

# Principles of Pharmacogenomics

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# Learning objectives

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**Define what pharmacogenetics/genomics (PGx) is**

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**Discuss pharmacogenes and possible genetic variations**

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**Learn what molecular methodologies are used in the field**

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**Discuss PGx Implementation challenges**

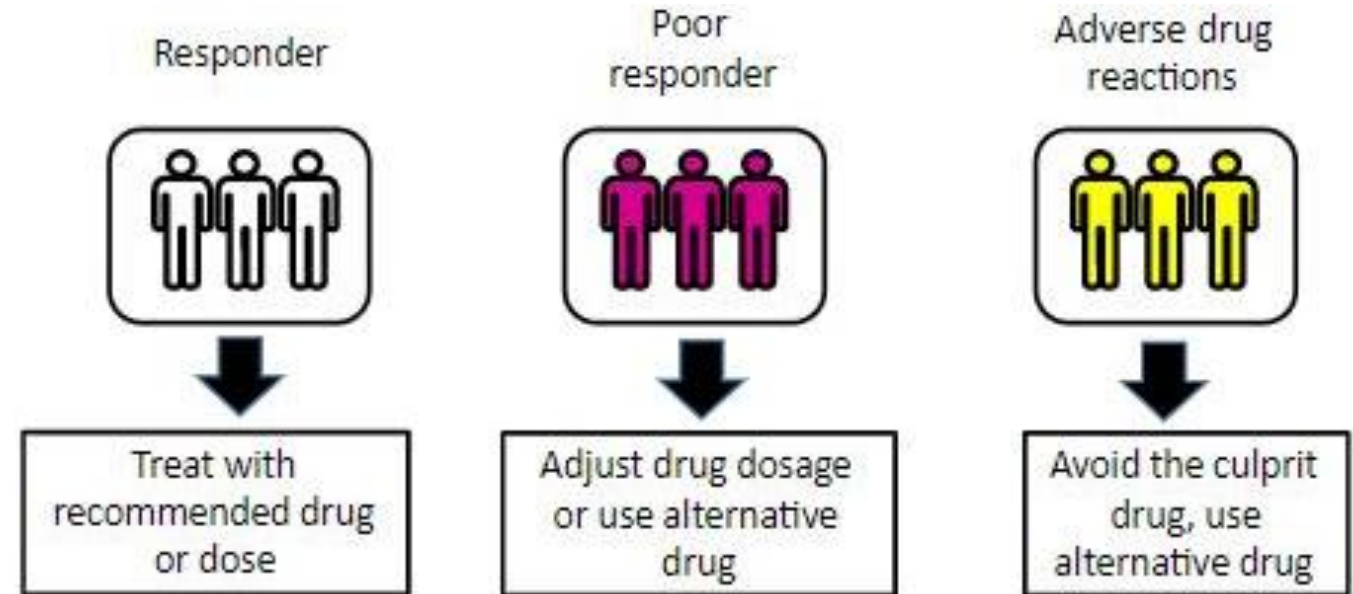
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# What is Pharmacogenetics?

## Layman's language

- Pharmacogenetics (sometimes called pharmacogenomics) is a field of research that studies how a person's genes affect how he or she responds to medications. Its long-term goal is to help doctors select the drugs and doses best suited for each person.
- It is part of the field of precision medicine, which aims to treat each patient individually.

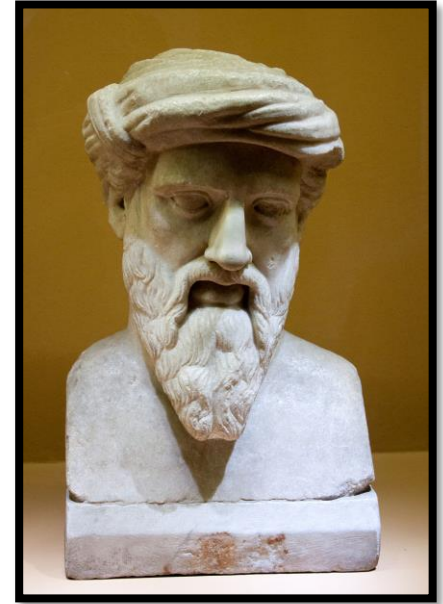


# History

## Pythagoras (570-495 BC):

The first pharmacogenetics observation was made by  
("be far from the fava beans consumption")

He noticed that fava beans caused a condition (*now known as acute hemolytic anemia*) in certain people but not in others



Archibald Garrod (1857-1936) discovered inherited alkaptonuria.

Established a concept: the metabolism of molecular compounds can be altered by inherited genetic factors and cause an abnormal accumulation of "intermediate" metabolites



Meletis J. Favism: a brief history from the "abstain from beans" of Pythagoras to the present. Arch Hellenic Med 2012; 29: 253-263





1957

## Arno Motulsky (1923–2018)

- Earned his title of the father of pharmacogenetics

“It is not unlikely that some drug sensitivity reactions ... be produced by (genetic) mechanisms”

Two major observations:

- Some soldiers developed hemolytic anemia when given Primaquine (antimalarial)
- Some patients developed prolonged apnea when given succinylcholine

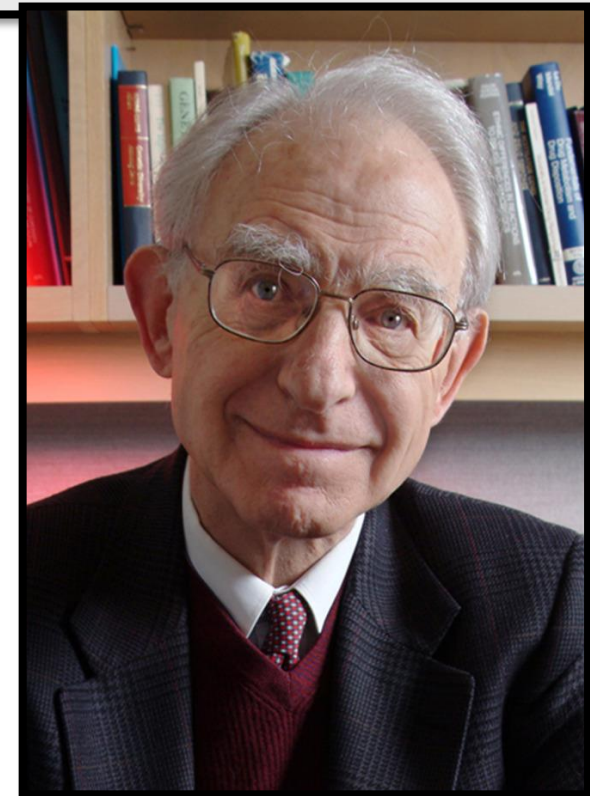
### DRUG REACTIONS, ENZYMES, AND BIOCHEMICAL GENETICS

Arno G. Motulsky, M.D., Seattle

In discussions of drug idiosyncrasy, careful distinction should be made between toxic reactions caused by immunologic mechanisms (drug allergy) and abnormal reaction caused by exaggeration or diminution of the usual effect of a given dose.<sup>1</sup> Although some progress has been made in the study of mechanisms of drug allergy, little was known until recently about the pathogenesis of hypersusceptibility reactions and hyposusceptibility reactions. Data are available now which suggest that reactions of this type may be caused by otherwise innocuous genetic traits or enzyme deficiencies.

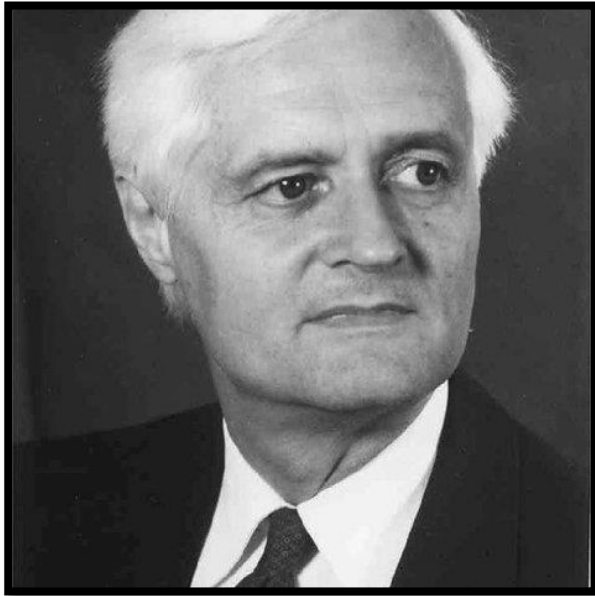
From the Department of Medicine, University of Washington Medical School. Dr. Motulsky is a John and Mary R. Markle Scholar in Medical Science.

Hockwald and his co-workers<sup>2</sup> demonstrated that approximately 10% of American Negroes and a very small number of caucasians developed hemolytic anemia when given an average dose of primaquine or chemically related drugs. Bentler and associates<sup>3</sup> showed that red blood cells of susceptible individuals possessed decreased numbers of nonprotein, sulfhydryl groups. It has now been pointed out that primaquine sensitivity is related to glucose-6-phosphate dehydrogenase activity.<sup>4</sup> Investigations of the genetics of this trait, now in progress, suggest that the abnormality is caused by a sex-linked gene of intermediate dominance.<sup>5</sup> The red blood cell abnormality per se has no known deleterious effect on the individual or on red blood cell life span. Excessive doses



# Friedrich Vogel (1925–2006)

He developed the term “pharmacogenetics” in 1959, as the close contact between genetics and pharmacology



## II. Moderne Probleme der Humangenetik

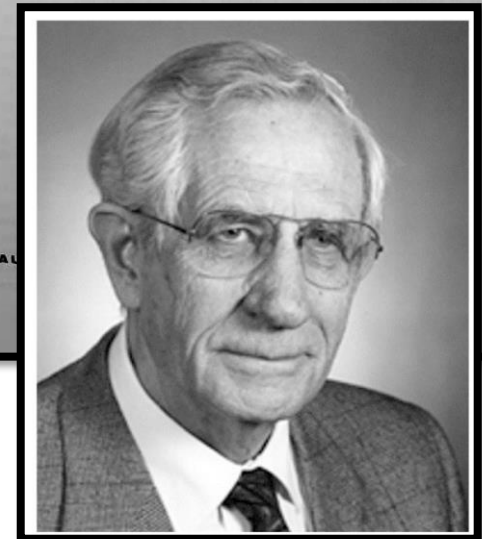
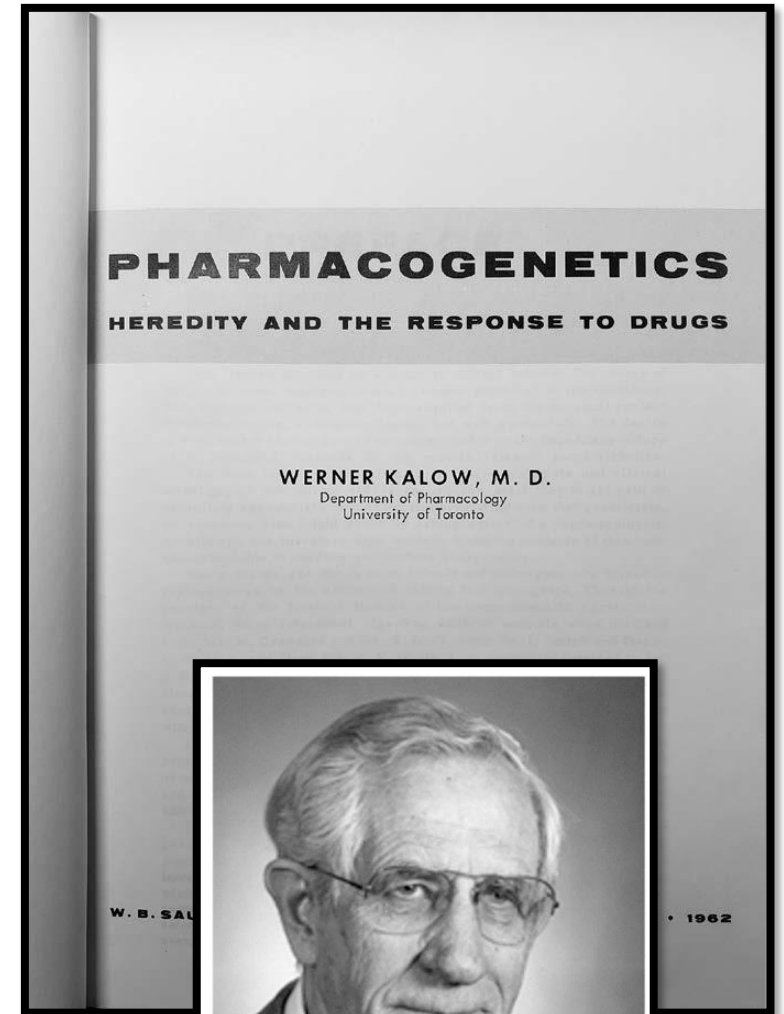
Von  
FRIEDRICH VOGEL\*

### 6. Ansätze zu einer Pharmakogenetik des Menschen

Der Ausdruck „Pharmakogenetik“ des Menschen, den wir zur Überschrift dieses Kapitels wählten, ist bisher noch *mehr ein Programm als eine Bezeichnung für ein Arbeitsgebiet*. Bisher sind zu einer Kenntnis der erblichen Varianten in der Reaktion auf Arzneimittel und sonstige von außen zugeführte Stoffe nur Ansätze vorhanden. Zum größten Teil liegt das sicher daran, daß diese Beziehungen einfach noch kaum untersucht sind; einzelne Beobachtungen scheinen darauf hinzudeuten, daß das ganze Gebiet in Zukunft immer wichtiger werden dürfte.

# Werner Kalow (1917–2008)

- The first book entirely dedicated to the field.
- Summarizes all the work and available knowledge of that time.
- Helped change pharmacogenetics from a subspecialty to an entire field

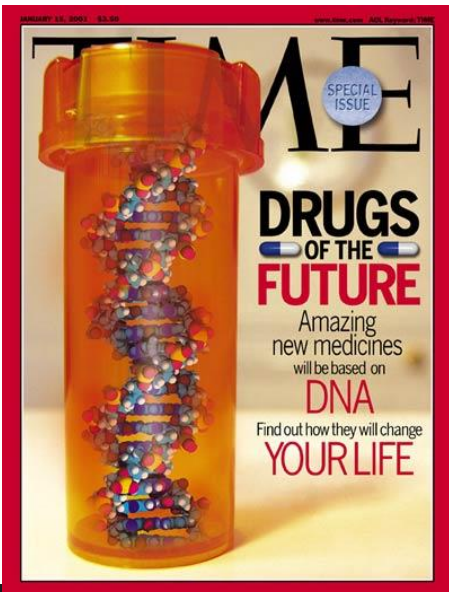


# What is

## Pharmacogenetics?

### Non-Layman's language

- Pharmacogenetics is the branch of pharmacology and genetics concerned with the inter-individual metabolic and therapeutic responses to a given medication.



Encompasses two closely related fields:

**Pharmacokinetics:** Studies the absorption, distribution, metabolism, and excretion pathways of the drug.

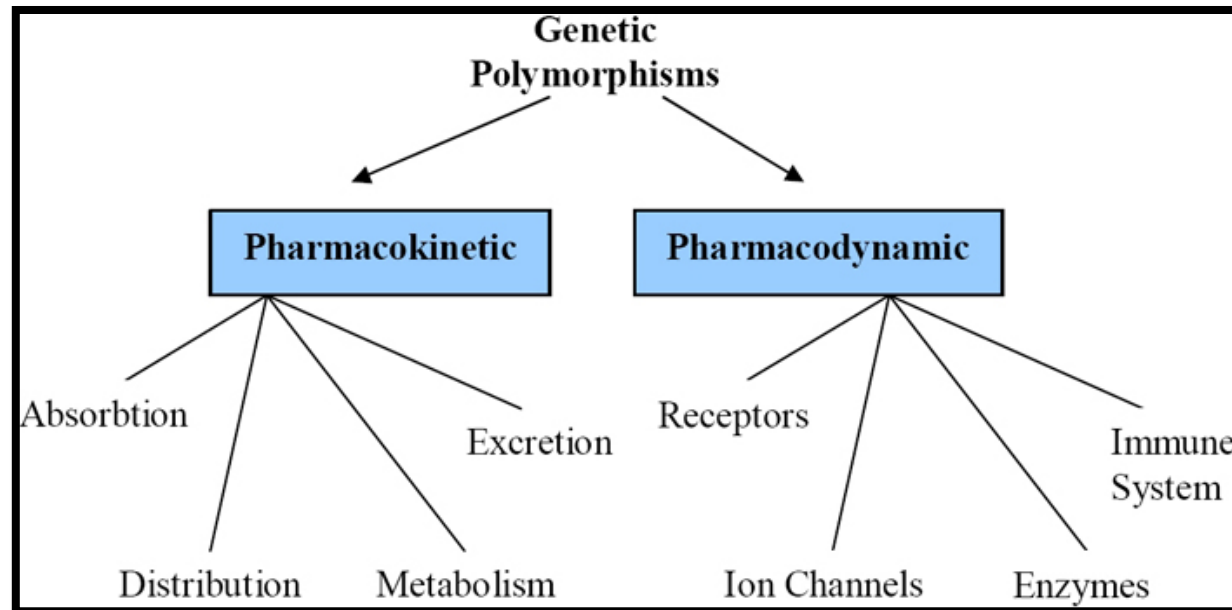
'What the body does to the drug'

BODY → DRUG

**Pharmacodynamics:** Concerned with the drug effects on the organism as a whole

'What the drug does to the body'

