovery

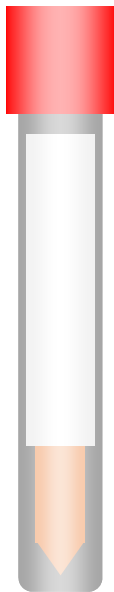
Specimen Variability

Last Resort

Promise

**Resources (Laboratory, Data Analysis)**

# First Diagnostic Applications, One Experience



Complete, Accurate, Validated  
Relevant TAT  
Cost-Effective  
Actionable



ARUP LABORATORIES

ARIA, Respiratory Pathogens by Next Generation Sequencing

ARUP Test Code 201084

Patient: DOB: Age: Gender: Client: ARUP Test Code 201084  
Patient Identifiers: Physician: Collection Date:  
Visit Number (P#): Physician: Referred to ARUP: Completion Date:

bioMérieux

bioMérieux PRECISION MEDICINE TO INFECTIOUS DISEASES

bioMérieux Inc. 1220 Boulevard Dine, Suzoville, CA 94088 www.biomérieux.com

| SUMMARY                 |                                                          |  |  |
|-------------------------|----------------------------------------------------------|--|--|
| Result:                 | Respiratory pathogens detected.                          |  |  |
| Analytical Sensitivity: | Reduced. Other respiratory pathogens cannot be excluded. |  |  |

| DETECTED PATHOGENS     |                       |                                           |                        |
|------------------------|-----------------------|-------------------------------------------|------------------------|
| <b>BACTERIA</b>        |                       |                                           |                        |
| Staphylococcus aureus  | Proportion (Bacteria) | Quantity                                  | Additional Information |
|                        | 72%                   | +++<br>10 <sup>7</sup> CFU equivalents/mL | USA300 MRSA            |
| Haemophilus influenzae | 12%                   | ++                                        | -                      |
| Streptococcus mitis    | 4%                    | +                                         | Inconclusive           |
| <b>VIROUSES</b>        |                       |                                           |                        |
| Rhinovirus A           | Proportion (Viruses)  | Quantity                                  | Additional Information |
|                        | 100%                  | n/a*                                      | -                      |
| <b>FUNGI</b>           |                       |                                           |                        |
| Pneumocystis jirovecii | Proportion (Fungi)    | Quantity                                  | Additional Information |
|                        | 77%                   | n/a*                                      | -                      |
| Mucor circinelloides   | 22%                   | n/a*                                      | Incidental             |

\* Quantity has not been established for this potential pathogen.

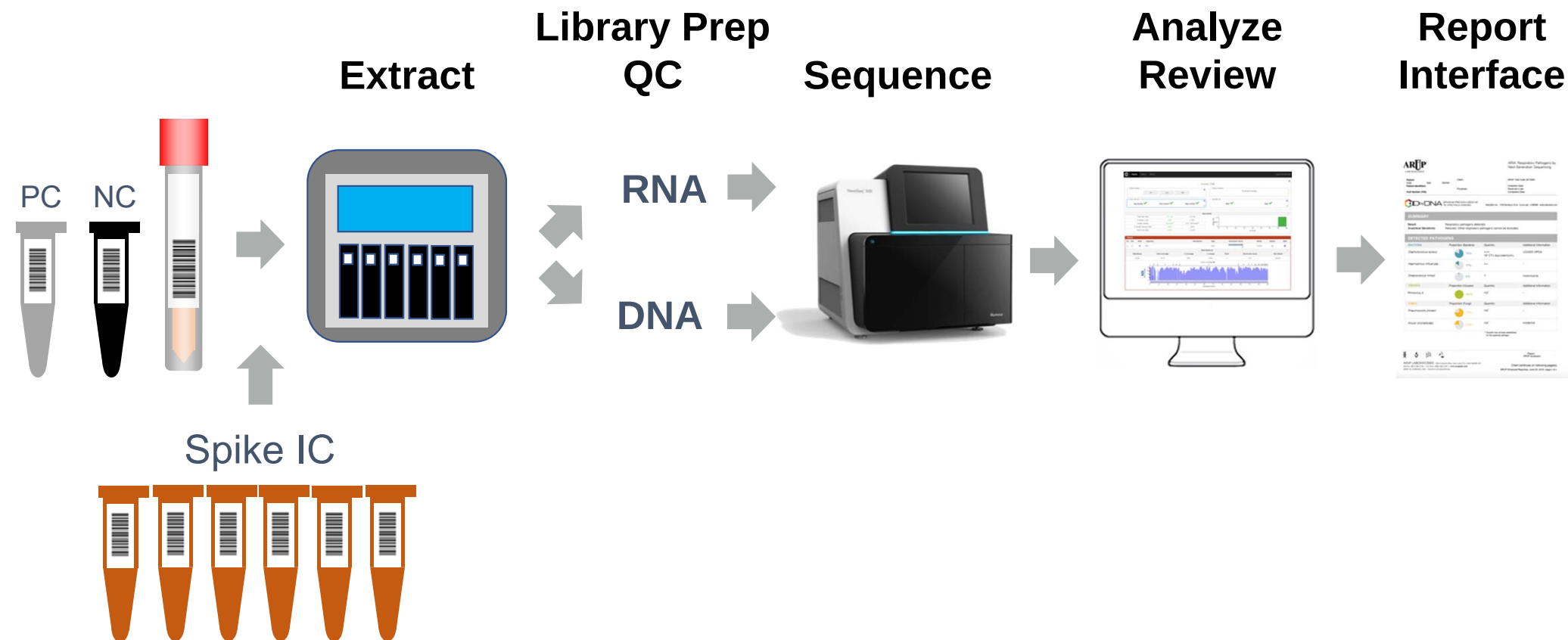
Report: ARUP accession:

ARUP LABORATORIES | 660 Chipmunk Way, Salt Lake City, Utah 84143-1077  
Phone: (801) 582-2747 | Toll Free: (800) 582-2747 | www.aruplab.com  
ARUP is an ISO15189:2013 | Certified reference laboratory

Chart continues on following page(s)  
ARUP Enhanced Reporting | June 28, 2016 | page 4 of 4



# Workflow



# Simplifying Diagnostic Workflows

## Specimen & Library Prep



## Sequencing

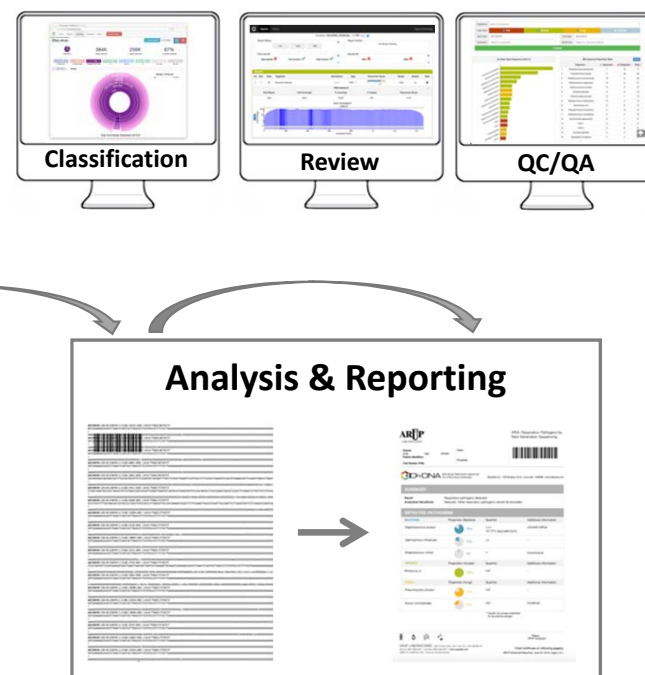
Validated

- Databases
- Algorithms
- Cutoffs



## Analysis

## Report



# Validation

## 200+ validated pathogens

- Viral, bacterial, fungal

## Patient Specimens

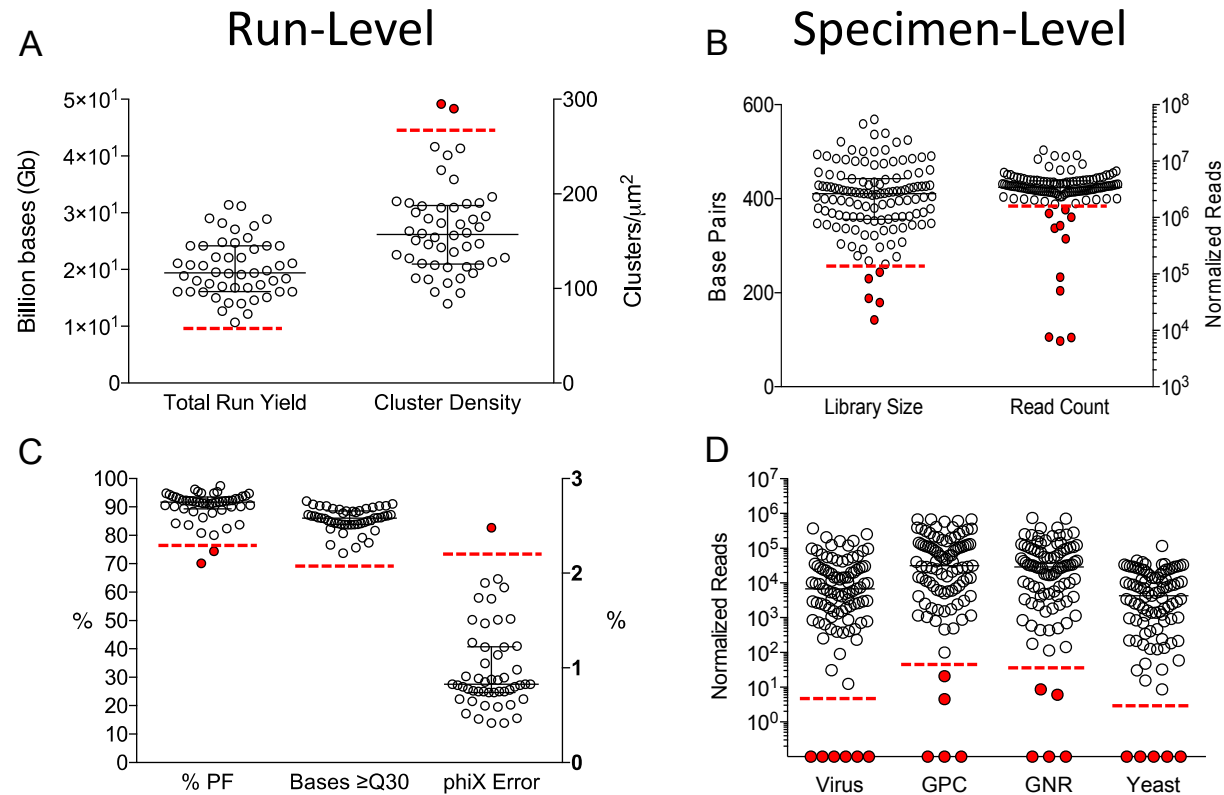
- Comparison to conventional tests
- Validation of testing system