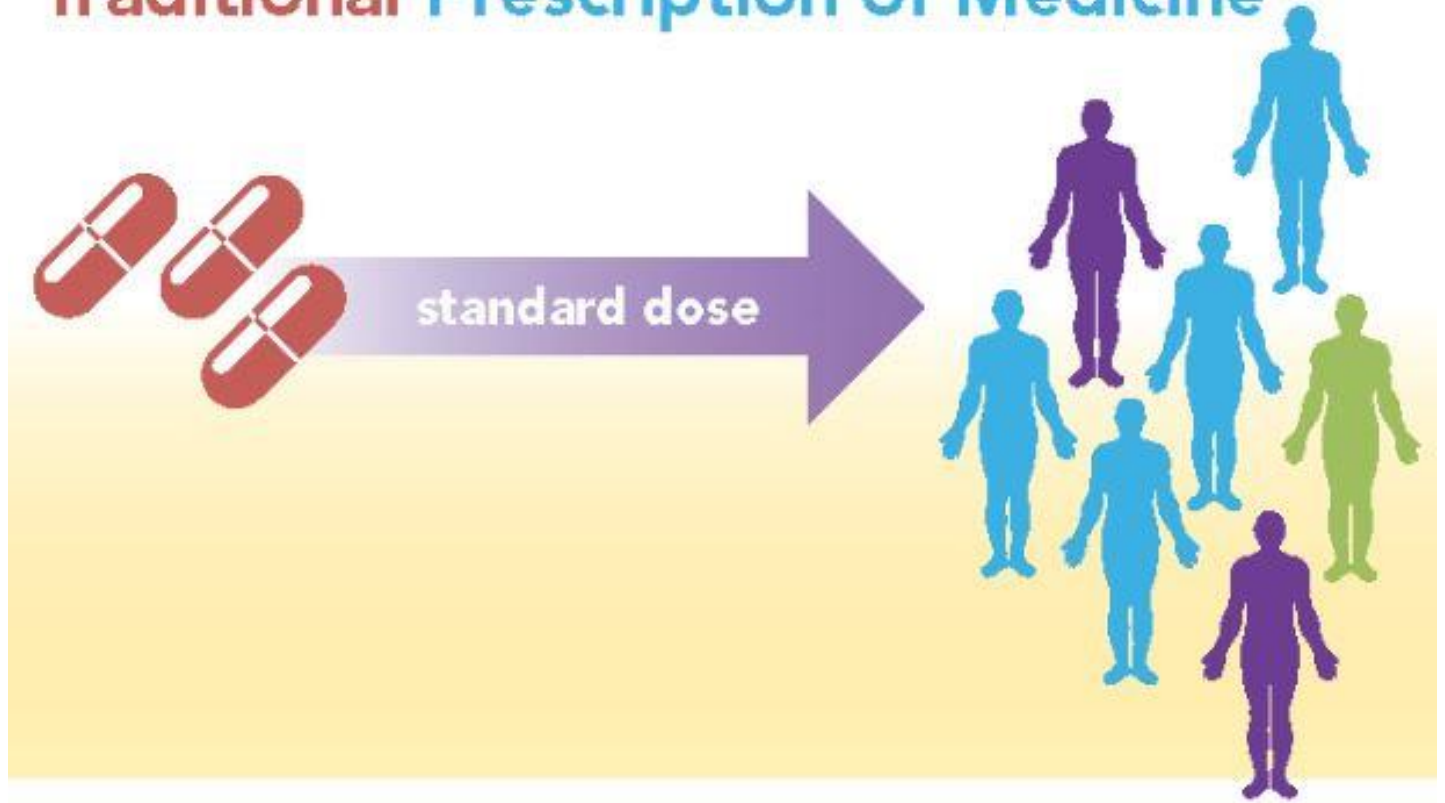
DRUG → BODY



Current  
practice

One size fits all

## Traditional Prescription of Medicine

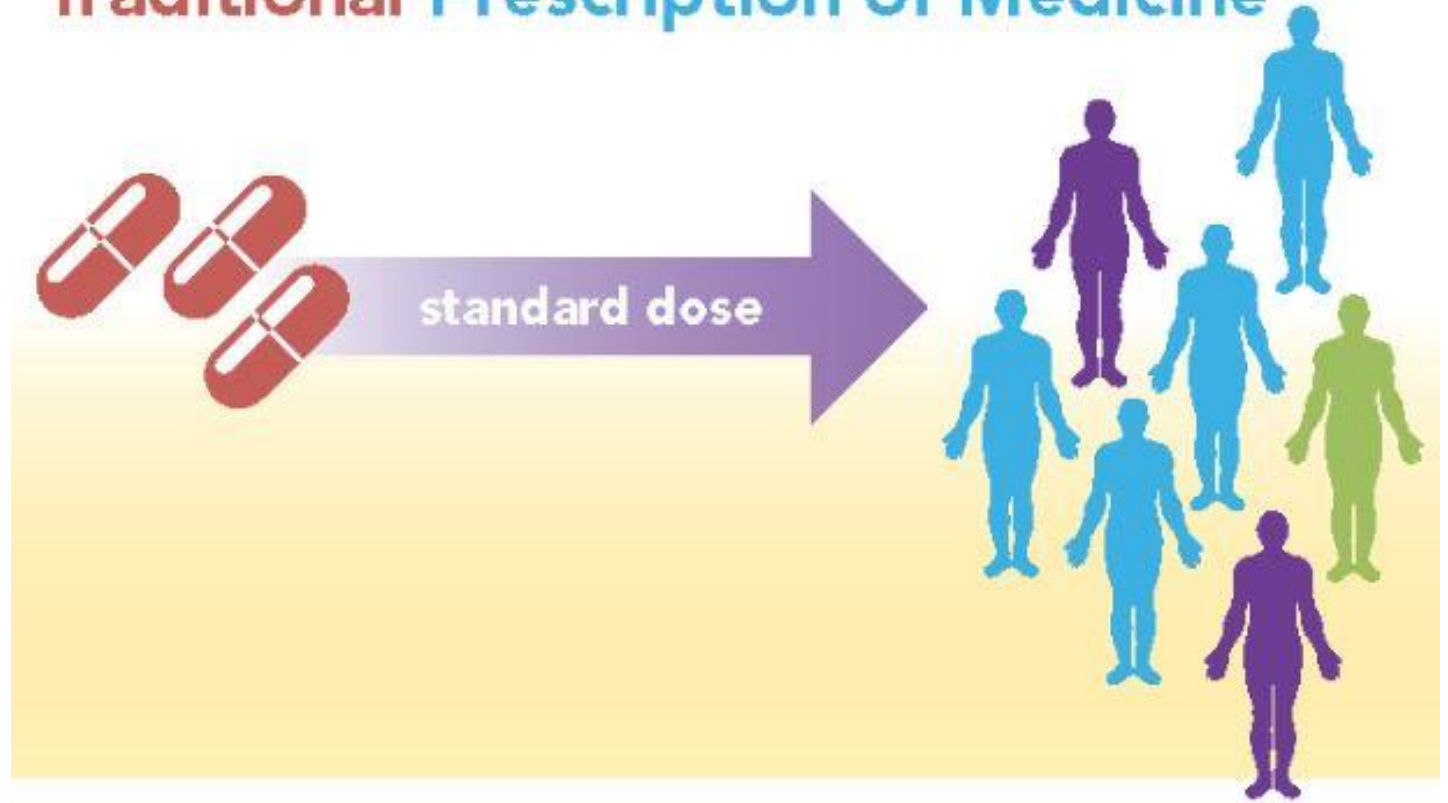




Current practice

One size fits all

## Traditional Prescription of Medicine



Dose works for many



Need less dose



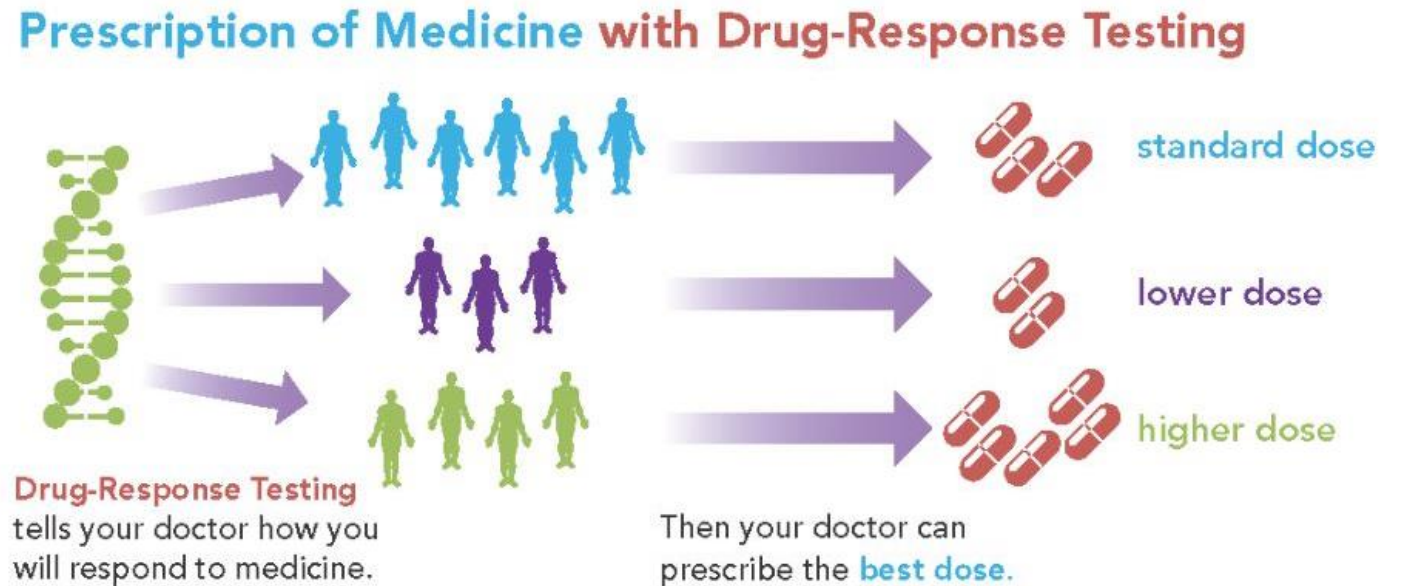
Need higher dose, or  
alternative drug



# Goals of Pharmacogenetics

- Maximize drug efficacy
- Minimize drug toxicity
- Predict patients who will respond to intervention
- Aid in new drug development

## Proactive Approach



# Application

## Drug selection/avoidance

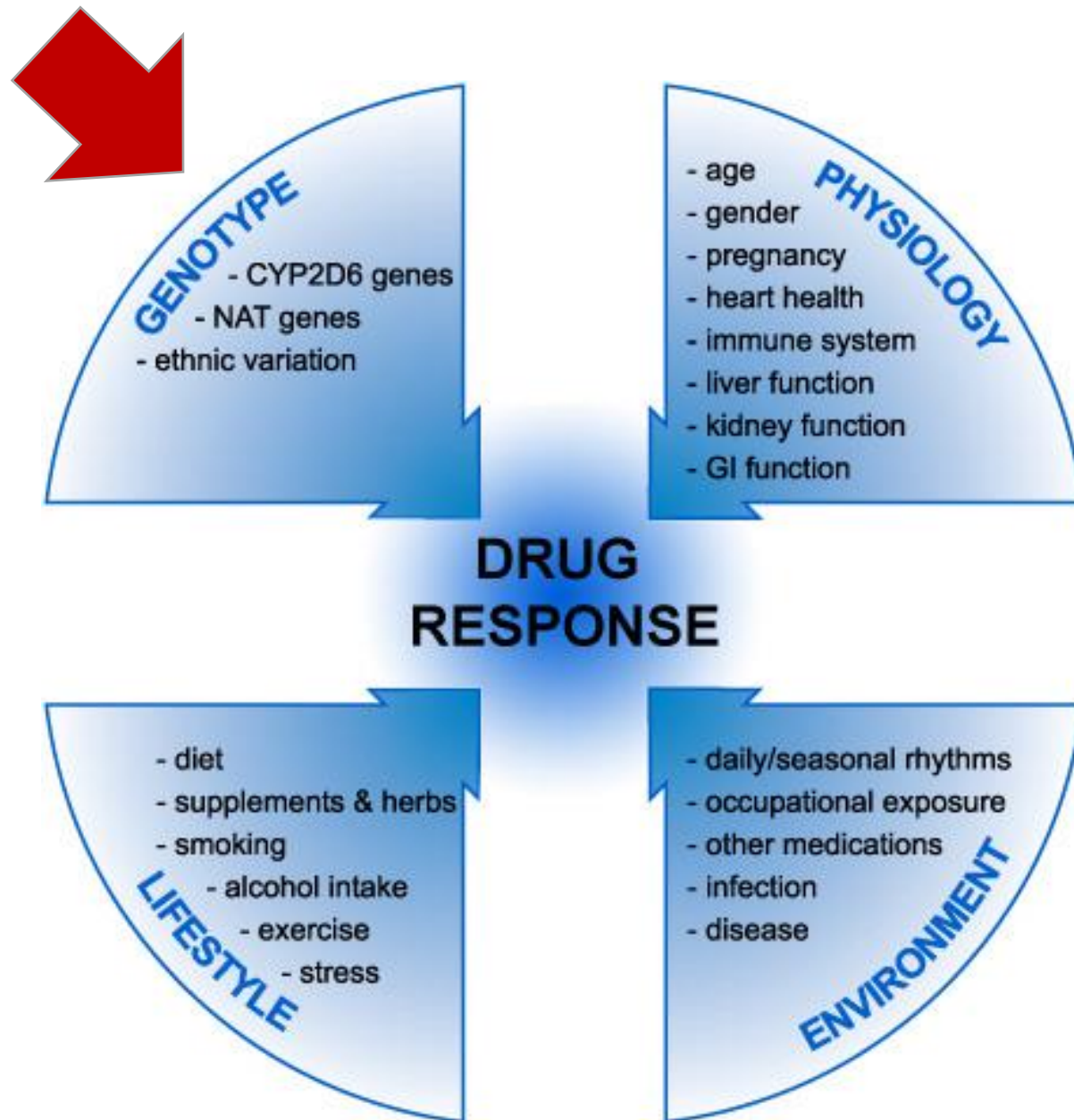
- » Who is at high risk of a serious ADR
- » Who is not likely to respond

## Dose optimization

- » Who is likely to be sensitive or resistant to a drug
- » What dose and what frequency is needed



# Factors Influencing Drug Response





# Frequency of PGx variants

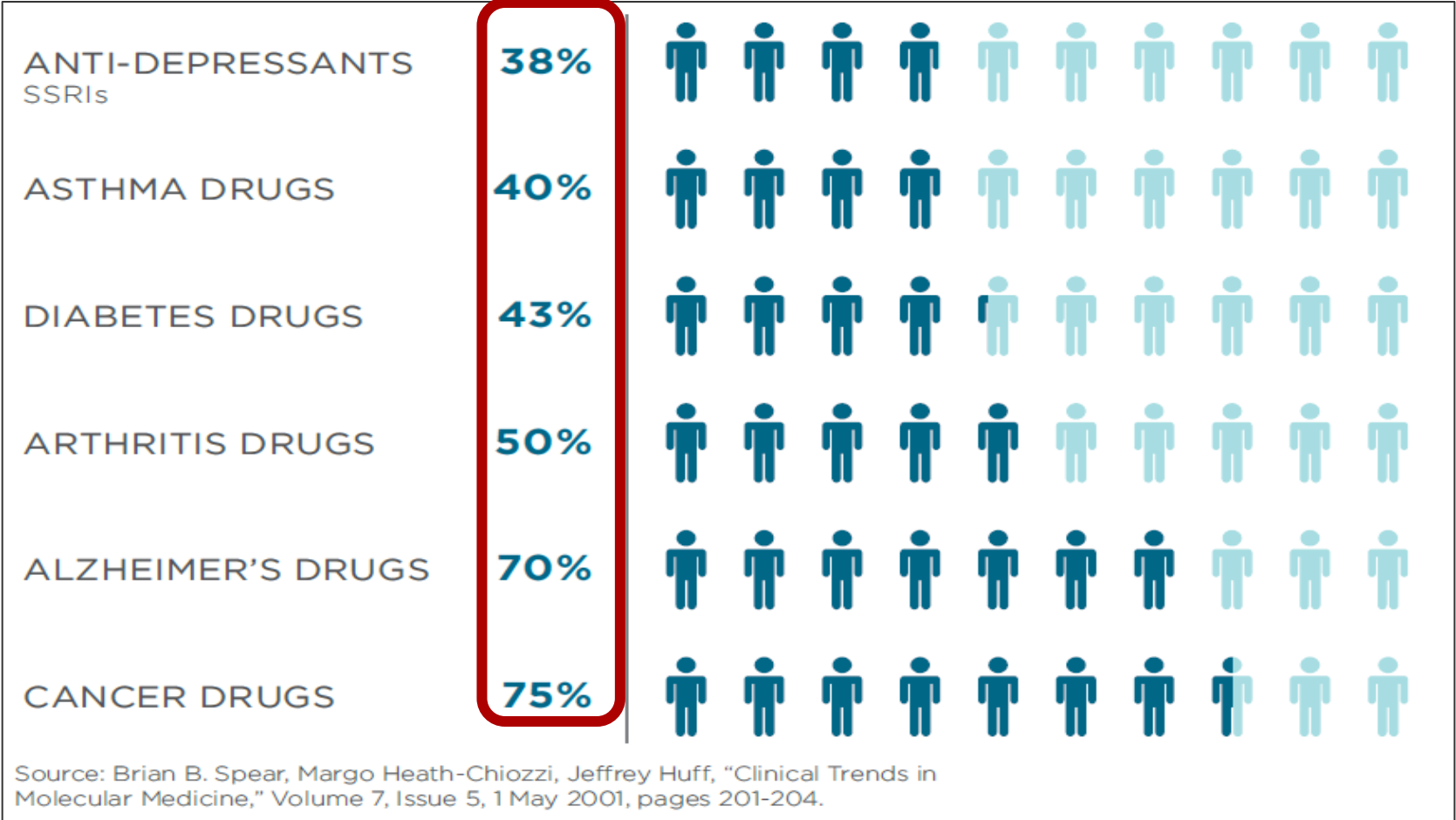
Around 97–98% of people have at least one actionable variant in their drug-related genes.

The possibility of the presence of a genetic variant [mainly loss of function (LOF) variant] in pharmacogenes is 93% for every individual



Genetic variation in human drug-related genes (2017)

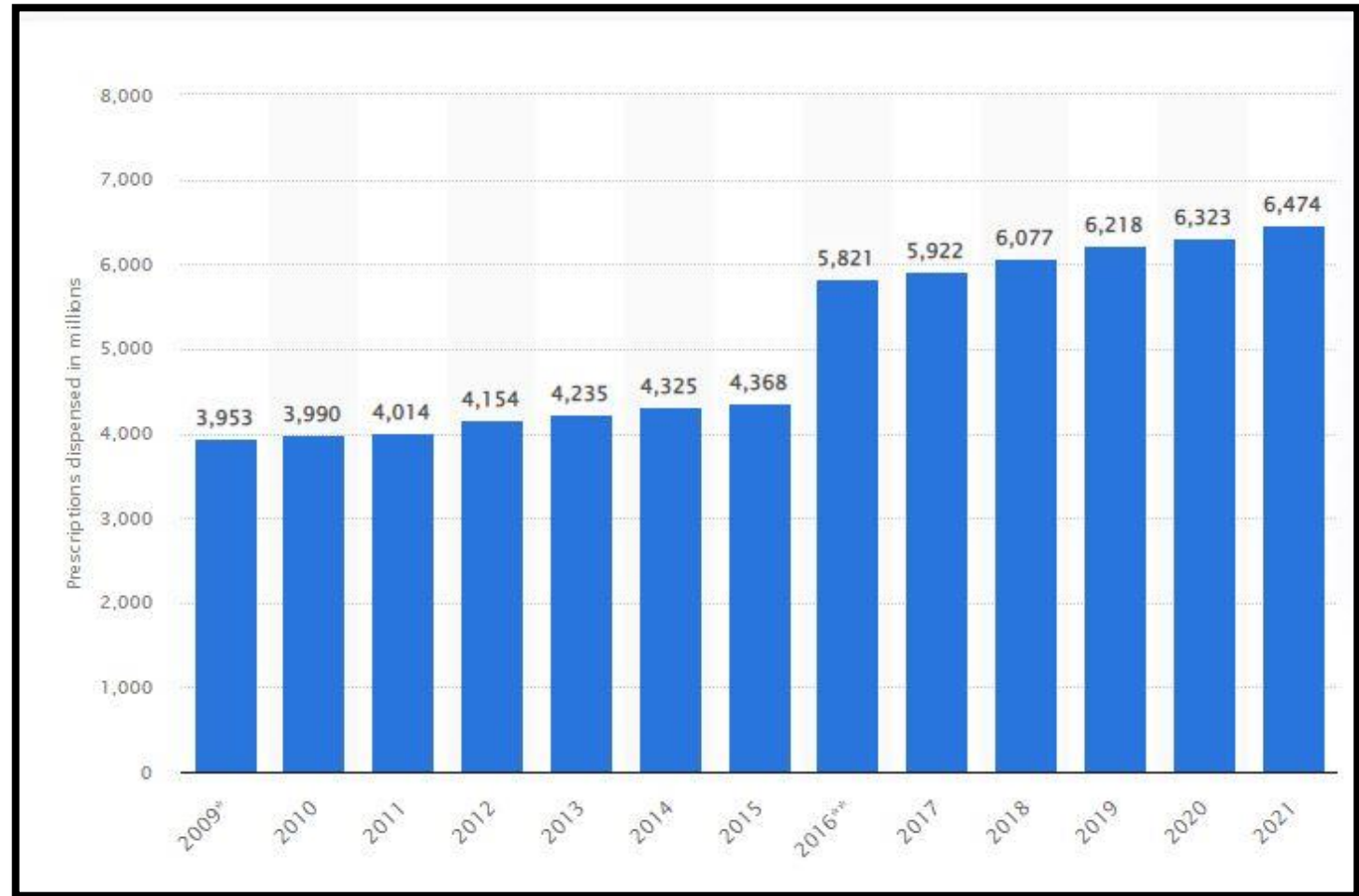
# Percentage of patient population for which a drug class is effective



<http://www.personalizedmedicinecoalition.org/>

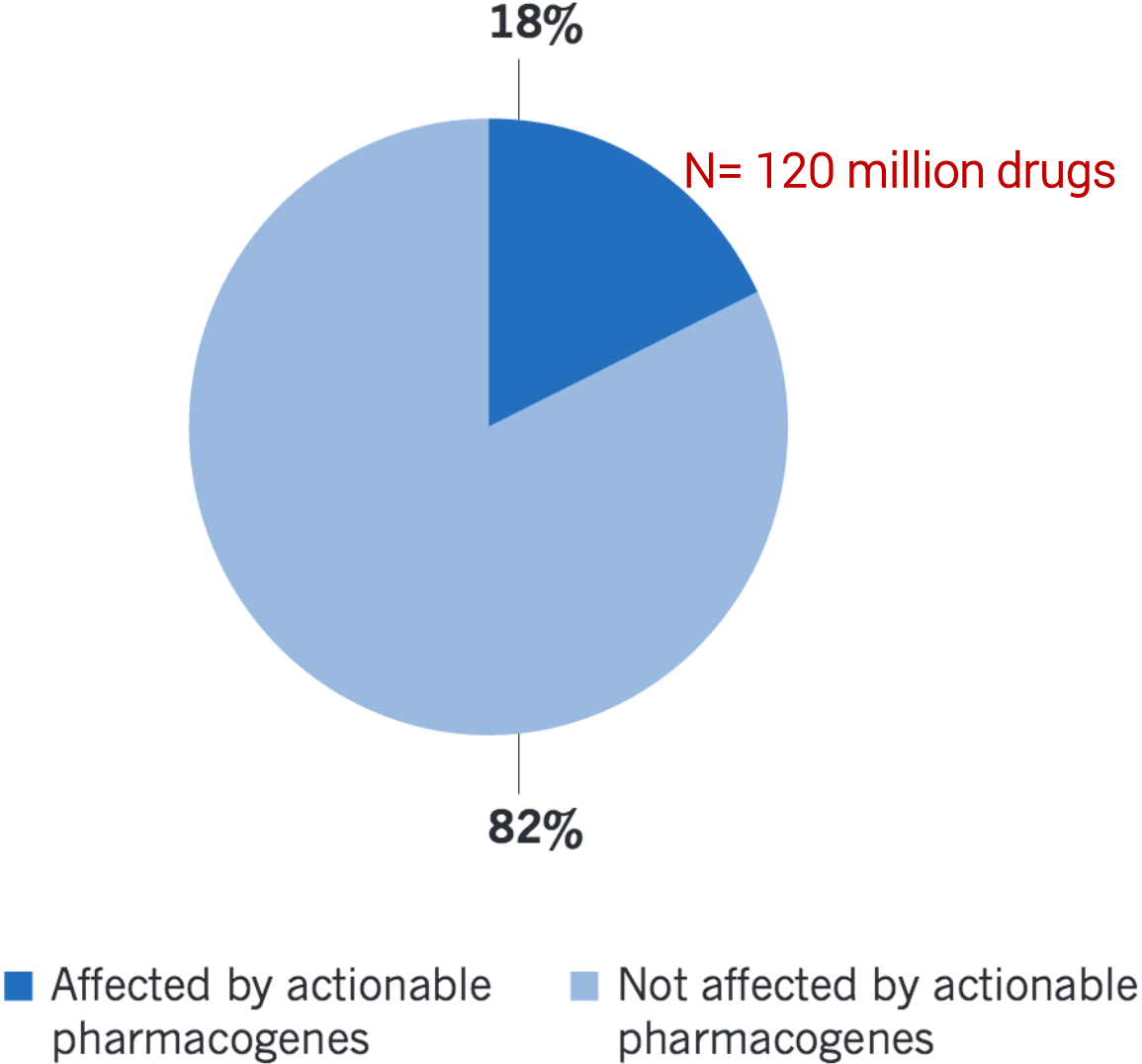


- In 2009 the number of prescriptions dispensed was near 3.95 billion.
- In 2021 the number of prescriptions dispensed was around 6.47 billion.



# Drugs prescribed in USA

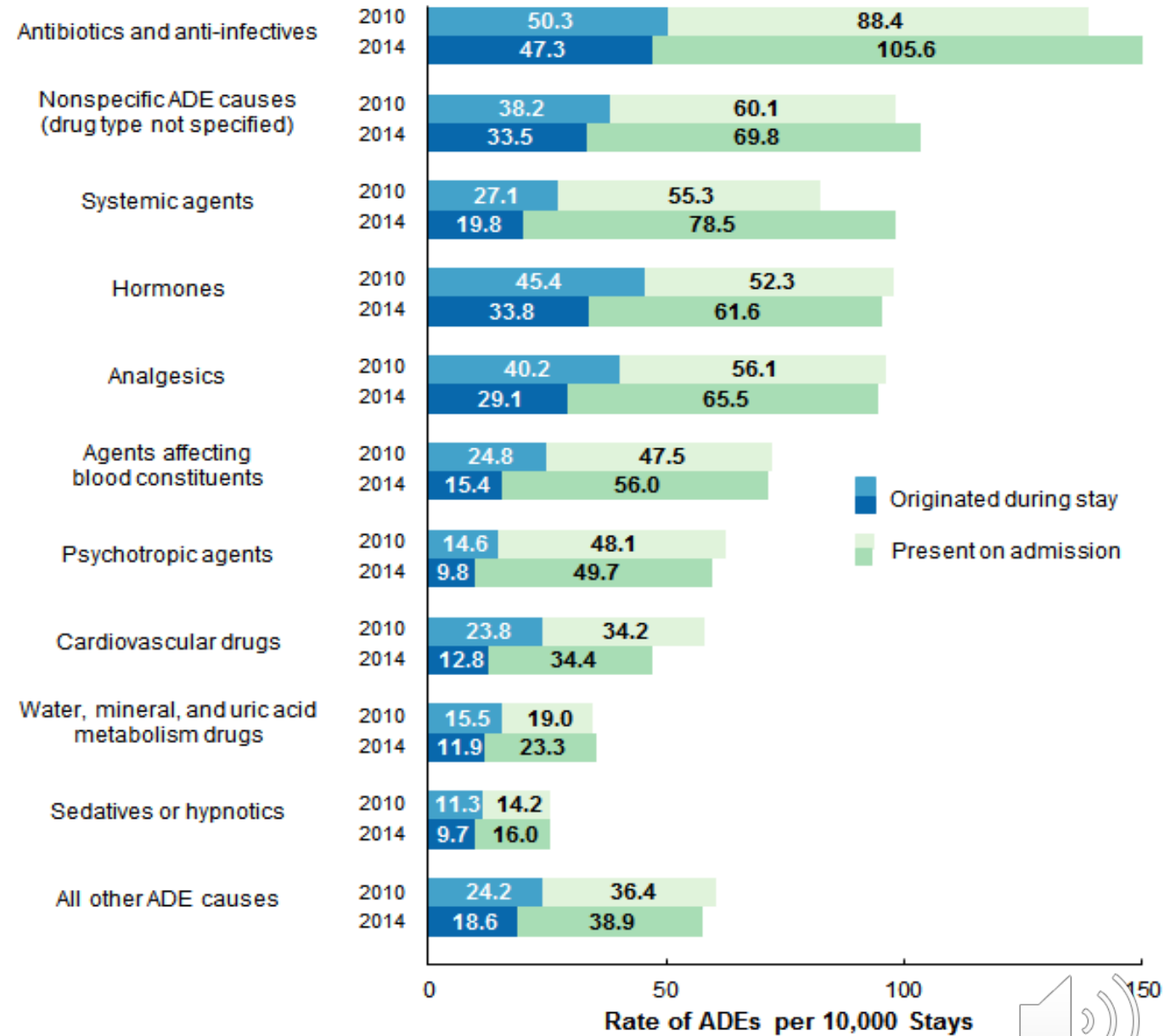
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# Adverse Drug Events (ADE)

- ADEs: leading type of nonsurgical adverse event occurring in hospitals in the US
- Some ADEs are the result of medication errors, but also may occur when medications are taken correctly.
- Patients hospitalized with an ADE have an increased length of stay, higher costs, and increased risk of in-hospital death.



# Adverse Drug Events: ADE

- 4th Leading cause of death ahead of pulmonary disease, diabetes, AIDS, pneumonia, accidents and automobile deaths.
- 100.000 deaths due to ADE per year
- \$136 Billion Costs of ADEs per year
- Large percentage is PREVENTABLE



# Adverse Drug Events: ADE

- Dose-related (Augmented)
  - Non-dose-related (Bizarre)
  - Dose-related and time-related (Chronic)
  - Time-related (Delayed)
  - Withdrawal (End of use)
  - Failure of therapy (Failure)
- 
- **Examples:** Rashes, jaundice, anemia, leucopenia, kidney damage, nerve injury or anaphylaxis
  - **\*Life threatening:** Stevens-Johnson syndrome or toxic epidermal necrolysis



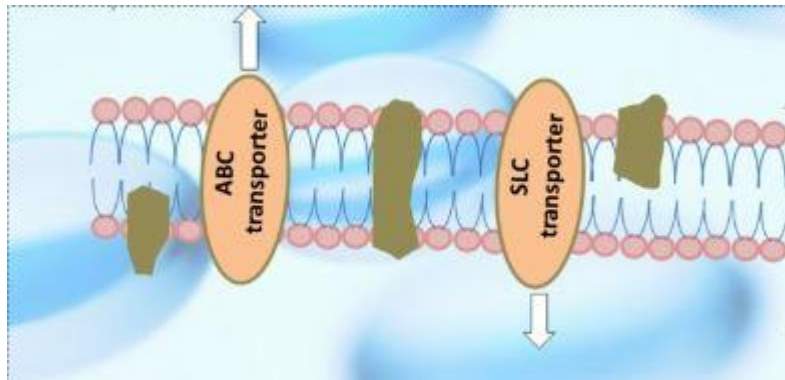
# How do we study pharmacogenetics?

*Pharmacogenes (focus on functional variants)*

## Drug transporters: cell surface proteins

The solute carrier (SLC) transporters e.g SLC01B1

The ATP-binding cassette (ABC) transporters e.g.  
ABCB1





# How do we study pharmacogenetics?

## *Pharmacogenes*

Drug transporters: cell surface proteins

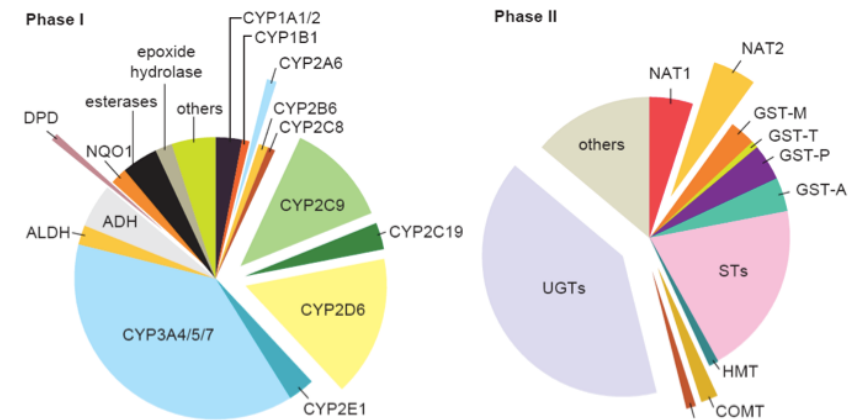
The solute carrier (SLC) transporters e.g. SLC01B1

The ATP-binding cassette (ABC) transporters e.g. ABCB1

## Drug Metabolizing Enzymes

Phase 1 metabolizing enzymes: e.g. CYP2D6, CYP3A5

Phase 2 metabolizing enzymes: TPMT, UGT1A1



Phase 1 – Oxidation, Reduction, Methylation, Hydroxylation, Deamination

Phase 2 – Conjugation (D-glucuronidation, O-sulfation, N-acetylation, O-, N-, S-methylation, glutathione, amino acid conjugation)



# How do we study pharmacogenetics?

## *Pharmacogenes (focus on functional variants)*

Drug transporters: cell surface proteins

The solute carrier (SLC) transporters e.g. SLC01B1

The ATP-binding cassette (ABC) transporters e.g. ABCB1

Drug Metabolizing Enzymes

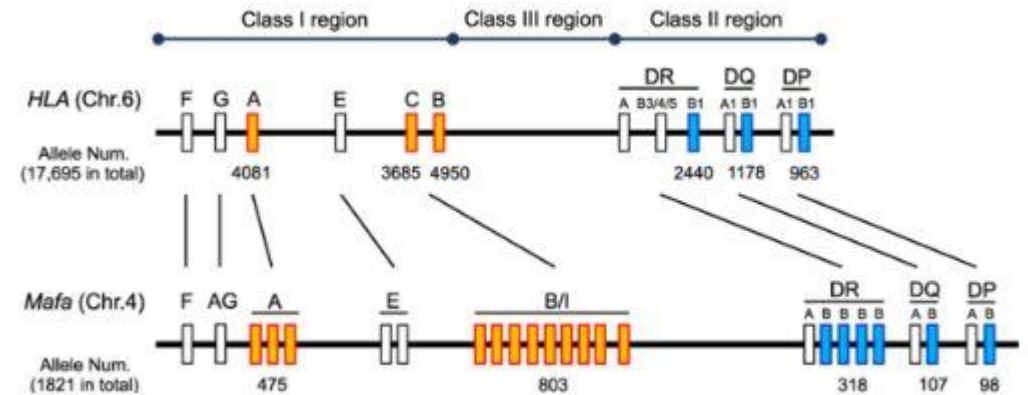
Phase 1 metabolizing enzymes: e.g. CYP2D6, CYP3A5

Phase 2 metabolizing enzymes: TPMT, UGT1A1

## Major histocompatibility complex genes (HLA alleles)

Some associated with increased risk for an allergic response to certain medications leading to Stevens-Johnson syndrome or toxic epidermal necrolysis.