## 1000+ Virtual Specimens

- Pathogens and near-neighbors
- Databases, algorithms, cutoffs

## QC Metrics

- Run-level
- Specimen-level



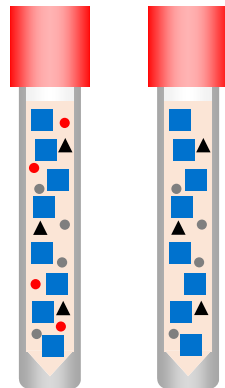
Schlaberg R, Chiu CY, Miller S, Procop G, Weinstock G, Arch Pathol Lab Med. 2017 Jun;141(6):776-786

# Validation: Generating Sequencing Reads

- ✓ Sensitivity
- ☐ Specificity
- ☐ Accuracy
- ✓ Reproducibility
- ✓ Stability

External Controls  
Internal Controls

Specimens  
Pos. Neg.



Sequencing QC  
Specimen QC  
Repeat Algorithms



Databases  
Algorithms  
Cutoffs



| Pathogen                     | Quantity   | Additional Information |
|------------------------------|------------|------------------------|
| Respiratory Syncytial Virus  | 1.2 x 10^4 | 100% RSV               |
| Human Rhinovirus             | 1.2 x 10^4 | 100% HRV               |
| Human Coronavirus            | 1.2 x 10^4 | 100% HCoV              |
| Human Parainfluenza Virus    | 1.2 x 10^4 | 100% HPIV              |
| Human Adenovirus             | 1.2 x 10^4 | 100% HAdV              |
| Human Rotavirus              | 1.2 x 10^4 | 100% HRV               |
| Human Norovirus              | 1.2 x 10^4 | 100% HNoV              |
| Human Bocavirus              | 1.2 x 10^4 | 100% HBoV              |
| Human Parvovirus B19         | 1.2 x 10^4 | 100% PPV               |
| Human Herpesvirus 6          | 1.2 x 10^4 | 100% HHV-6             |
| Human Herpesvirus 8          | 1.2 x 10^4 | 100% HHV-8             |
| Human Cytomegalovirus        | 1.2 x 10^4 | 100% CMV               |
| Human Epstein-Barr Virus     | 1.2 x 10^4 | 100% EBV               |
| Human Hepatitis A Virus      | 1.2 x 10^4 | 100% HAV               |
| Human Hepatitis B Virus      | 1.2 x 10^4 | 100% HBV               |
| Human Hepatitis C Virus      | 1.2 x 10^4 | 100% HCV               |
| Human Immunodeficiency Virus | 1.2 x 10^4 | 100% HIV               |
| Human Papillomavirus         | 1.2 x 10^4 | 100% HPV               |
| Human Papillomavirus 16      | 1.2 x 10^4 | 100% HPV16             |
| Human Papillomavirus 18      | 1.2 x 10^4 | 100% HPV18             |
| Human Papillomavirus 31      | 1.2 x 10^4 | 100% HPV31             |
| Human Papillomavirus 33      | 1.2 x 10^4 | 100% HPV33             |
| Human Papillomavirus 35      | 1.2 x 10^4 | 100% HPV35             |
| Human Papillomavirus 39      | 1.2 x 10^4 | 100% HPV39             |
| Human Papillomavirus 45      | 1.2 x 10^4 | 100% HPV45             |
| Human Papillomavirus 52      | 1.2 x 10^4 | 100% HPV52             |
| Human Papillomavirus 58      | 1.2 x 10^4 | 100% HPV58             |
| Human Papillomavirus 66      | 1.2 x 10^4 | 100% HPV66             |
| Human Papillomavirus 68      | 1.2 x 10^4 | 100% HPV68             |
| Human Papillomavirus 73      | 1.2 x 10^4 | 100% HPV73             |
| Human Papillomavirus 82      | 1.2 x 10^4 | 100% HPV82             |
| Human Papillomavirus 84      | 1.2 x 10^4 | 100% HPV84             |
| Human Papillomavirus 89      | 1.2 x 10^4 | 100% HPV89             |
| Human Papillomavirus 91      | 1.2 x 10^4 | 100% HPV91             |
| Human Papillomavirus 93      | 1.2 x 10^4 | 100% HPV93             |
| Human Papillomavirus 94      | 1.2 x 10^4 | 100% HPV94             |
| Human Papillomavirus 96      | 1.2 x 10^4 | 100% HPV96             |
| Human Papillomavirus 97      | 1.2 x 10^4 | 100% HPV97             |
| Human Papillomavirus 98      | 1.2 x 10^4 | 100% HPV98             |
| Human Papillomavirus 99      | 1.2 x 10^4 | 100% HPV99             |
| Human Papillomavirus 101     | 1.2 x 10^4 | 100% HPV101            |
| Human Papillomavirus 102     | 1.2 x 10^4 | 100% HPV102            |
| Human Papillomavirus 103     | 1.2 x 10^4 | 100% HPV103            |
| Human Papillomavirus 104     | 1.2 x 10^4 | 100% HPV104            |
| Human Papillomavirus 105     | 1.2 x 10^4 | 100% HPV105            |
| Human Papillomavirus 106     | 1.2 x 10^4 | 100% HPV106            |
| Human Papillomavirus 107     | 1.2 x 10^4 | 100% HPV107            |
| Human Papillomavirus 108     | 1.2 x 10^4 | 100% HPV108            |
| Human Papillomavirus 109     | 1.2 x 10^4 | 100% HPV109            |
| Human Papillomavirus 110     | 1.2 x 10^4 | 100% HPV110            |
| Human Papillomavirus 111     | 1.2 x 10^4 | 100% HPV111            |
| Human Papillomavirus 112     | 1.2 x 10^4 | 100% HPV112            |
| Human Papillomavirus 113     | 1.2 x 10^4 | 100% HPV113            |
| Human Papillomavirus 114     | 1.2 x 10^4 | 100% HPV114            |
| Human Papillomavirus 115     | 1.2 x 10^4 | 100% HPV115            |
| Human Papillomavirus 116     | 1.2 x 10^4 | 100% HPV116            |
| Human Papillomavirus 117     | 1.2 x 10^4 | 100% HPV117            |
| Human Papillomavirus 118     | 1.2 x 10^4 | 100% HPV118            |
| Human Papillomavirus 119     | 1.2 x 10^4 | 100% HPV119            |
| Human Papillomavirus 120     | 1.2 x 10^4 | 100% HPV120            |