HLA-B\*57:01, HLA-B\*15:02

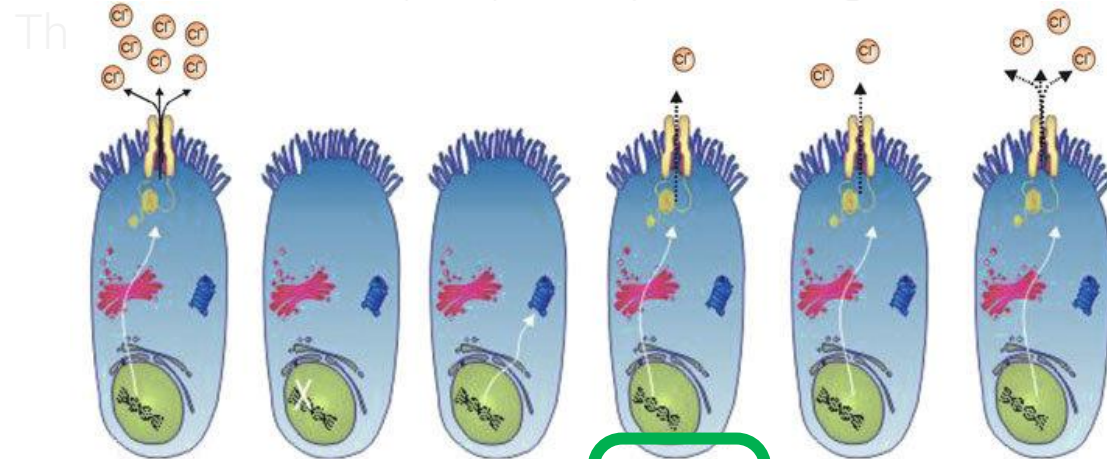


# How do we study pharmacogenetics?

## *Pharmacogenes (focus on functional variants)*

Drug transporters: cell surface proteins

The solute carrier (SLC) transporters e.g. SLC01B1



| Normal               | Class I   | Class II                           | Class III                         | Class IV                          | Class IV                                    |
|----------------------|---|------------------------------------|-----------------------------------|-----------------------------------|---|
| Defect               | Defective Synthesis                             | Defective processing or maturation | Defective regulation              | Defective conductance             | Reduced synthesis and stability             |
| Therapy              | Readthrough                                     | Correctors (+ potentiators)        | Potentiators                      | Potentiators                      | Potentiators Spicing modulators             |
| Mutations (examples) | G542X<br>W1282X<br>R553X, E882X<br>621 + 1G → T | ΔF508<br>N1303K<br>ΔI507<br>R1066C | G551D<br>G551S<br>G178R<br>G1244E | R117H<br>R354W<br>R347P<br>R1070W | 3272 6A→G<br>A455E<br>D565G<br>3849+1kb C→T |

Drug Metabolizing Enzymes

Phase 1 metabolizing enzymes: e.g. CYP2D6, CYP3A5

Phase 2 metabolizing enzymes: TPMT, UGT1A1

## Drug targets

- Molecules or pathways that a drug is designed to affect in order to deliver therapy.
- Work by altering the amount of the target protein or by delivering therapy only to specific genetic variants.
- e.g. Ivacaftor to treat cystic fibrosis



# Pharmacogenes variations

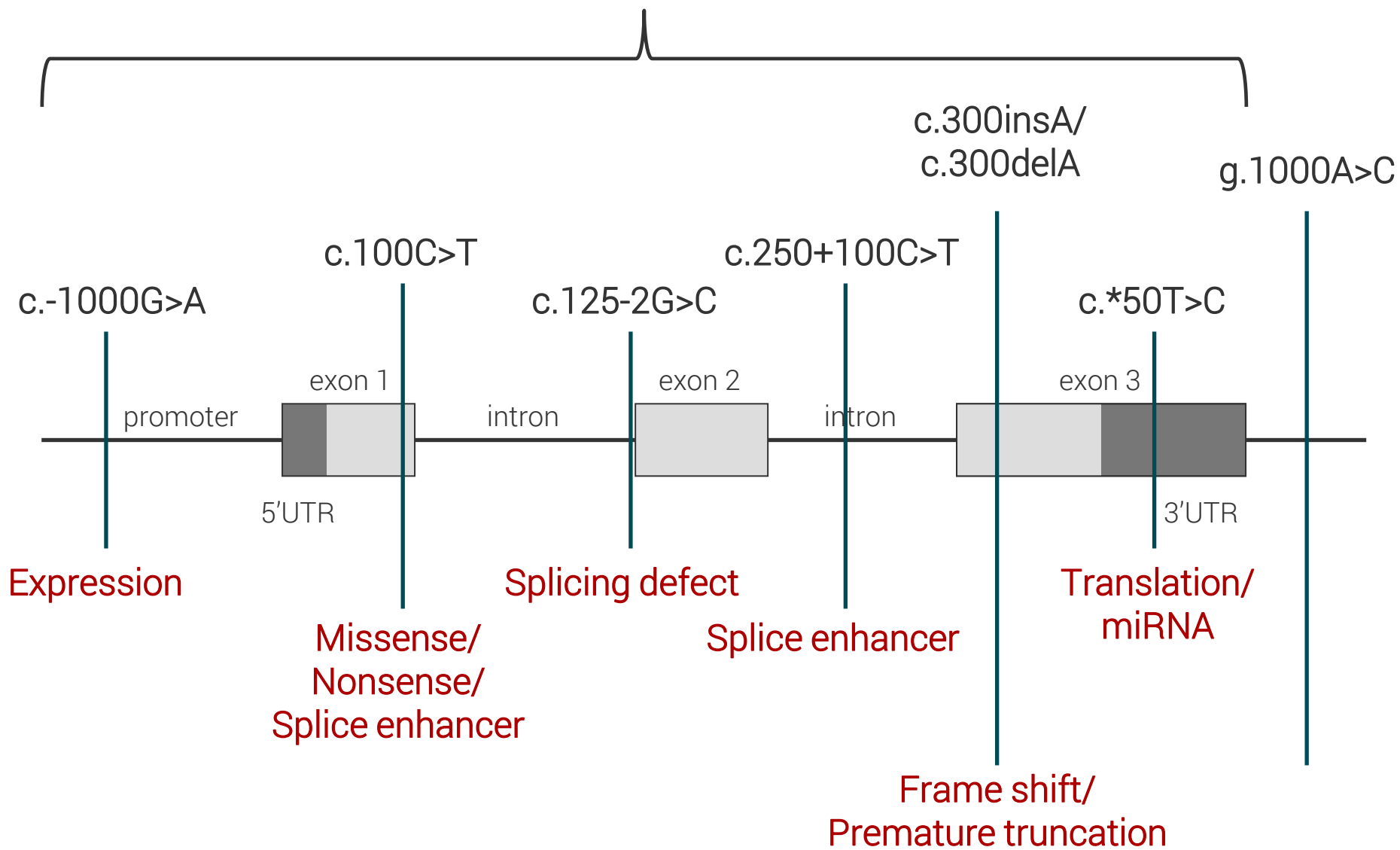
- SNPs
- Small Insertion and deletions
- Structural variants
- CNV: e.g. *CYP2D6*
- Tandem repeats e.g. *UGT1A1*

**Table 2** Counts of Ensembl consequence type for variants mapped to canonical transcripts of PGRNseq captured genes

| ENSEMBL consequence type                   | IN PGx | IN 1KG | IN EXAC |
|--|--------|--------|---------|
| Upstream Gene Variant                      | 6,094  | 2,122  | 23      |
| Intron Variant                             | 5,542  | 2,016  | 460     |
| Missense Variant                           | 4,806  | 1,485  | 1,792   |
| 3 Prime UTR Variant                        | 4,245  | 1,539  | 65      |
| Downstream Gene Variant                    | 3,574  | 1,239  | 44      |
| Synonymous Variant                         | 3,147  | 1,335  | 1,255   |
| 5 Prime UTR Variant                        | 931    | 287    | 59      |
| Missense Variant, Splice Region Variant    | 147    | 48     | 62      |
| Splice Region Variant, Intron Variant      | 142    | 60     | 49      |
| Stop Gained                                | 97     | 20     | 31      |
| Splice Region Variant, Synonymous Variant  | 90     | —      | 36      |
| Splice Acceptor Variant                    | 18     | 5      | 3       |
| Splice Donor Variant                       | 15     | 3      | 6       |
| Splice Region Variant, 5 Prime UTR Variant | 14     | 3      | 3       |
| Initiator Codon Variant                    | 11     | 2      | 2       |
| Stop Gained, Splice Region Variant         | 3      | 1      | 1       |
| Stop Lost                                  | 2      | —      | —       |
| Stop Retained Variant                      | 1      | 1      | —       |
| Splice Region Variant, 3 Prime UTR Variant | 1      | 1      | —       |
| Total                                      | 28,880 | 10,167 | 3,891   |



# PGx 'GENE'





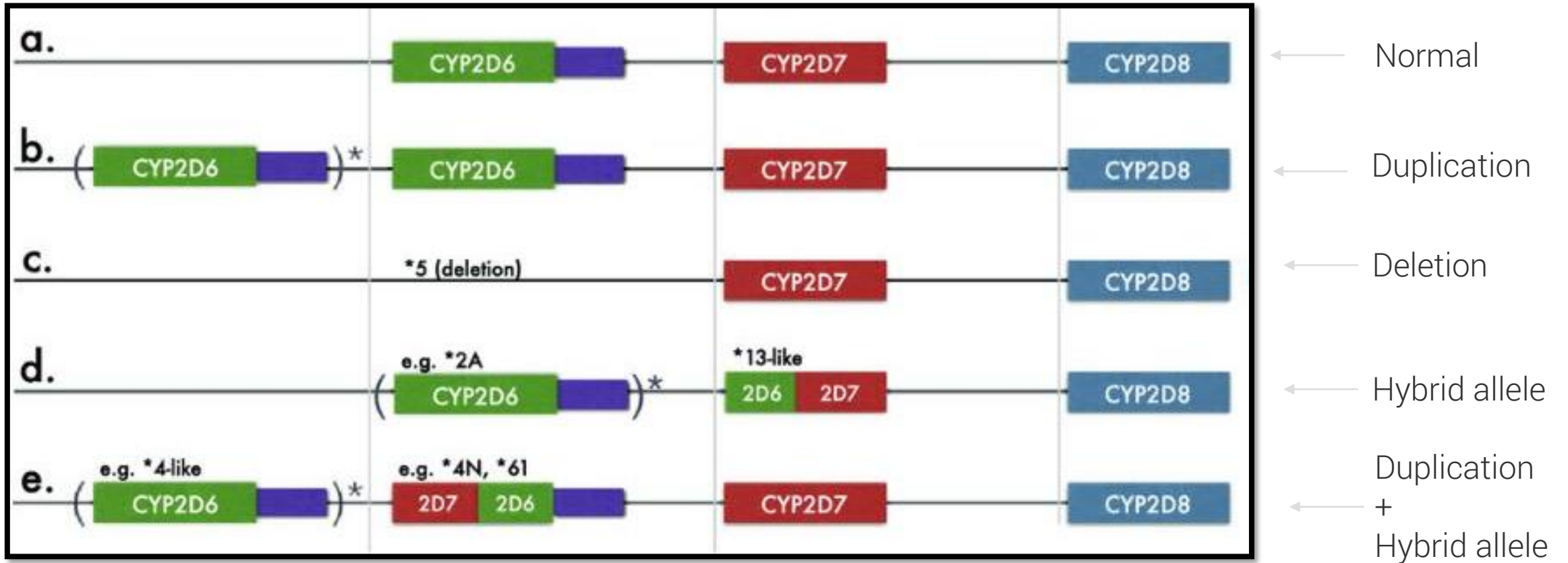
## Example variants and predicted function

### Cytochrome P450 2D6 (CYP2D6) alleles and their effects on CYP2D6 enzyme activity

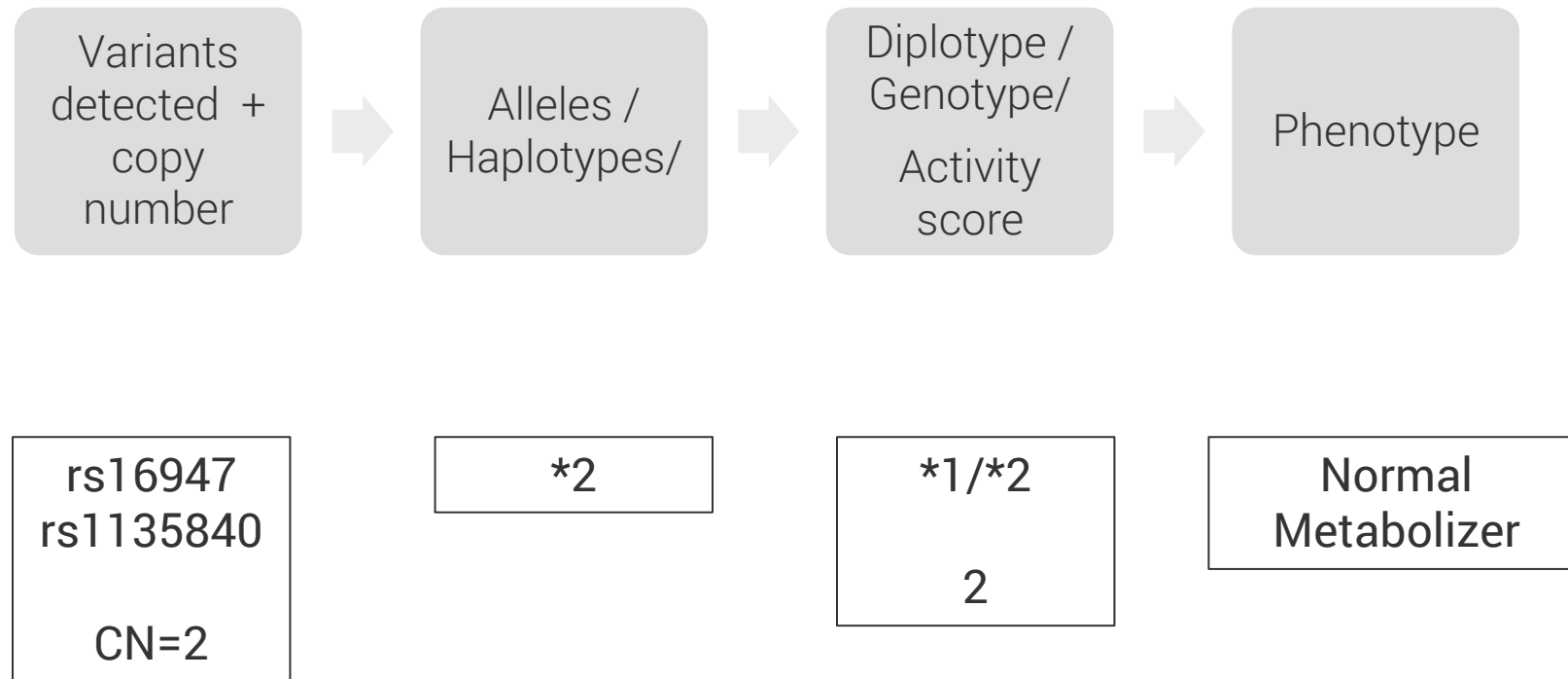
| CYP2D6 alleles   | Allele designation  | Enzyme activity                       | Allele abbreviation |
|--|---------------------|---------------------------------------|---------------------|
| *1, *2, *33, *35   | Normal or wild type | Normal                                | EM                  |
| *3, *4, *5-8, *11-16, *18-21,<br>*36, *38, *40, *42, *44, *56, *62 | Null                | No protein, inactive<br>or negligible | PM                  |
| *9, *10, *17, *29, *41, *59  | Reduced activity    | Decreased                             | IM                  |
| *22-28, *30-32, *34, *37,<br>*39, *43, *45-55                      | Unknown activity    | Unknown                               | Not applicable      |



# Example copy number variation/re-arrangements e.g CYP2D6

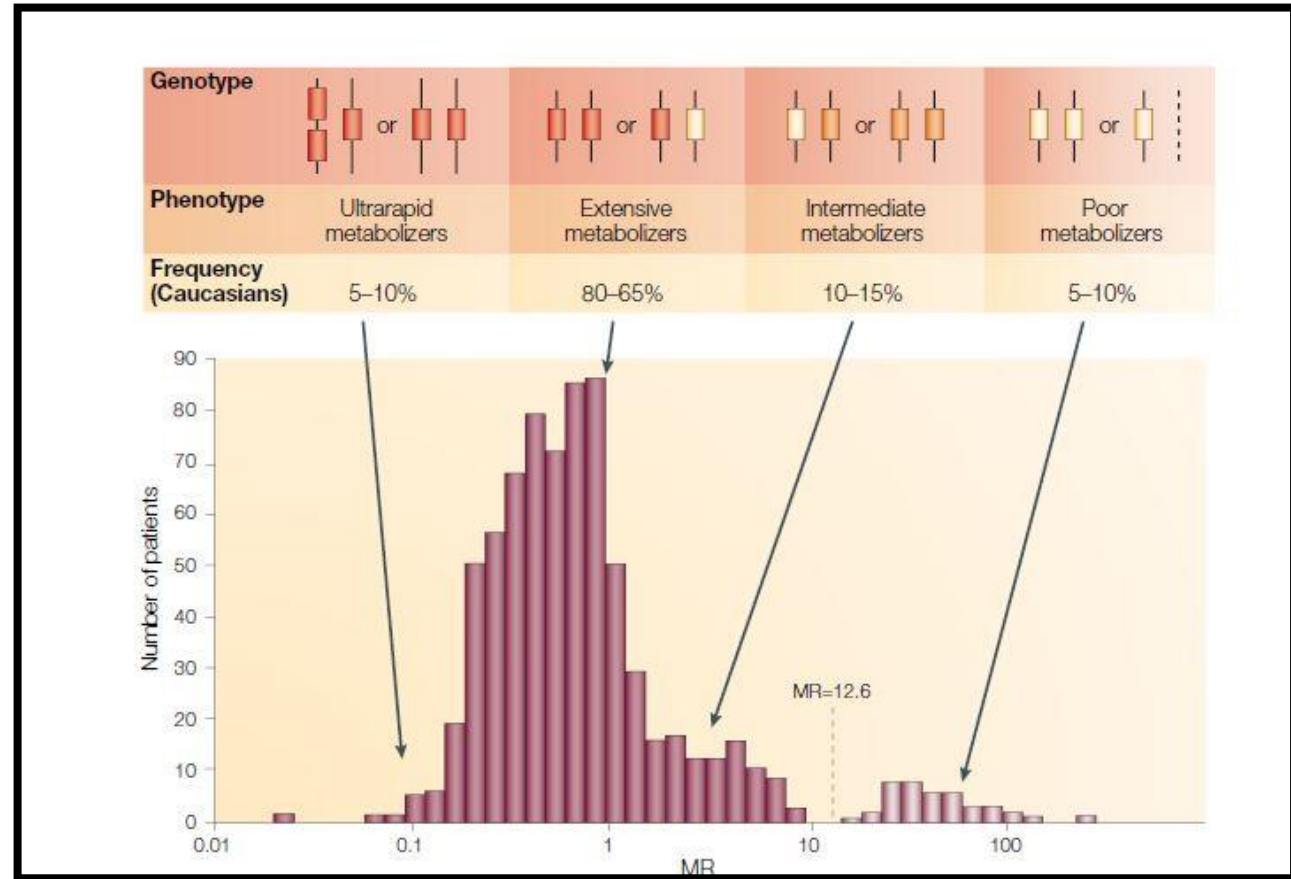


# How is the phenotype determined?



# Genetic variation in pharmacogenes and its effect on protein function is translated into “Metabolizer phenotype”

|                     |   |
|---------------------|---|
| <b>Ultra-rapid</b>  | Extreme metabolic activity, which may result in poor efficacy and therapeutic failure of the drug |
| <b>Extensive</b>    | Normal to high metabolic activity   |
| <b>Intermediate</b> | Impaired or slow metabolic activity   |
| <b>Poor</b>         | Low to absent metabolic activity, which may result in a higher risk of toxicity                   |



# Resources for PGx Knowledge

- National and international drug agencies
  - » FDA, EMA, PMDA
- Professional consensus guidelines
  - » CPIC, DPWG, CPNDS
  - » Professional societies (AMP, ACMG)
- Peer-reviewed literature





Objective:

Create, curate, and post freely available, peer-reviewed, evidence-based, updatable, and detailed **gene/drug clinical practice guidelines**



|  |   |  |
|--|---|--|
| <a href="#"><i>CYP2C9, HLA-B and Phenytoin</i></a>                                 | fosphenytoin<br>phenytoin   | <a href="#">CYP2C9</a><br><a href="#">HLA-B</a>                            |
| <a href="#"><i>CYP2C9, VKORC1, CYP4F2 and Warfarin</i></a>                         | warfarin  | <a href="#">CYP2C9</a><br><a href="#">CYP4F2</a><br><a href="#">VKORC1</a> |
| <a href="#"><i>CYP2D6 and Atomoxetine</i></a>                                      | atomoxetine   | <a href="#">CYP2D6</a>   |
| <a href="#"><i>CYP2D6 and Ondansetron and Tropisetron</i></a>                      | ondansetron<br>tropisetron  | <a href="#">CYP2D6</a>   |
| <a href="#"><i>CYP2D6 and Tamoxifen</i></a>  | tamoxifen   | <a href="#">CYP2D6</a>   |
| <a href="#"><i>CYP2D6, CYP2C19 and Selective Serotonin Reuptake Inhibitors</i></a> | citalopram<br>escitalopram<br>fluvoxamine<br>paroxetine<br>sertraline | <a href="#">CYP2C19</a><br><a href="#">CYP2D6</a>                          |



**Table 1 Assignment of likely CYP2D6 phenotypes based on genotypes**

| Phenotype <sup>a</sup>   |                | Genotype  | Examples of CYP2D6 diplotypes <sup>b</sup> |
|--|----------------|---|--|
| Metabolizer  | Activity score |   |  |
| CYP2D6 ultrarapid metabolizer  | > 2.0          | An individual carrying duplications of functional alleles   | *1/*1xN, *1/*2xN, *2/*2xN <sup>f</sup>     |
| CYP2D6 normal metabolizer  | 1.5 and 2.0    | An individual carrying two normal function alleles or one normal function and one decreased function allele   | *1/*1, *1/*2, *1/*9, *1/*41, *2/*2,        |
| CYP2D6 normal metabolizer or intermediate metabolizer (controversy remains) <sup>d</sup> | 1.0            | An individual carrying two decreased function alleles or one normal function and one no function allele.<br><i>An activity score (AS) of 1.0 is associated with decreased tamoxifen metabolism to endoxifen compared to those with an AS of 1.5 or 2.</i> | *1/*4, *1/*5, *41/*41                      |
| CYP2D6 intermediate metabolizer  | 0.5            | An individual carrying one decreased function and one no function allele  | *4/*10, *4/*41, *5/*9                      |
| CYP2D6 poor metabolizer  | 0              | An individual carrying only no functional alleles   | *3/*4, *4/*4, *5/*5, *5/*6                 |



**Table 2 Dosing recommendations for tamoxifen based on CYP2D6 phenotype**

| Phenotype                       |                | Implications  | Therapeutic recommendation <sup>b</sup>   | Classification of recommendation <sup>a</sup> |
|---------------------------------|----------------|---|---|---|
| Metabolizer status              | Activity score |   |   |   |
| CYP2D6 ultrarapid metabolizer   | >2.0           | Therapeutic endoxifen concentrations  | Avoid moderate and strong CYP2D6 inhibitors. Initiate therapy with recommended standard of care dosing (tamoxifen 20 mg/day).   | Strong  |
| CYP2D6 intermediate metabolizer | 0.5            | Lower endoxifen concentrations compared to normal metabolizers; higher risk of breast cancer recurrence, event-free and recurrence-free survival compared to normal metabolizers. | Consider hormonal therapy such as an aromatase inhibitor for postmenopausal women or aromatase inhibitor along with ovarian function suppression in premenopausal women, given that these approaches are superior to tamoxifen regardless of CYP2D6 genotype. <sup>43</sup> If aromatase inhibitor use is contraindicated, consideration should be given to use a higher but FDA approved tamoxifen dose (40 mg/day). <sup>45</sup> Avoid CYP2D6 strong to weak inhibitors. | Moderate                                      |



# Methods in pharmacogenetics testing

- DNA microarray (allele-specific hybridization)
- Invader assay (cleavage-based with endonuclease enzyme)
- Mass spectrometry
- PCR-RFLP (restriction endonuclease)
- Pyrosequencing (primer extension)
- Sanger sequencing (chain termination)
- SNaPShot (primer extension)
- TaqMan (allele-specific hybridization)
- Next Generation Sequencing

Cost

Throughput

Required prior knowledge



# SNV Panels

- SNV panel testing is the most used technology in PGx practice
- Commercial-panels or custom panels
- Typically contain a preselected set of SNVs: few variants in specific genes OR genome-wide
- Typically contain variants linked to drug response in PGx guidelines or on PharmGKB
- The evidence underlying the selected variants can vary from only the most strongly associated variants, to containing all variants potentially or theoretically associated with drug response
- Quick result at low costs
- Most have no CNV detection, no phasing, and no hybrid detection





# Commercial vs Custom panels

## Why the need?

- Many commercial arrays contain a high number of variants, making a fast turnaround time and interpretation challenging.
- Commercial arrays would include variants which may not be of direct interest in a clinical setting due to lack of evidence of clinical utility

## Solution

- Many institutes choose to customize a panel with genes/variants of interest.

# Genome wide panels

- Offers genome wide coverage + PGx coverage
- Hundreds of thousands of markers
- Can miss specific alleles for PGx genes and CN which is not ideal



# Next Generation Sequencing

- NGS technologies are not yet routinely applied in clinical PGx. e.g. in a recent market analyses out of 25 labs, only two used NGS
- Many research studies conducted using NGS for PGx
- While SNV panels only cover a limited set of selected variants, sequencing data cover the full exome or genome or targeted panels

## The Identification of Novel *CYP2D6* Variants in US Hmong: Results From Genome Sequencing and Clinical Genotyping

Ya Feng Wen<sup>1</sup>, Andrea Gaedigk<sup>2,3</sup>, Erin C. Boone<sup>2</sup>, Wendy Y. Wang<sup>2</sup> and Robert J. Straka<sup>1\*</sup>

<sup>1</sup>Department of Experimental and Clinical Pharmacology, College of Pharmacy, University of Minnesota, Twin Cities, MN, United States, <sup>2</sup>Division of Clinical Pharmacology, Toxicology and Therapeutic Innovation, Children's Hospital of Philadelphia, Philadelphia, PA, United States, <sup>3</sup>School of Medicine, University of Missouri-Kansas City, Kansas City, MO, United States

ORIGINAL RESEARCH ARTICLE | Genetics in Medicine

### A curated gene list for reporting results of newborn genomic sequencing

Ozge Ceyhan-Birsoy, PhD<sup>1,2,3</sup>, Kalotina Machini, PhD<sup>1,2,3</sup>, Matthew S. Lebo, PhD<sup>1,2,3</sup>, Tim W. Yu, MD<sup>3,4,5</sup>, Pankaj B. Agrawal, MD, MMSC<sup>3,4,5</sup>, Richard B. Parad, MD, MPH<sup>3,7</sup>, Ingrid A. Holm, MD, MPH<sup>3,4</sup>, Amy McGuire, PhD<sup>3</sup>, Robert C. Green, MD, MPH<sup>3,8,10</sup>, Alan H. Beggs, PhD<sup>3,4</sup>, Heidi L. Rehm, PhD<sup>1,2,3,10</sup>, for the BabySeq Project

## Characterization of ADME Gene Variation in Colombian Population by Exome Sequencing

Daniel Felipe Silgado-Guzmán<sup>1†</sup>, Mariana Angulo-Aguado<sup>2†</sup>, Adrien Morel<sup>2</sup>, María José Niño-Orrego<sup>2</sup>, Daniel-Armando Ruiz-Torres<sup>2</sup>, Nora Constanza Contreras Bravo<sup>2</sup>, Carlos Martin Restrepo<sup>2</sup>, Oscar Ortega-Recalde<sup>2,4‡</sup> and Dora Janeth Fonseca-Mendoza<sup>2,4‡</sup>

<sup>1</sup>Department of Molecular Diagnosis, Genética Molecular de Colombia SAS, Bogotá, Colombia, <sup>2</sup>Center for Research in Genetics and Genomics—CIGGUR, GENIUIROS Research Group, School of Medicine and Health Sciences, Universidad Del Rosario, Bogotá, Colombia

## Applying Next-Generation Sequencing Platforms for Pharmacogenomic Testing in Clinical Practice

Alireza Tafazoli<sup>1,2</sup>, Henk-Jan Guchelaar<sup>3,4</sup>, Wojciech Milyk<sup>1</sup>, Adam J. Kretowski<sup>2,5</sup> and Jesse J. Swen<sup>3,4\*</sup>

## Assessing the capability of massively parallel sequencing for opportunistic pharmacogenetic screening

David Ng, MD<sup>1</sup>, Celine S. Hong, PhD<sup>1</sup>, Larry N. Singh, PhD<sup>1</sup>, Jennifer J. Johnston, PhD<sup>1</sup>, James C. Mullikin, PhD<sup>2,3</sup>, Leslie G. Biesecker, MD<sup>1,2</sup>; on behalf of the NISC Comparative Sequencing Program

## *CYP2C8*, *CYP2C9*, and *CYP2C19* Characterization Using Next-Generation Sequencing and Haplotype Analysis

A GeT-RM Collaborative Project



# NGS Panels

- Custom-capture panels of genes with associations to pharmacogenetic phenotypes.
- Generate deep coverage data
- >99% concordant with orthogonal datasets
- Identify novel, rare variants of interest. Value in research and clinical settings.
- Limitations:
  - ❖ Miss non-coding and complex structural variants for specific pharmacogenes (including CYP2A6, CYP2D6, and HLA-B)
  - ❖ Require better computational resources for data interpretation



## HHS Public Access

Author manuscript

*Pharmacogenet Genomics*. Author manuscript; available in PMC 2017 July 05.

Published in final edited form as:

*Pharmacogenet Genomics*. 2016 April ; 26(4): 161–168. doi:10.1097/FPC.0000000000000202.

### **PGRNseq: A Targeted Capture Sequencing Panel for Pharmacogenetic Research and Implementation**

Adam Gordon<sup>1</sup>, Robert S. Fulton<sup>3</sup>, Xiang Qin<sup>2</sup>, Elaine R. Mardis<sup>3</sup>, Deborah A. Nickerson<sup>1,\*</sup>, and Steve Scherer<sup>2</sup>

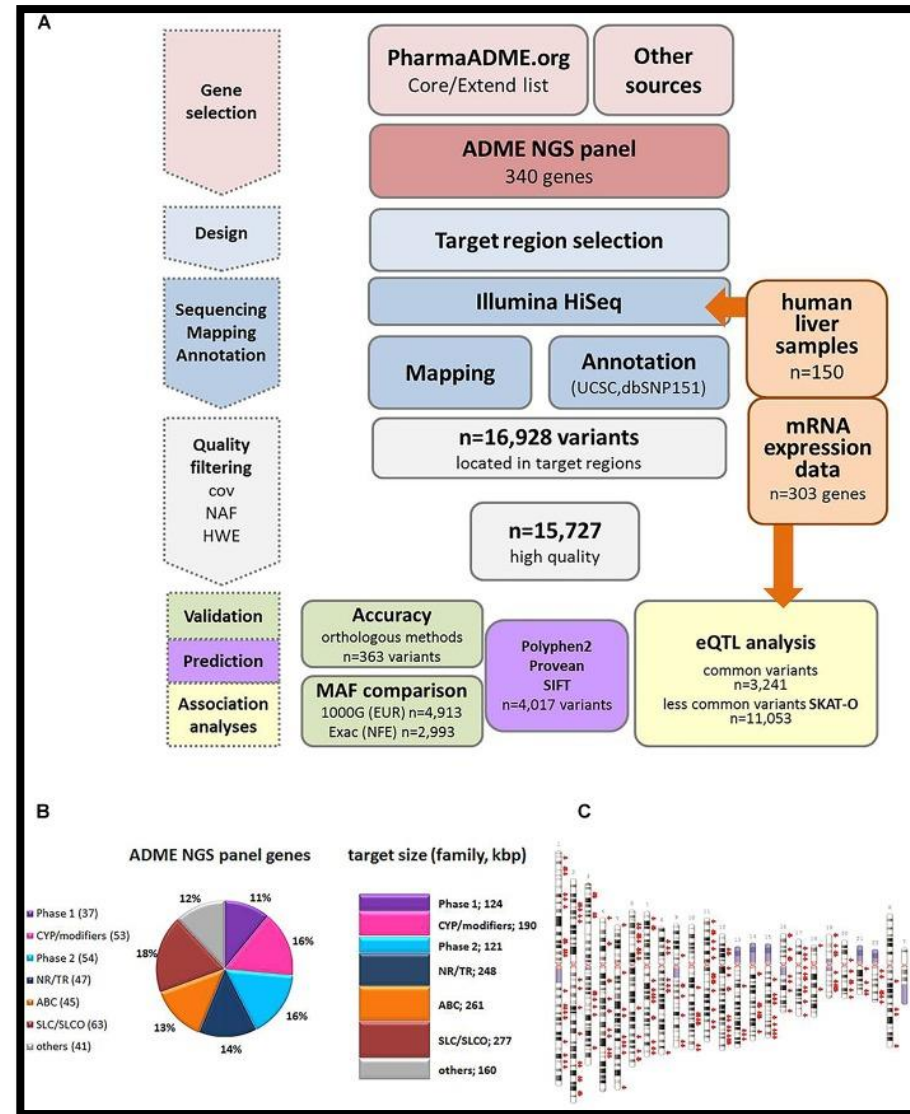


# NGS Panels

**A New Panel-Based Next-Generation Sequencing Method for ADME Genes Reveals Novel Associations of Common and Rare Variants With Expression in a Human Liver Cohort**

*Kathrin Klein<sup>1,2</sup>, Roman Tremmel<sup>1,2</sup>, Stefan Winter<sup>1,2</sup>, Sarah Fehr<sup>3,4</sup>, Florian Battke<sup>3,4</sup>, Tim Scheurenbrand<sup>3,4</sup>, Elke Schaeffeler<sup>1,2</sup>, Saskia Biskup<sup>3,4</sup>, Matthias Schwab<sup>1,2,5,6</sup> and Ulrich M. Zanger<sup>1,2\*</sup>*

- Genes coding for phase I and II enzymes, drug transporters and regulator/modifier genes
- Coding regions, adjacent introns, and 5' and 3' UTRs in flanking sequences
- >99% concordance.
- Very high read-depth
- Combined in-silico prediction with expression data, identified eQTLs .