- ✓ Sensitivity
- ✓ Specificity
- ✓ Accuracy
- ✓ Reproducibility
- ✓ Stability

- ✓ Sensitivity
- ✓ Specificity
- ✓ Accuracy
- ✓ Reproducibility
- ✓ Stability



x10s

# Virtual Matrix

Virtual  
Specimen  
x1000's

[illegible][illegible]

ATGCATGTCATGCATGCATGC  
ATGCATGTCATGCATGCATGC  
ATGCATGTCATGCATGCATGC  
ATGCATGTCATGCATGCATGC  
ATGCATGTCATGCATGCATGC  
**ATGCATGTCATGCATGCATGC**  
ATGCATGTCATGCATGCATGC  
ATGCATGTCATGCATGCATGC  
ATGCATGTCATGCATGCATGC  
ATGCATGTCATGCATGCATGC  
ATGCATGTCATGCATGCATGC  
ATGCATGTCATGCATGCATGC  
ATGCATGTCATGCATGCATGC  
**ATGCATGTCATGCATGCATGC**  
ATGCATGTCATGCATGCATGC  
ATGCATGTCATGCATGCATGC



# Diagnostic Yield

## Immunocompromised Children with PNA (ICU)

**Table 1.** Underlying Conditions

| Condition                   | n  | %  |
|-----------------------------|----|----|
| Bone marrow transplantation | 17 | 42 |
| Solid organ transplantation | 11 | 27 |
| Primary immunodeficiency    | 7  | 17 |
| Chemotherapy                | 6  | 15 |
| High dose steroid therapy   | 2  | 5  |

**Table 2.** Radiographic Findings

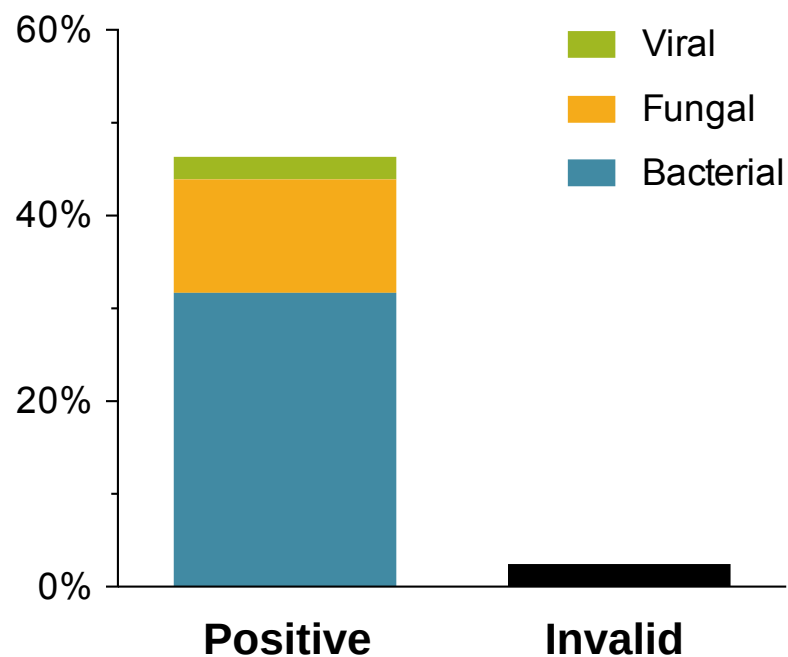
| Condition                              | n      | %       |
|----------------------------------------|--------|---------|
| Diffuse opacities [with consolidation] | 28 [7] | 68 [17] |
| Nodules                                | 6      | 15      |
| Pleural effusions                      | 6      | 15      |