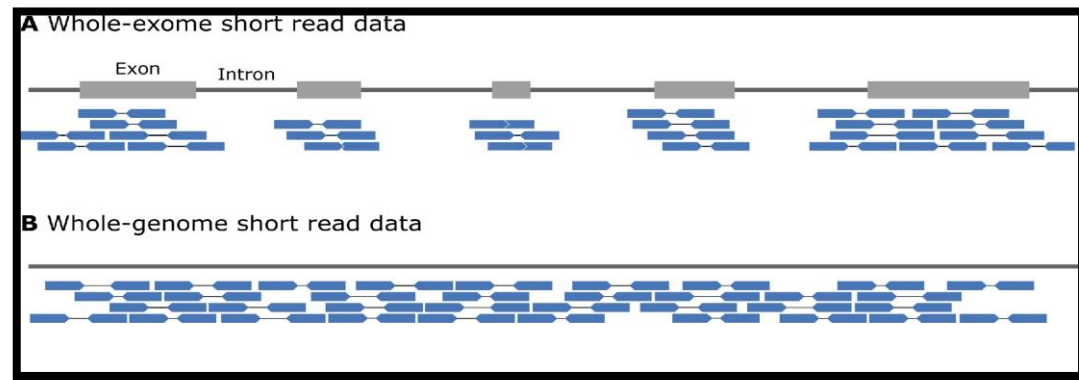




# WES and WGS (Short reads)

## Cross-Comparison of Exome Analysis, Next-Generation Sequencing of Amplicons, and the iPLEX® ADME PGx Panel for Pharmacogenomic Profiling

Eng Wee Chua<sup>1,2†</sup>, Simone L. Cree<sup>1†</sup>, Kim N. T. Ton<sup>1</sup>, Klaus Lehnert<sup>3</sup>, Phillip Shepherd<sup>4</sup>, Nuala Helsby<sup>5</sup> and Martin A. Kennedy<sup>1\*</sup>

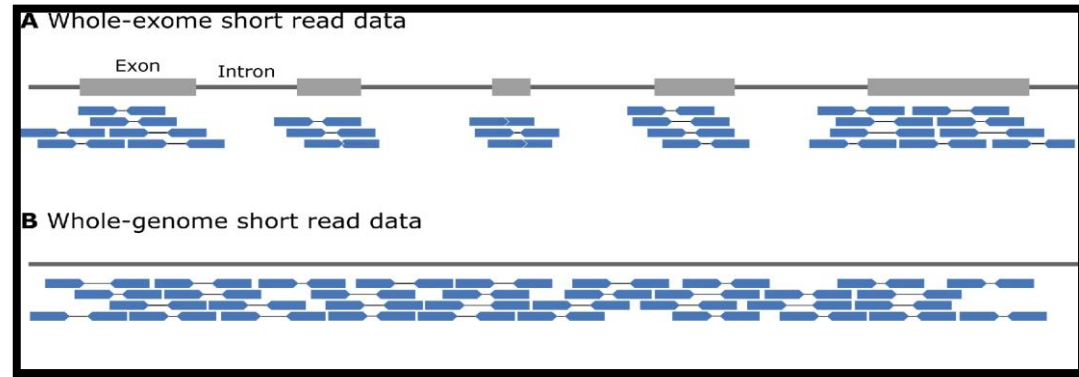


- Initial efforts: re-purpose already existent exomes or genomes to detect PGx variants.
- 94% concordance between PGx panel and WES
- 96% concordance between PGx panel and WGS.
- Some very important alleles could be missed by WES or WGS. e.g.
  - Non-coding: CYP2C19\*17 variants ; VKORC1, CYP2D6\*4 and \*41
  - CYP2D6 copy number variation
  - CYP2D6/2D7 hybrids
  - HLA-genes
- Short reads pose a limitation: the identification of structural variants, repetitive regions, phasing of alleles and distinguishing highly homologous regions.

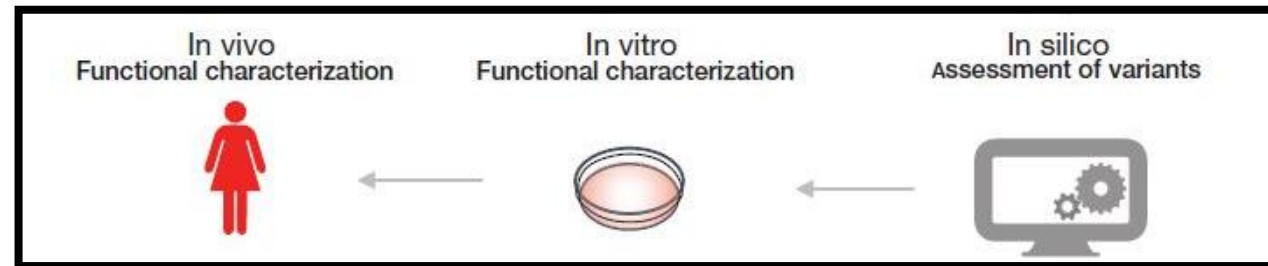




# WES and WGS (Short reads)

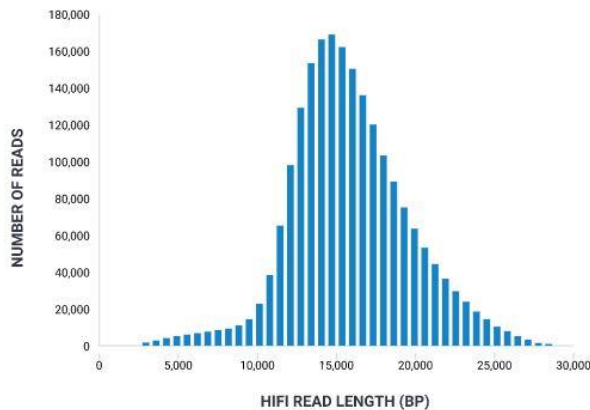


- May facilitate the discovery of novel loci (but will need a confirmative study or extensive invitro research to attribute potential, newly identified variants in a particular gene to drug response).
- WGS: Structural variants, non-coding, copy numbers..etc



# NGS (Long reads)


- Long-reads (>10 kilobase on average, sometimes tens to thousands of kb in length)
- Sequencing process occurs in real-time.
- Sequencing and library preparation without PCR amplification (no PCR bias)
- Two major technologies:
  - ✓ **Pacific Bioscience (PacBio) technology:** Uses SMRT (single molecule real-time)-sequencing
  - ✓ **Oxford Nanopore Technologies (ONT):** Nanopores through which the DNA strand is pulled, the disruption in the current is specific to a codon, allowing for the full assembly of the DNA sequence



## REVIEWS

 Check for updates

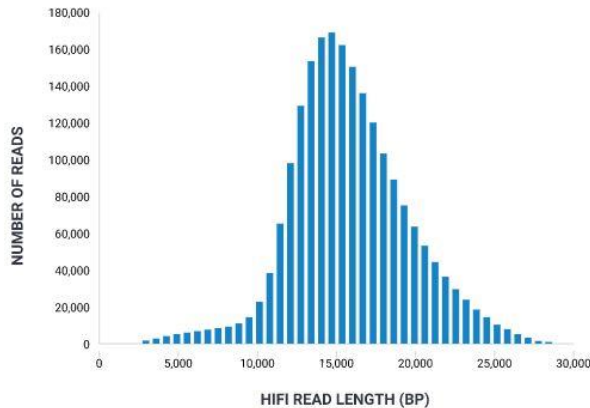
### Long-read human genome sequencing and its applications

Glenn A. Logsdon<sup>1</sup>, Mitchell R. Vollger<sup>1</sup> and Evan E. Eichler<sup>1,2</sup> 



# NGS (Long reads)

- ✓ **Pacific Bioscience (PacBio) technology:** Uses SMRT (single molecule real-time)-sequencing
- ✓ **Oxford Nanopore Technologies (ONT):** Nanopores through which the DNA strand is pulled, the disruption in the current is specific to a codon, allowing for the full assembly of the DNA sequence

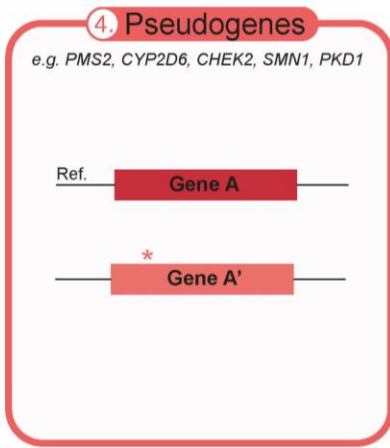
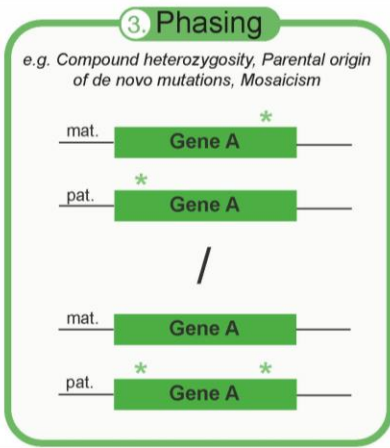
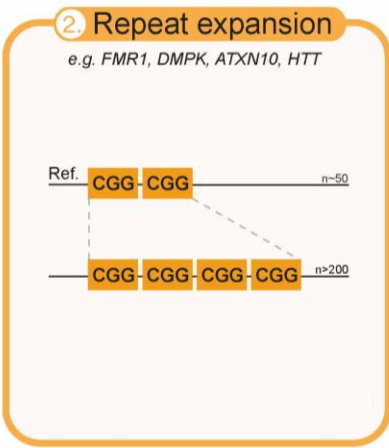
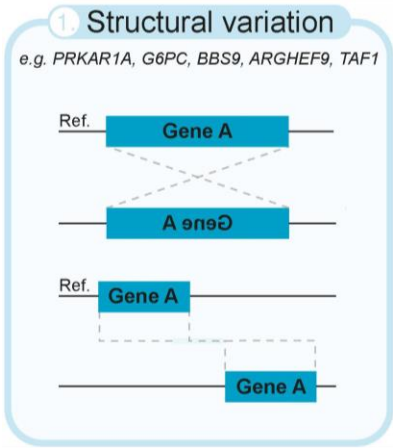
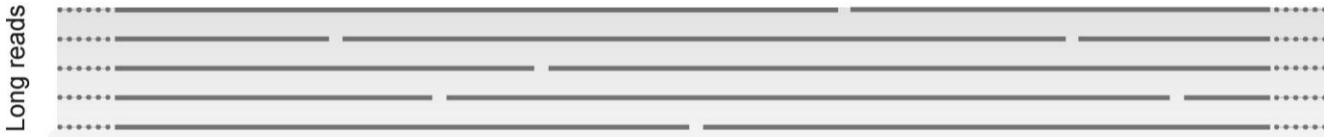
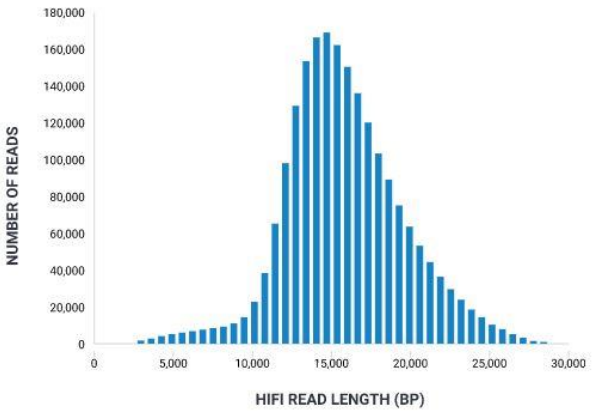


| Sequencing technology              | Platform           | Data type  | Read length (kb) |           | Read accuracy (%) | Throughput per flow cell (Gb) |         | Estimated cost per Gb (US\$) | Maximum throughput per year (Gb) <sup>a</sup> |
|------------------------------------|--------------------|------------|------------------|-----------|-------------------|-------------------------------|---------|------------------------------|---|
|                                    |                    |            | N50              | Maximum   |                   | Mean                          | Maximum |                              |   |
| Pacific Biosciences (PacBio)       | RS II <sup>b</sup> | CLR        | 5–15             | >60       | 87–92             | 0.75–1.5                      | 2       | 333–933 <sup>c</sup>         | 4,380   |
|                                    | Sequel             | CLR        | 25–50            | >100      |                   | 5–10                          | 20      |                              |   |
|                                    | Sequel II          | CLR        | 30–60            | >200      | >99               | 50–100                        | 160     | 13–26 <sup>e</sup>           | 93,440  |
|                                    |                    | HiFi       | 10–20            | >20       |                   | 15–30                         | 35      |                              |   |
| Oxford Nanopore Technologies (ONT) | MinION/ GridION    | Long       | 10–60            | >1,000    | 87–98             | 2–20                          | 30      | 50–500 <sup>f</sup>          | 21,900 (MinION)<br>109,500 (GridION)          |
|                                    |                    | Ultra-long | 100–200          | >1,500    |                   | 0.5–2                         | 2.5     |                              | 500–2,000 <sup>f</sup>                        |
|                                    | PromethION         | Long       | 10–60            | >1,000    |                   | 50–100                        | 180     | 21–42 <sup>f</sup>           | 3,153,600                                     |
| Illumina                           | NextSeq 550        | Single-end | 0.075–0.15       | 0.15      | >99.9             | 16–30                         | >30     | 50–63 <sup>g</sup>           | >47,782                                       |
|                                    |                    | Paired-end | 0.075–0.15 (×2)  | 0.15 (×2) |                   | 32–120                        | >120    |                              | 40–60 <sup>g</sup>                            |
|                                    | NovaSeq 6000       | Single-end | 0.05–0.25        | 0.25      |                   | 65–3,000                      | >3,000  | 10–35 <sup>h</sup>           | >1,194,545                                    |
|                                    |                    | Paired-end | 0.05–0.25 (×2)   | 0.25 (×2) |                   |                               |         |                              |   |



# NGS (Long reads)

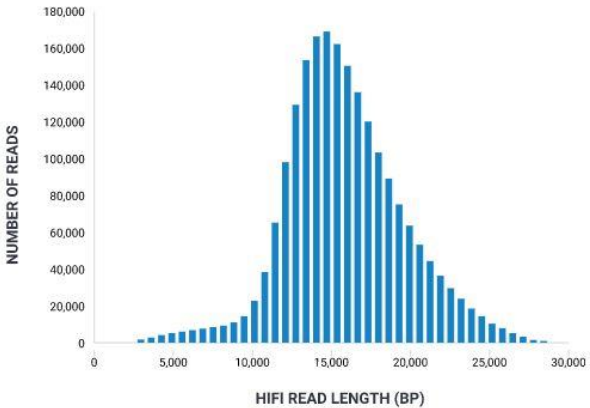
- Expensive
- Data processing is significantly more intensive
- Throughput and accuracy lower compared to short-reads (Gb/year)



# NGS (Long reads)

## Promise:

- Resolve some of the most challenging regions of the human genome
- Detect previously inaccessible structural variants
- Telomere-to-telomere assemblies of whole chromosomes



**1. Structural variation**  
e.g. *PRKAR1A, G6PC, BBS9, ARGHEF9, TAF1*

**2. Repeat expansion**  
e.g. *FMR1, DMPK, ATXN10, HTT*

**3. Phasing**  
e.g. *Compound heterozygosity, Parental origin of de novo mutations, Mosaicism*

**4. Pseudogenes**  
e.g. *PMS2, CYP2D6, CHEK2, SMN1, PKD1*





# NGS (Long reads) In PGx

- No clinical adoption yet
- Few single gene studies
- Advantages: All apply to PGx implementation

RESEARCH ARTICLE

Human Mutation  
OFFICIAL JOURNAL  
**HGV**  
HUMAN GENOME VARIATION SOCIETY  
www.hgvs.org

**Flexible and Scalable Full-Length CYP2D6 Long Amplicon PacBio Sequencing**

Henk P.J. Buermans,<sup>1\*</sup> Rolf H.A.M. Vossen,<sup>1</sup> Seyed Yahya Anvar,<sup>1</sup> William G. Allard,<sup>1</sup> Henk-Jan Guchelaar,<sup>2</sup> Stefan J. White,<sup>1</sup> Johan T. den Dunnen,<sup>1,3</sup> Jesse J. Swen,<sup>2</sup> and Tahar van der Straaten<sup>2</sup>

HHS Public Access  
Author manuscript  
*Hum Mutat.* Author manuscript; available in PMC 2017 March 01.