# Diagnostic Yield

## Immunocompromised Children with PNA (ICU)

**A**

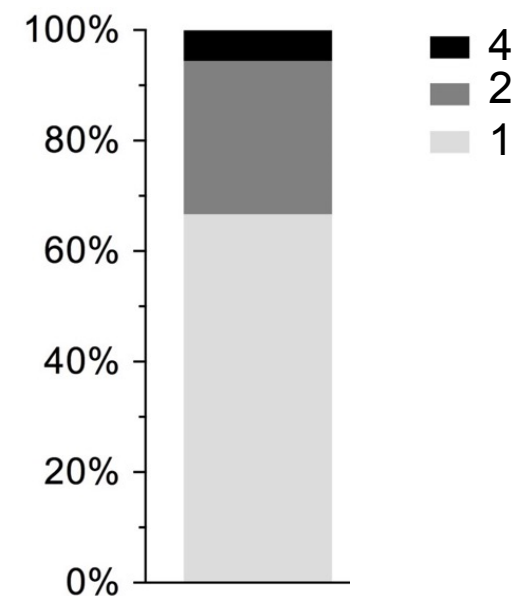
**Explify™ Respiratory Results**



|                 |                                                                           |
|-----------------|---------------------------------------------------------------------------|
| <b>Viruses</b>  | Parainfluenza virus<br>Human Bocavirus<br>Rhinovirus                      |
| <b>Fungi</b>    | <i>Pneumocystis</i><br><i>Mucor</i><br><i>Fusarium</i>                    |
| <b>Bacteria</b> | <i>Staph. aureus</i><br><i>Strep. pneumoniae</i><br><i>S. maltophilia</i> |

**B**

**Number of Pathogens/Case**



Graf ... Schlager, manuscript in preparation

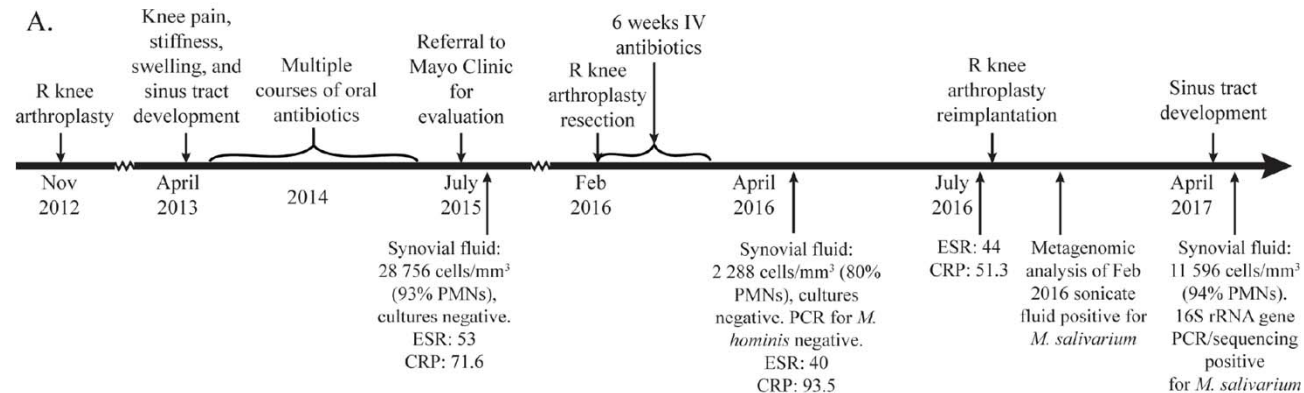
# Other Applications



# Unexpected Pathogens in PJI

52-year-old man with chronic right knee prosthetic joint infection

- Swelling, drainage from multiple sinus tracts 6 mo after the TKA
- *Mycoplasma salivarium* by unbiased DNA sequencing

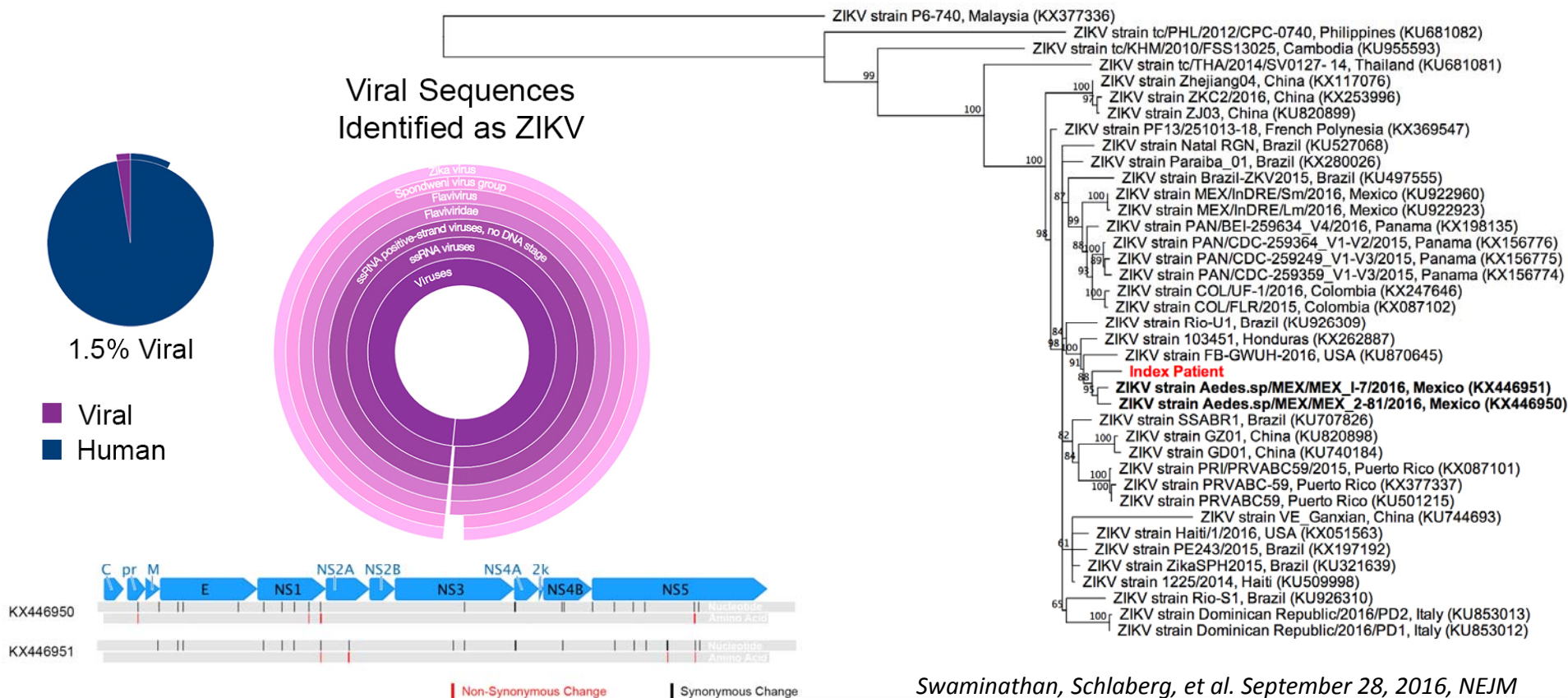


Thoendel, ... , Patel R; Clin Infect Dis. 2017 Jul 15;65(2):332-335



# Identification of ZIKV (Only) in Fatal Case

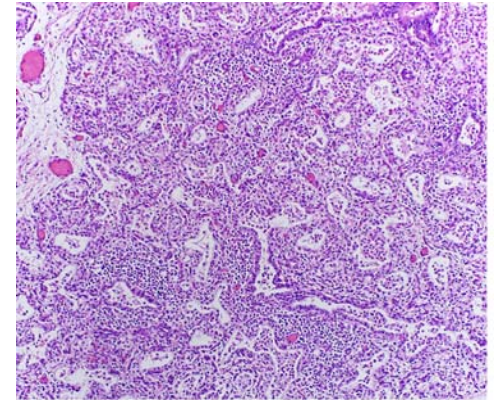
Serum, RNA-seq & Taxonomer



Swaminathan, Schlager, et al. September 28, 2016, NEJM

# Gestational Psittacosis, Neonatal Death

- 32-year-old pregnant female (16 weeks gestation)
- Persistent fever, neck pain, headache, nausea
- Normal WBC, left shift, thrombocytopenia
- Extensive workup without etiologic diagnosis
- Pre-term labor at 21 weeks gestation
  - Acute chorioamnionitis
  - Congenital pneumonia
- *Chlamydia psittaci* by RNA-seq



Paul, ..., Schlager, Submitted

# Other Infectious Syndromes

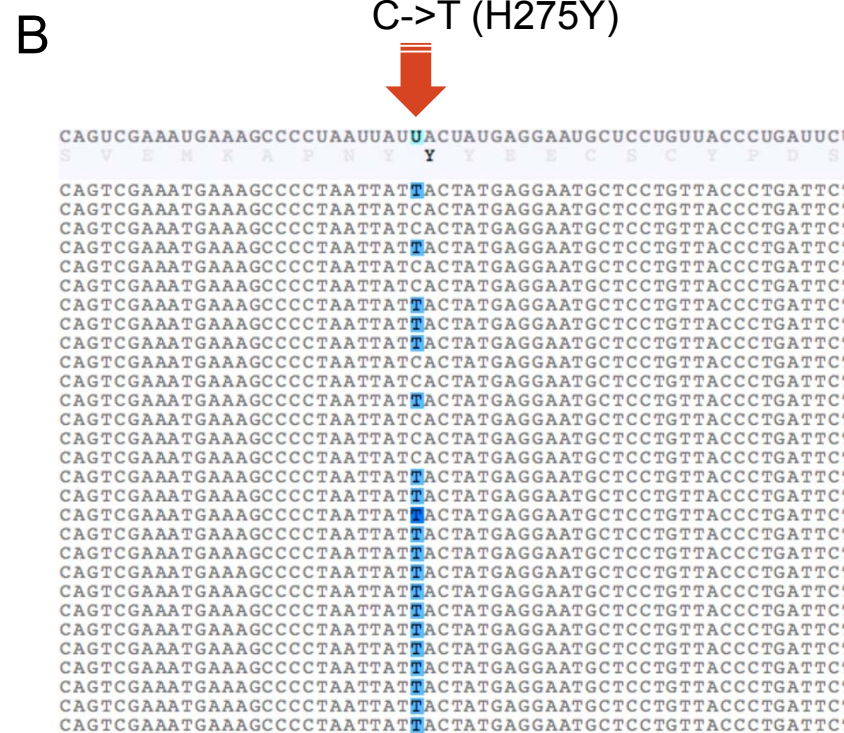
| Organ System     | Diagnosis                | Pathogen (Examples)                        |
|------------------|--------------------------|--------------------------------------------|
| CNS              | Chronic meningitis       | <i>Leptospira borgpetersenii</i>           |
|                  | Progressive encephalitis | Variegated squirrel bornavirus 1           |
|                  | Meningoencephalitis      | <i>Balamuthia mandrillaris</i>             |
| Ocular           | Chronic uveitis          | Rubella virus                              |
|                  | Keratitis                | <i>Fusarium sp.</i>                        |
| Bloodstream      | Fever of unknown origin  | Parvovirus B19, <i>P. falciparum</i> , ... |
|                  | Sepsis                   | <i>Enterobacter cloacae</i> , ...          |
| Gastrointestinal | Diarrhea                 | <i>Clostridium difficile</i> , ...         |