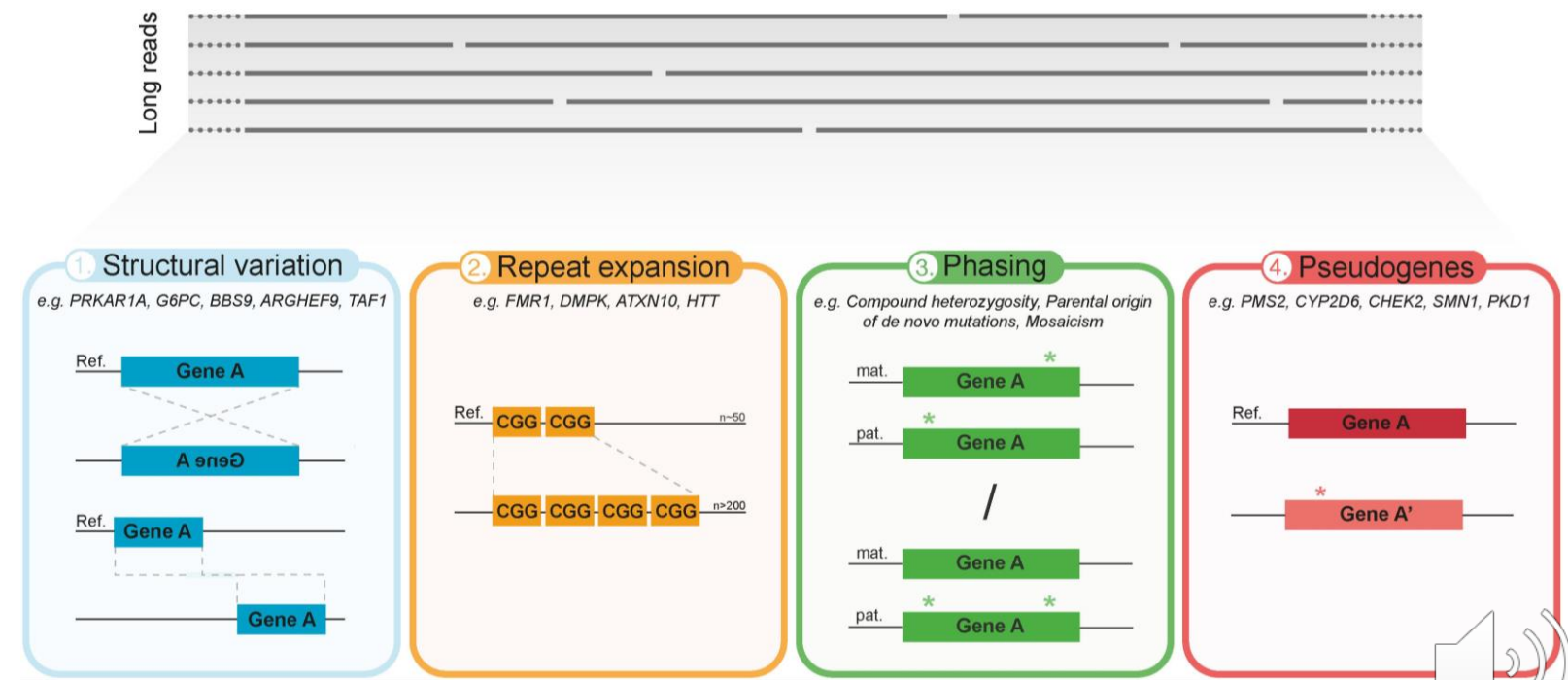
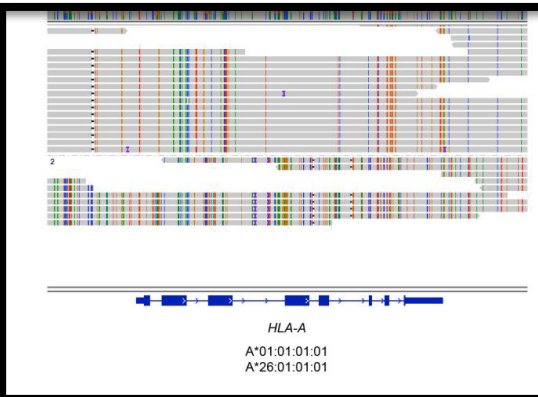
Published in final edited form as:  
*Hum Mutat.* 2016 March ; 37(3): 315–323. doi:10.1002/humu.22936.

**Long-read single-molecule real-time (SMRT) full gene sequencing of cytochrome P450-2D6 (CYP2D6)**

Wanqiong Qiao<sup>1\*</sup>, Yao Yang<sup>1\*</sup>, Robert Sebra<sup>1,2</sup>, Geetu Mendiratta<sup>1</sup>, Andrea Gaedigk<sup>3,4</sup>, Robert J. Desnick<sup>1</sup>, and Stuart A. Scott<sup>1</sup>

nature biotechnology ARTICLES  
<https://doi.org/10.1038/s41587-019-0217-9>

Accurate circular consensus long-read sequencing improves variant detection and assembly of a human genome



# PGx Application Challenges

Metabolizer  
Phenotype  
Inference

Haplotype  
Phasing

Structural  
Variants

Variants of  
Unknown Effect

Difficult genes

Limitations in  
clinical  
implementation



# PGx Application Challenges

Metabolizer  
Phenotype  
Inference

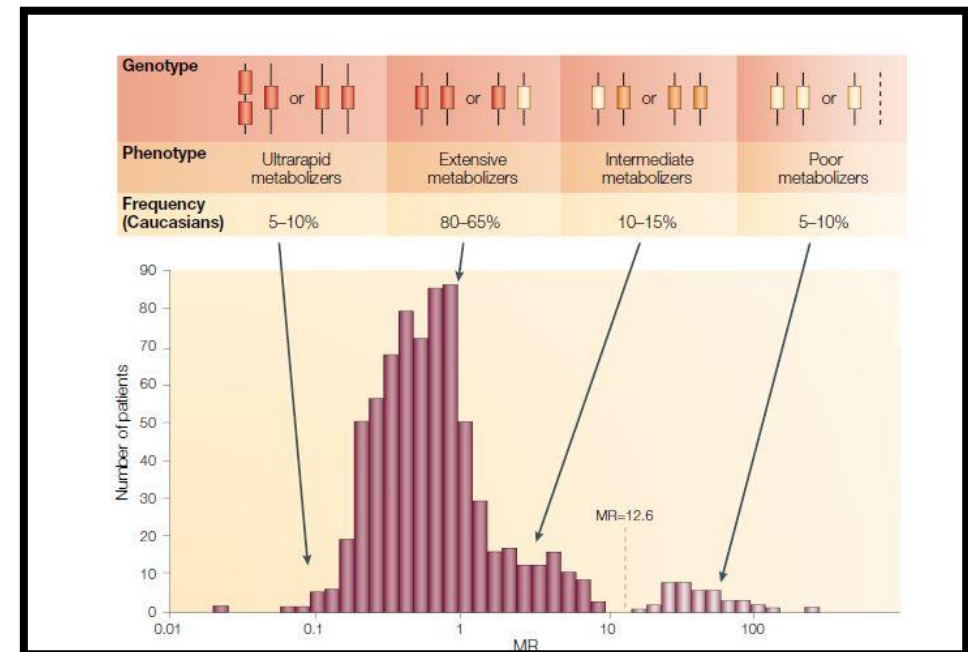


# Drug Metabolizer Phenotype Inference

- Genotypes are obtained
- For some genes (CYP genes) those genotypes make up haplotypes referred to as star alleles (\*)
- The combination of the two \* alleles makes a diplotype and translated into drug metabolizer phenotype.



The Pharmacogene Variation (PharmVar) Consortium is a central repository for pharmacogene (PGx) variation that focuses on haplotype structure and allelic variation.



# Drug Metabolizer Phenotype Inference

↓ CYP2C19\*1 PV00598 [80161A>G](#) (I331V)

↓ CYP2C19\*2 PV00599 [12662A>G](#) (splice defect), [19154G>A](#) (splice defect), [80161A>G](#) (I331V)

CYP2D6\*2 has 30 sub-alleles!!!

|                |           |         |   |
|----------------|-----------|---------|---|
| ↓ CYP2D6*2     |           | PV00427 | <a href="#">2851C&gt;T</a> (R296C), <a href="#">4181G&gt;C</a> (S486T)  |
| ↓ CYP2D6*2.001 | CYP2D6*2A | PV00129 | <a href="#">-1584C&gt;G</a> , <a href="#">-1235A&gt;G</a> , <a href="#">-740C&gt;T</a> , <a href="#">-678G&gt;A</a> , <a href="#">214G&gt;C</a> , <a href="#">221C&gt;A</a> , <a href="#">223C&gt;G</a> , <a href="#">227T&gt;C</a> , <a href="#">232G&gt;C</a> , <a href="#">233A&gt;C</a> , <a href="#">245A&gt;G</a> , <a href="#">310G&gt;T</a> , <a href="#">745C&gt;G</a> , <a href="#">842T&gt;G</a> , <a href="#">1662G&gt;C</a> , <a href="#">2851C&gt;T</a> (R296C), <a href="#">3385A&gt;C</a> , <a href="#">3585G&gt;A</a> , <a href="#">3791C&gt;T</a> , <a href="#">4181G&gt;C</a> (S486T), <a href="#">4482G&gt;A</a>  |
| ↓ CYP2D6*2.002 | CYP2D6*2B | PV00150 | <a href="#">1038C&gt;T</a> , <a href="#">1662G&gt;C</a> , <a href="#">2851C&gt;T</a> (R296C), <a href="#">4181G&gt;C</a> (S486T)  |
| ↓ CYP2D6*2.003 | CYP2D6*2C | PV00149 | <a href="#">1662G&gt;C</a> , <a href="#">2471T&gt;C</a> , <a href="#">2851C&gt;T</a> (R296C), <a href="#">4181G&gt;C</a> (S486T)  |
| ↓ CYP2D6*2.004 | CYP2D6*2D | PV00152 | <a href="#">2851C&gt;T</a> (R296C), <a href="#">4181G&gt;C</a> (S486T)  |
| ↓ CYP2D6*2.005 | CYP2D6*2E | PV00836 | <a href="#">-1584C&gt;G</a> , <a href="#">-1235A&gt;G</a> , <a href="#">-984G&gt;A</a> , <a href="#">-740C&gt;T</a> , <a href="#">-678G&gt;A</a> , <a href="#">214G&gt;C</a> , <a href="#">221C&gt;A</a> , <a href="#">223C&gt;G</a> , <a href="#">227T&gt;C</a> , <a href="#">232G&gt;C</a> , <a href="#">233A&gt;C</a> , <a href="#">245A&gt;G</a> , <a href="#">310G&gt;T</a> , <a href="#">745C&gt;G</a> , <a href="#">842T&gt;G</a> , <a href="#">996C&gt;G</a> , <a href="#">1662G&gt;C</a> , <a href="#">2851C&gt;T</a> (R296C), <a href="#">3385A&gt;C</a> , <a href="#">3585G&gt;A</a> , <a href="#">3791C&gt;T</a> , <a href="#">4181G&gt;C</a> (S486T), <a href="#">4482G&gt;A</a> |





# Drug Metabolizer Ph

CYP3A4\*14

CYP3A4\*16

CYP2D6\*2 has 30 sub

|                |           |         |   |
|----------------|-----------|---------|---|
| ↓ CYP2D6*2     |           | PV00427 |   |
| ↓ CYP2D6*2.001 | CYP2D6*2A | PV0017  |   |
| ↓ CYP2D6*2.002 | CYP2D6*2B | PV00156 |   |
| ↓ CYP2D6*2.003 | CYP2D6*2C |         |   |
| ↓ CYP2D6*2.004 | CYP2D6*2D | PV00152 |   |
| ↓ CYP2D6*2.005 | CYP2D6*2E | PV00336 | -1584...SA>G...-740C>G...>A, 214G>C...C>A, 223C>G, 227T>C, 232G>C, 233A>C, 245A>G, 310G>T, 745C>G, 842T>G, 996C>G, 1662G>C, 128... (R296C), 3585G>A...C>T, 4181G>C (3486T), 4482G>A |

CYP2D6 has over a 100 \* alleles and over 2000 variants that define that



# Drug Metabolizer Phenotype Inference

- Several tools have been developed to assign \*-allele and haplotypes based on sequencing data
- Performance is quite variable, in a study Aldy showed the least errors, compared to Stargazer and Astrolabe (2 compared to 9 and 10 respectively out of 21 alleles tested)
- All tools use PharmVar as their database. PharmVar is updated continuously leading to potential differences in assignments if not every tool is updated at the same time.
- Most tools require training datasets, and variation in those training sets can result in different sensitivity.



Stargazer

PharmCAT

Aldy

Astrolabe

Cyprip

g-Nomic

PHARMIP

Cyrius



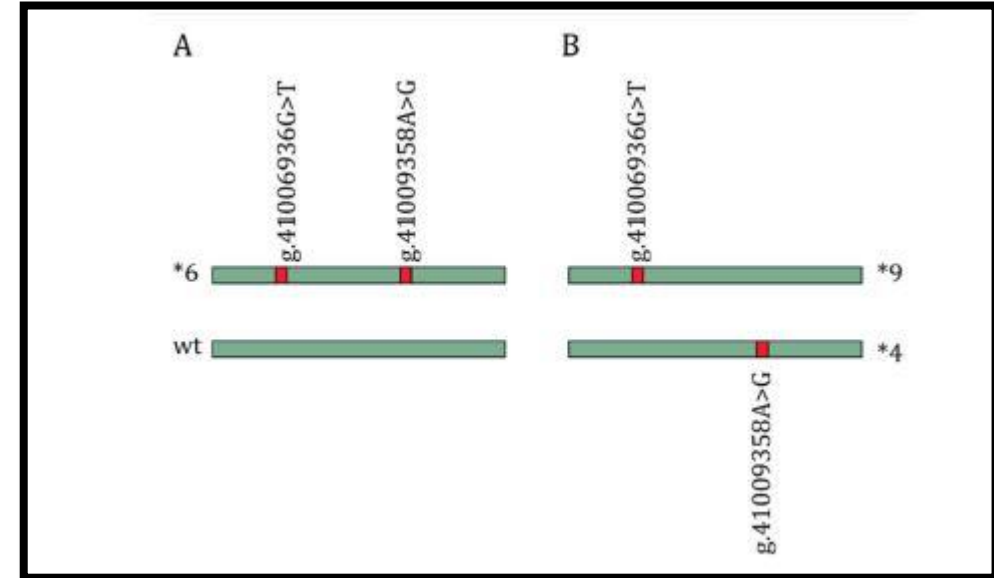
# PGx Application Challenges

## Haplotype Phasing



# Haplotype phasing

- Determining if variants are located on the same allele or if they are on different alleles, leading to differences in phenotype assignment
- Given the polymorphic nature of many pharmacogenes, the likelihood of identifying multiple heterozygous variants within the gene locus of interest is highly likely
- While CPIC and the DPWG report which diplotype translate into which phenotypes; but no guidance on phasing
- Having two variants on one allele is different than having them on opposing alleles.