Simner et al., CID;66(5):778-788

Shigeyasu et al., J Infect Chemother. 2018 Jan 20. pii: S1341-321X(17)30320-3

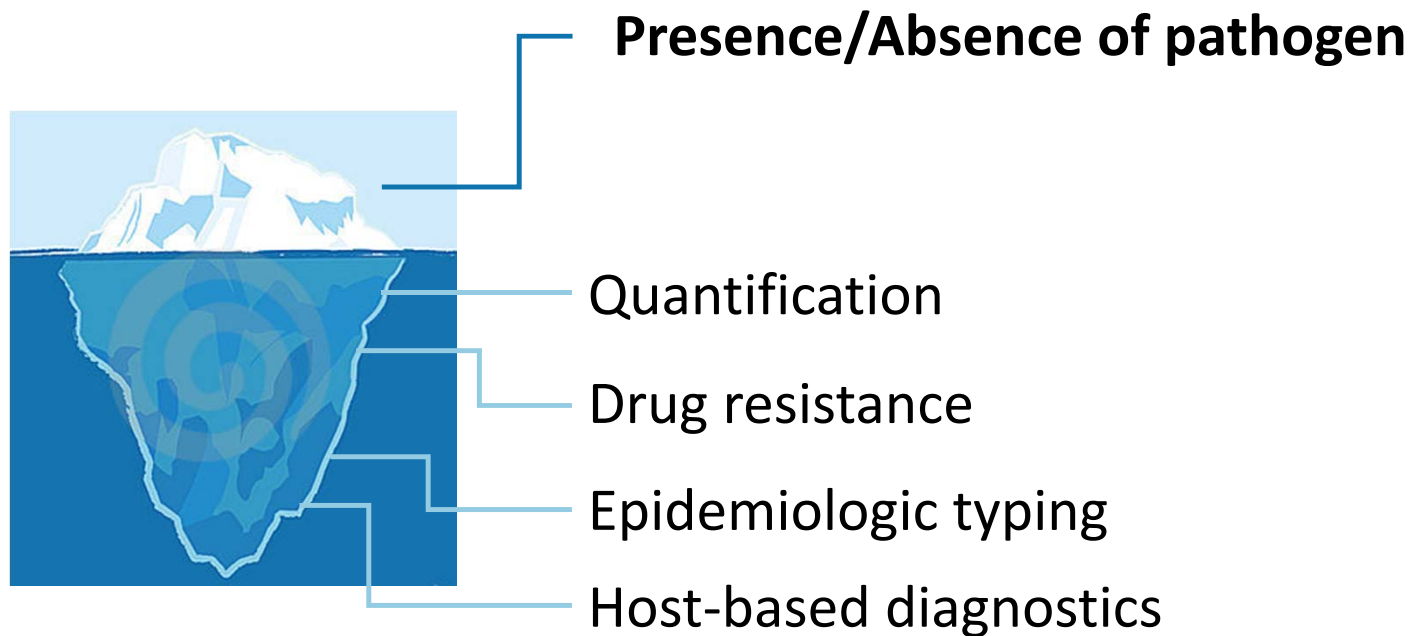
Grumaz et al., Genome Med. 2016 Jul 1;8(1):73

# Monitoring Oseltamivir Resistance, 3-Year-Old Immunocompromised Child



NATIONAL REFERENCE LABORATORY

# Much More Information than Pathogen Detection





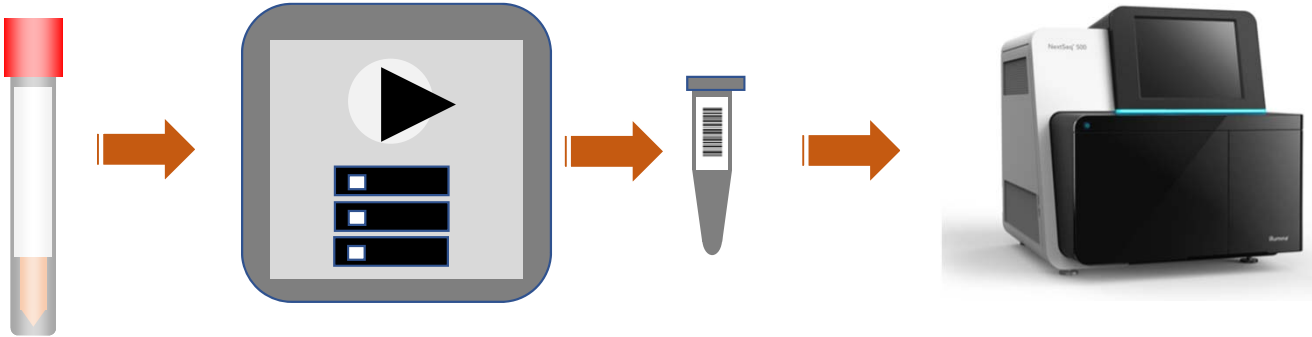
# Identification , Quantification, AMR

## Automated Sample & Library Prep

## Rapid Sequencing Flexible Batch Size

## Rapid, Automated Data Analysis

Various



### Reagents & Consumables

- DNA/RNA-free
- Closed system

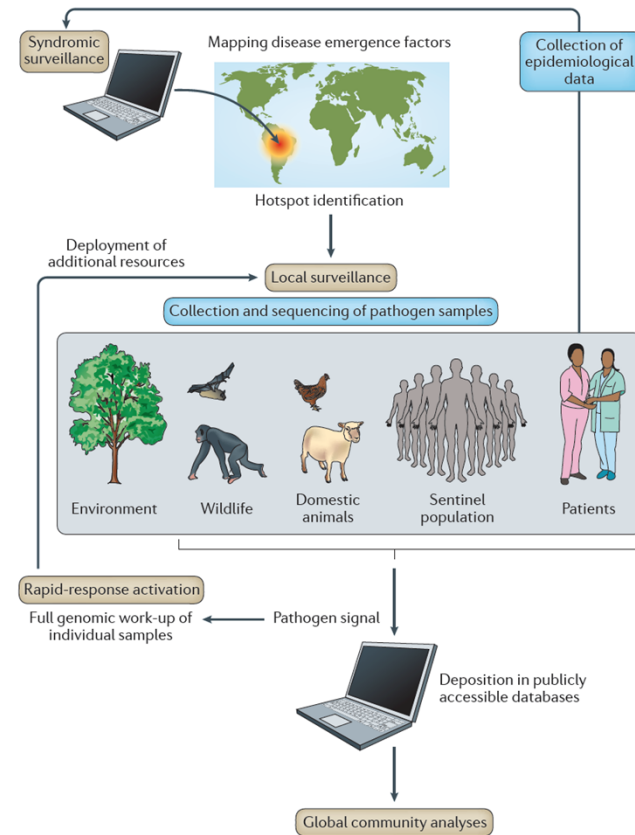
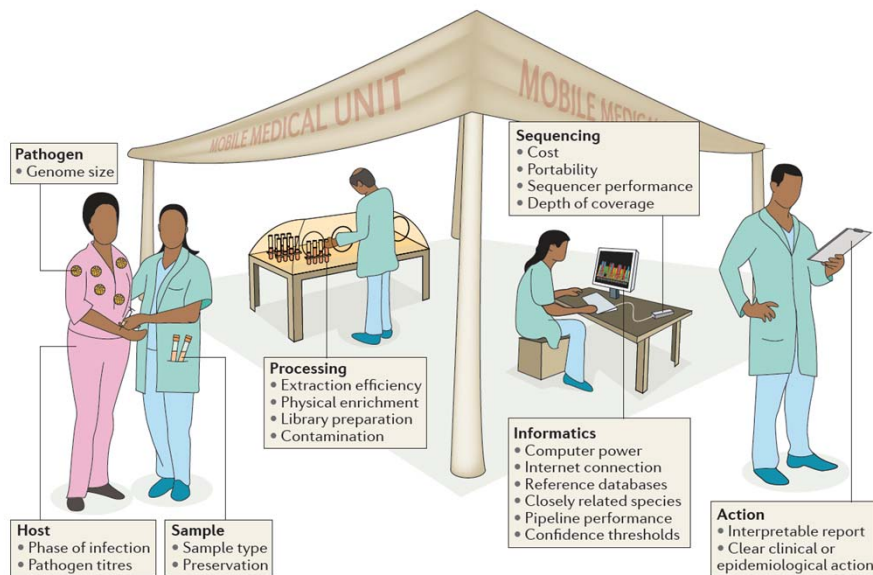
### Reagents

- Consistent performance
- Stable reagents
- Few lot changes

### Validated

- Databases
- Algorithms
- Cutoffs

# Sequencing-based, real-time, global pathogen surveillance



Gardy & Lorman; Nat Rev Genet. 2018 Jan;19(1):9-20



# Metagenomics In Routine Diagnostic Practice

## Advantages

- Does not rely on differential diagnoses, 'unbiased' to get rid of 'hypothesis-free'
- Resistant to genetic variation, suitable for discovery
- Detection of multiple microorganisms
- Does not rely on viability
- Workflow

## Challenges

- Technologies need to become faster, easier to use, cheaper
- Specimen composition affecting performance
- Complex to validate and maintain
- Skills & resources often not available in microbiology laboratories

*Allcock et al. J Clin Microbiol. 2017 Nov;55(11):3175-3182*

### **Human Genetics**

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Mark Yandell

### **ARUP Laboratories**

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# Questions?

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