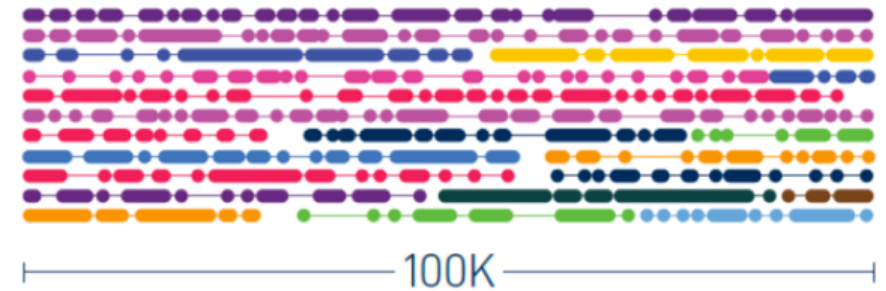
CYP2B6



# Haplotype phasing

- Resolving phasing by short-read NGS:
  - » Linked-read sequencing: use of barcoded short fragments sequenced with conventional short-read methods.
  - » Using barcodes, every read can be linked back to the original position and artificial long input DNA can be reconstructed
- Resolving phasing by Long-read NGS:
  - » The length of the reads can be utilized for haplotype phasing

Linked-Reads



PacBio HiFi and Hi-C





# PGx Application Challenges

## Structural Variants



# Structural Variants

- Majority of pharmacogenes Are largely characterized by complex regions
- CNVs, structural rearrangements and repetitive regions

| Protein        | Gene           | Related Drugs |      | Locus Size (bp) | Rare Variants, n (% of Known Variants) | Part of Locus Defined as Complex, %(bp) |
|----------------|----------------|---------------|------|-----------------|--|---|
|                |                | CPIC          | DPWG |                 |  |   |
| CACNA1S        | CACNA1S        | 7             | -    | 73,055          | 2520 (98%)                             | 33.3                                    |
| CFTR           | CFTR           | 1             | -    | 250,187         | 1684 (99%)                             | 42.2                                    |
| CYP2B6         | CYP2B6         | 1             | 1    | 27,149          | 761 (98%)                              | 100.0                                   |
| CYP2C9         | CYP2C9         | 10            | 2    | 50,734          | 632 (98%)                              | 72.0                                    |
| CYP2C19        | CYP2C19        | 15            | 10   | 90,525          | 712 (99%)                              | 83.6                                    |
| CYP2D6         | CYP2D6         | 14            | 21   | 4408            | 992 (97%)                              | 100.0                                   |
| CYP3A5         | CYP3A5         | 1             | 1    | 31,833          | 643 (98%)                              | 49.4                                    |
| CYP4F2         | CYP4F2         | 1             | -    | 20,098          | 766 (97%)                              | 51.4                                    |
| DPD            | DPYD           | 2             | 4    | 917,258         | 1211 (98%)                             | 40.0                                    |
| FACT. V LEIDEN | FACT. V LEIDEN | -             | 1 †  | 72,423          | 1679 (97%)                             | 41.9                                    |
| G6PD           | G6PD           | 1             | -    | 16,183          | 465 (98%)                              | 36.4                                    |
| HLA-A          | HLA-A          | 2             | 1    | 4625            | 423 (71%)                              | 100.0                                   |
| HLA-B          | HLA-B          | 6             | 7    | 87,698          | 308 (78%)                              | 62.1                                    |
| IFNL3          | IFNL3          | 2             | -    | 1577            | 317 (95%)                              | 100.0                                   |
| IFNL4          | IFNL4          | 2             | -    | 3543            | 404 (97%)                              | 100.0                                   |
| NUDT15         | NUDT15         | 3             | 3    | 9656            | 244 (99%)                              | 64.7                                    |
| RYR-1          | RYR1           | 7             | -    | 153,866         | 6584 (98%)                             | 51.4                                    |
| SLCO1B1        | SLCO1B1        | 1             | 2    | 108,045         | 951 (96%)                              | 69.6                                    |
| TPMT           | TPMT           | 3             | 3    | 26,764          | 346 (97%)                              | 52.3                                    |
| UGT1A1         | UGT1A1         | 1             | 1    | 13,052          | 470 (99%)                              | 40.3                                    |
| VKORC1         | VKORC1         | 1             | 3    | 5139            | 370 (98%)                              | 41.8                                    |



# Structural Variants by NGS

- Multiple tools designed to extract CNVs:  
XHMM , CoNIFER, Varseq, CNVnator
- Agreement between methods is low
- There is bias towards smaller CNVs vs. large CNVs
- Distinction between a pharmacogene and a pseudogene can be challenging (e.g. CYP2D6 and CYP2D7 share >98% of their sequence)
- Long-read sequencing can distinguish gene from pseudogene, can better assess large insertions and deletions, and structural variants
- Full characterization of the complexity of pharmacogenes is still in the research phase

| Protein | Gene    | Related Drugs |      | Locus Size (bp) | Rare Variants, n (% of Known Variants) | Part of Locus Defined as Complex, %(bp) |
|---------|---------|---------------|------|-----------------|--|---|
|         |         | CPIC          | DPWG |                 |  |   |
| CACNA1S | CACNA1S | 7             | -    | 73,055          | 2520 (98%)                             | 33.3                                    |
| CFTR    | CFTR    | 1             | -    | 250,187         | 1684 (99%)                             | 42.2                                    |
| CYP2B6  | CYP2B6  | 1             | 1    | 27,149          | 761 (98%)                              | 100.0                                   |
| CYP2C9  | CYP2C9  | 10            | 2    | 50,734          | 632 (98%)                              | 72.0                                    |
| CYP2C19 | CYP2C19 | 15            | 10   | 90,525          | 712 (99%)                              | 83.6                                    |
| CYP2D6  | CYP2D6  | 14            | 21   | 4408            | 992 (97%)                              | 100.0                                   |
| CYP3A5  | CYP3A5  | 1             | 1    | 31,833          | 643 (98%)                              | 49.4                                    |
| CYP4F2  | CYP4F2  | 1             | -    | 20,098          | 766 (97%)                              | 51.4                                    |
| DPD     | DPYD    | 2             | 4    | 917,258         | 1211 (98%)                             | 40.0                                    |
| FACT.V  | FACT.V  | -             | 1 †  | 72,423          | 1679 (97%)                             | 41.9                                    |
| LEIDEN  | LEIDEN  | -             | -    | -               | -                                      | -                                       |
| G6PD    | G6PD    | 1             | -    | 16,183          | 465 (98%)                              | 36.4                                    |
| HLA-A   | HLA-A   | 2             | 1    | 4625            | 423 (71%)                              | 100.0                                   |
| HLA-B   | HLA-B   | 6             | 7    | 87,698          | 308 (78%)                              | 62.1                                    |
| IFNL3   | IFNL3   | 2             | -    | 1577            | 317 (95%)                              | 100.0                                   |
| IFNL4   | IFNL4   | 2             | -    | 3543            | 404 (97%)                              | 100.0                                   |
| NUDT15  | NUDT15  | 3             | 3    | 9656            | 244 (99%)                              | 64.7                                    |
| RYR-1   | RYR1    | 7             | -    | 153,866         | 6584 (98%)                             | 51.4                                    |
| SLCO1B1 | SLCO1B1 | 1             | 2    | 108,045         | 951 (96%)                              | 69.6                                    |
| TPMT    | TPMT    | 3             | 3    | 26,764          | 346 (97%)                              | 52.3                                    |
| UGT1A1  | UGT1A1  | 1             | 1    | 13,052          | 470 (99%)                              | 40.3                                    |
| VKORC1  | VKORC1  | 1             | 3    | 5139            | 370 (98%)                              | 41.8                                    |



# PGx Application Challenges

Variants of  
Unknown Effect



# Variants of unknown effect

- The clinical utility is limited and not much is known about their function
- Solutions could be:
  - Cell-line models
  - In silico predictions (sequence conservation, the physiochemical and crystal structure of the protein, or on evolutionary scores)
  - Studying patients displaying the most extreme phenotypes





# PGx Application Challenges

Difficult genes



# Difficult genes

| Gene          | Challenges                                   |
|---------------|--|
| <i>CYP2D6</i> | Structural variants and gene re-arrangements |
|               | Pseudogenes                                  |
|               | CNVs   |
|               | Highly polymorphic                           |
| <i>UGTA1A</i> | Rare population specific variants            |
|               | Non-coding variants                          |
| <i>VKORC1</i> | Non-coding variants                          |
| <i>HLA</i>    | Highly polymorphic regions                   |
|               | Rare population specific variants            |



# PGx Application Challenges

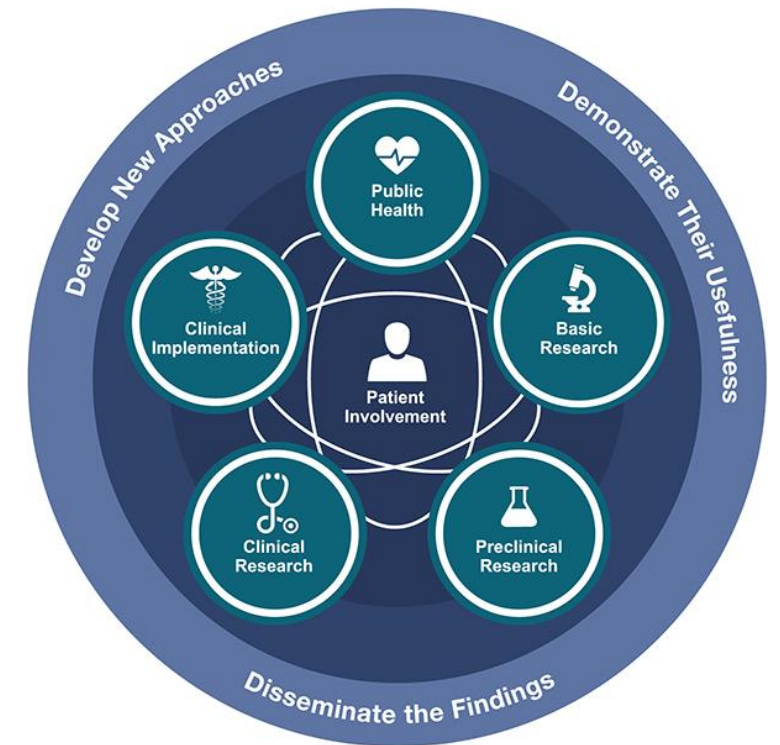
Clinical  
implementation



# Challenges to clinical application

## Translating basic science findings into clinical practice:

- \*Clinical Studies/Trials
- \*Clinical practice recommendations and guidelines
- \*Adoption of guidelines into evidence-based practice
- \*Assessing efficacy, cost, outcomes.etc.

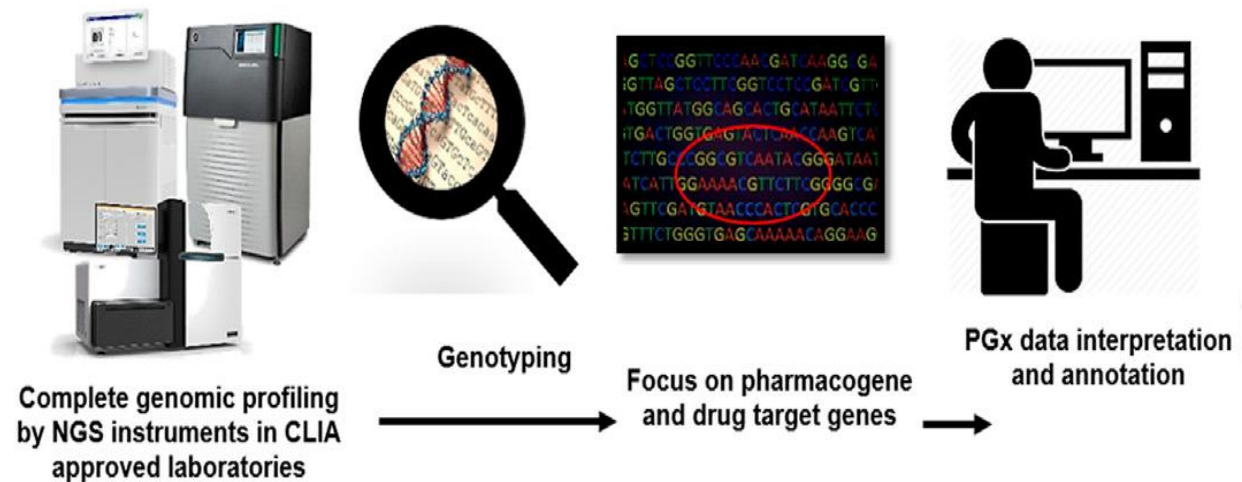


Credit: National Center for Advancing Translational Sciences



# Challenges to clinical application

- Need results fast...point-of-care, pre-emptive testing  
New molecular testing assays can be performed in ~1 hour
- Need for infrastructure:  
High throughput sequencers and molecular Sequencing technologies  
Specific software and computational tools

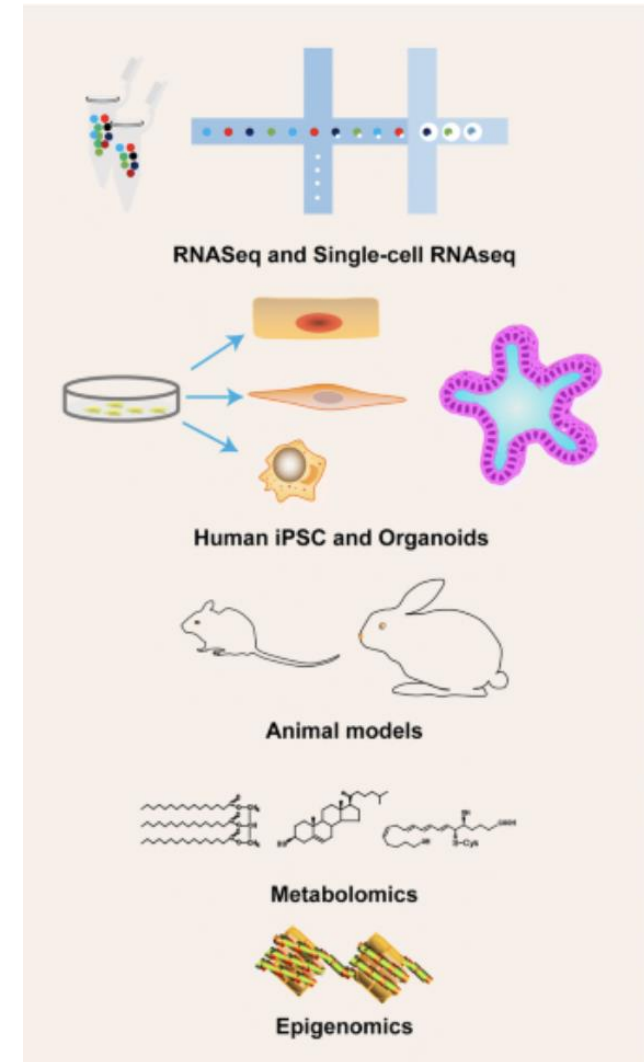




# Challenges to clinical application

## Functional characterization of variants:

- \*In silico tools
- \*In Vitro studies
- \*In Vivo models
- \*Non-coding elements and regulatory regions



# Challenges to clinical application

- Physicians training  
Translating genotypes into phenotypes and using guidelines to guide managements is not common knowledge
- Laboratory consultations  
Laboratories offering tests should offer consultations
- PGx teams/ clinics  
Trained to use clinical support systems



**PGx trained clinicians**

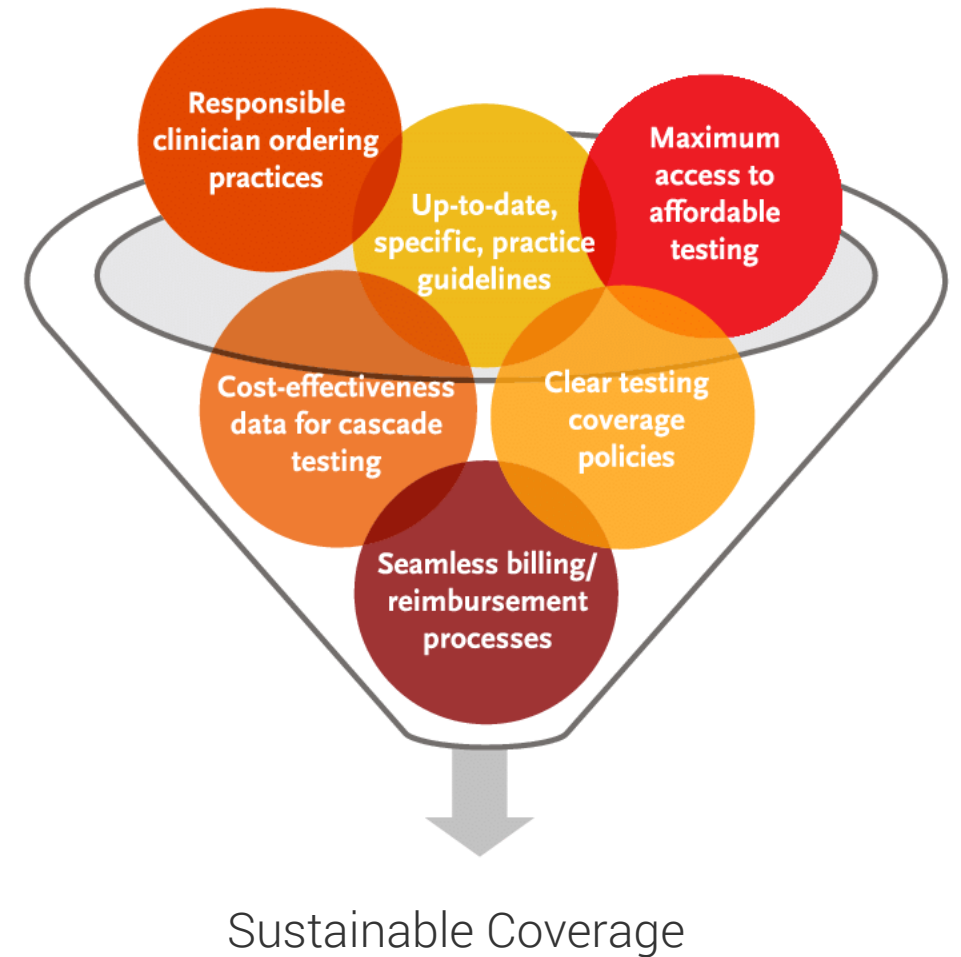


# Challenges to clinical application

## Research funding and Reimbursement

\*While still low, reimbursement for PGx testing almost doubled in the last few years.

\*Allies within legislative bodies needed to ensure adequate funding for both research as well as for efforts to build precision medicine and PGx programs



# PHARMACOGENOMIC RESOURCES

1. <http://www.cypalleles.ki.se/>
  - Catalog of *CYP450* genetic variation and nomenclature.
2. <http://www.pharmGKB.org/>
  - A central PGx resource: collect, encode, and disseminate knowledge about the impact of human genetic variations on drug response.
3. <http://www.cpicpgx.org>
  - **CPIC**: continued guidelines on PGx-based clinical management.
4. <http://www.warfarindosing.org/>
  - Online tool to predict warfarin dose using both clinical and genetic variables.
5. <http://medicine.iupui.edu/clinpharm/ddis/>
  - *CYP450* drug interaction database.
6. <http://www.fda.gov/Drugs/ScienceResearch/ResearchAreas/Pharmacogenetics/ucm083378.htm>
  - FDA pharmacogenetics biomarker table.





*A nonprofit enterprise of the University of Utah and its Department of Pathology*